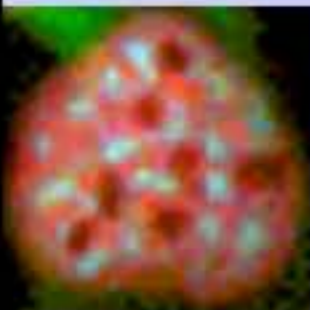


A visual scripting interface for ImageJ and
its usage at the microscopy facility
Montpellier RIO Imaging

Volker Baecker and Pierre Travo

19.05.2006

Montpellier RIO Imaging



Overview

A visual scripting interface for ImageJ

- introduction and motivation
 - using visual scripting
 - simple applications
 - batch applications
 - interactive applications
 - extending the framework
 - adding operations
 - accessing ImageJ methods
 - the implementation
- ... and its usage at the microscopy facility MRI
- DNA combing application
 - measuring intensity ratios
 - measuring fluorescent stained proteins
 - counting cells and nuclei
 - automation of simple tasks
 - further project
 - conclusions





Who am I

- Born: 29.08.1970
- Nationality: German (living in France since Nov. 2003)
- 2000
Master thesis in computer science at the University of Dortmund:
“A development environment for cellular automata with fuzzy rules”
- 2000 – 2003
Software developer and consultant for object oriented systems
George Heeg eK
- Nov. 2003 – Feb. 2004
Centre de Recherches de Biochimie Macromoléculaire,
Structural Bioinformatics and Molecular Modelling, Andrey Kajava
- From Feb. 2004
Software development at the Microscopy Facility: Montpellier RIO Imaging



Montpellier RIO Imaging

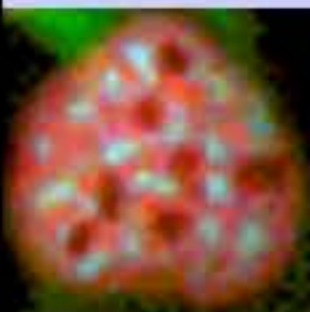
Responsible



PierreTravo, IRCE CNRS

- regional imaging facility
- 4 sites
- 23 microscopes
- 420 users
- 25413 hours in 2005
- 3M images in 2005
- promote the usage of microscopy
- participate in the development of microscopy
- provide training

MRI-EDU					
	Composition variable Toutes opérations de formation et communication Resp. J. Cau	MRI-DEV			
	Corine Tran-Aupiais IE2 CNRS Optronique		Olivier Miquel T3 CNRS Systèmes Réseaux	Sylvain de Rossi IE2 INSERM Optique et systèmes LASER	Volker Bäcker, IE CDD Conception développement
		Optique adaptative	Soutien à la plate-forme	Soutien à la plate-forme	MRI Cell Image Analyzer



Responsible

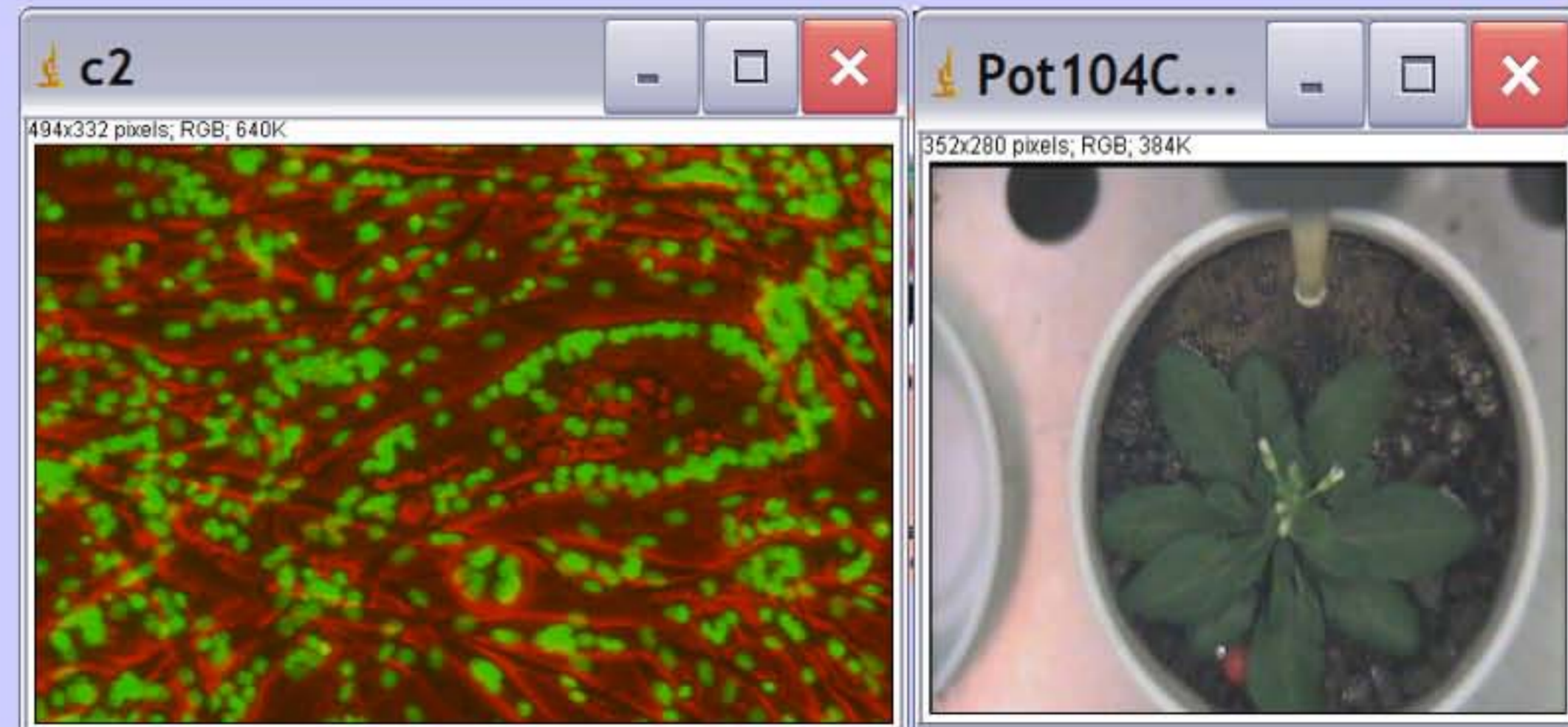


PierreTravo, IRCE CNRS

Montpellier RIO Imaging

- regional imaging facility
- 4 sites
- 23 microscopes
- 420 users
- 25413 hours in 2005
- 3M images in 2005
- promote the usage of microscopy
- participate in the development of microscopy
- provide training

wide range of mostly
biological applications
images from sub-cellular
to entire organisms



The problem

robotized acquisition,
time series, volume images

- large amounts of data

manual analysis



- time consuming
- biased results?

analysis is the
bottleneck

→ automatic analysis
needed

wide range of

- analysis needs
- image qualities

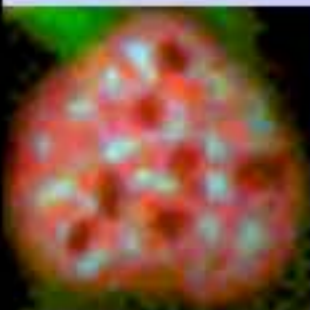


→ analysis must use a priori knowledge

if automatic analysis not possible

- partial automation can augment efficiency
- let the user only do what the software can't do

→ semi-automatic analysis needed



Problem / Requirements



wanted:

rapid prototyping framework for
image analysis applications

requirements:

- interactive experimentation to find solutions
- create interactive and batch applications
- build prototype-applications from existing operations rapidly
- extendable - add new operations
- allow to parametrize and run applications
- easy to use for end user





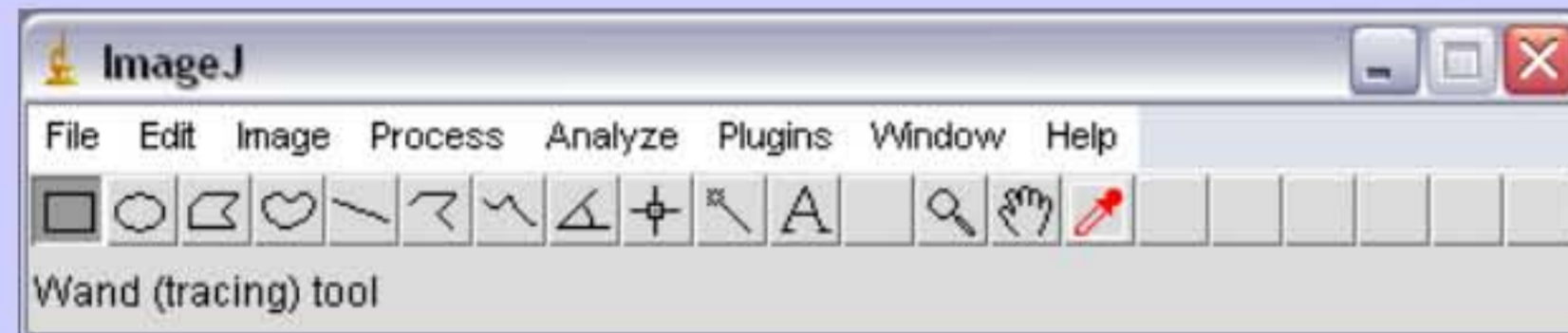
Process

process:

- use framework
 - to create prototype solutions on demand
 - in close collaboration with biologist
 - extend framework, when necessary for a project
 - **only then**
- eventually create full featured application



ImageJ



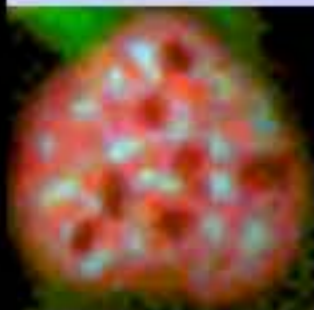
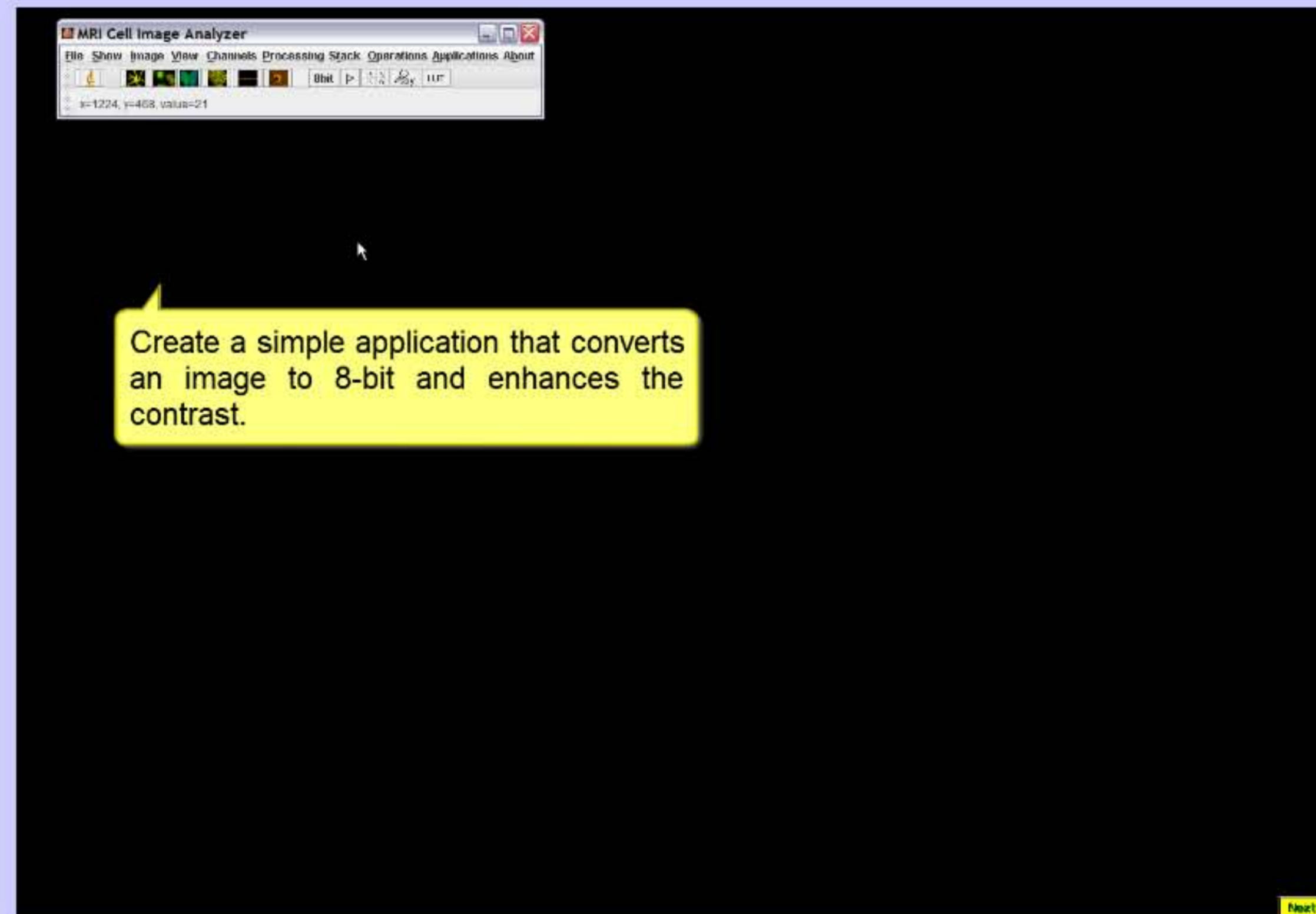
Is ImageJ the desired framework?

- interactive experimentation
 - solid image processing / analysis framework
- create interactive and batch applications
- allow to parametrize and run applications
 - macros
- extendable, configurable
 - plug-ins, source code available, public domain
- build prototype-applications from existing operations rapidly
 - yes, but macro language not understandable for end user
- easy to use
 - yes, but users have to learn command shortcuts or search in menus
- great for image analysis specialist
- some aspects too technical for end user



Using visual scripting

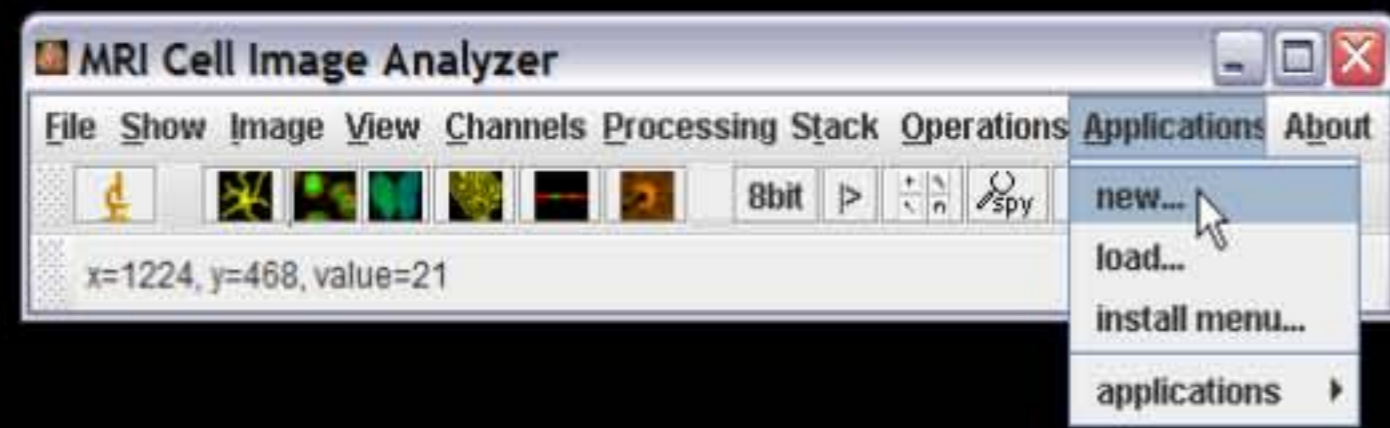
- create a simple application





Create a simple application that converts an image to 8-bit and enhances the contrast.





Menu Applications->new...

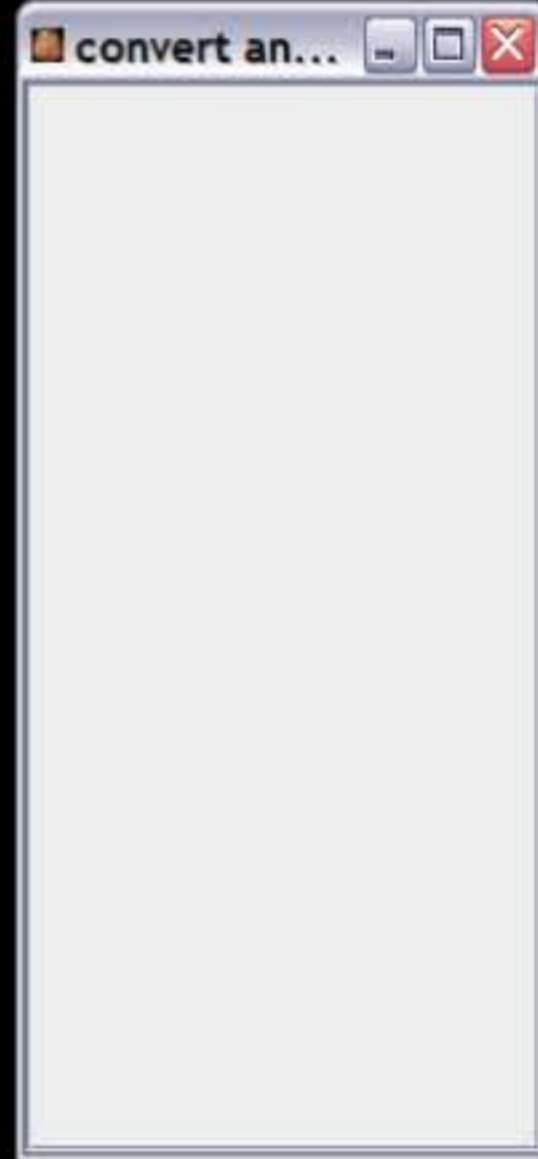


Enter the name of the new application







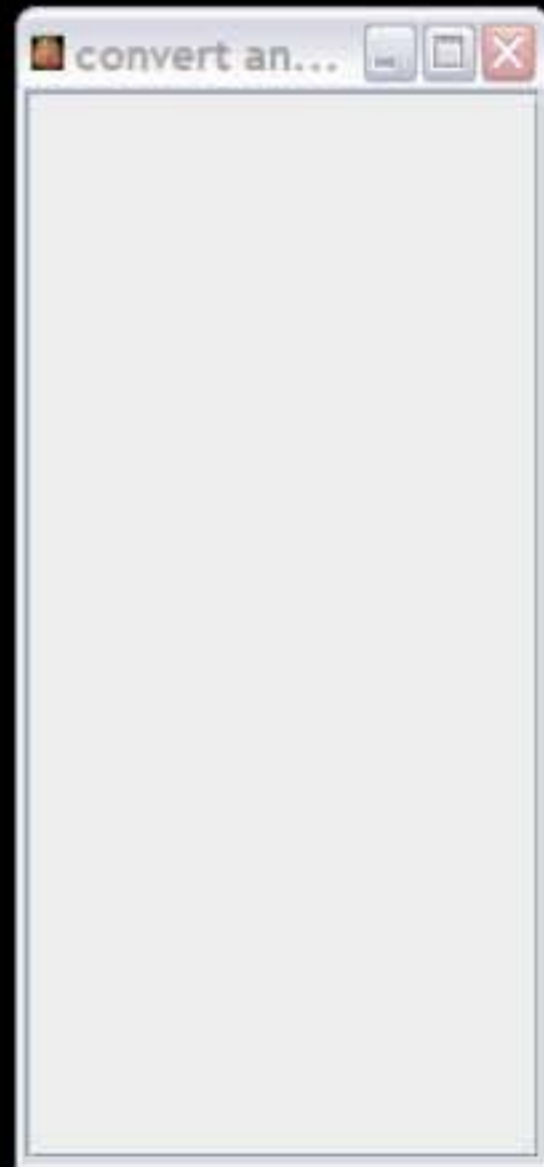
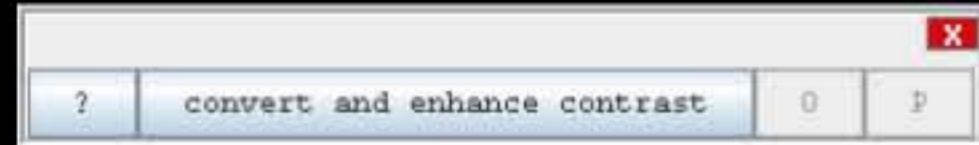


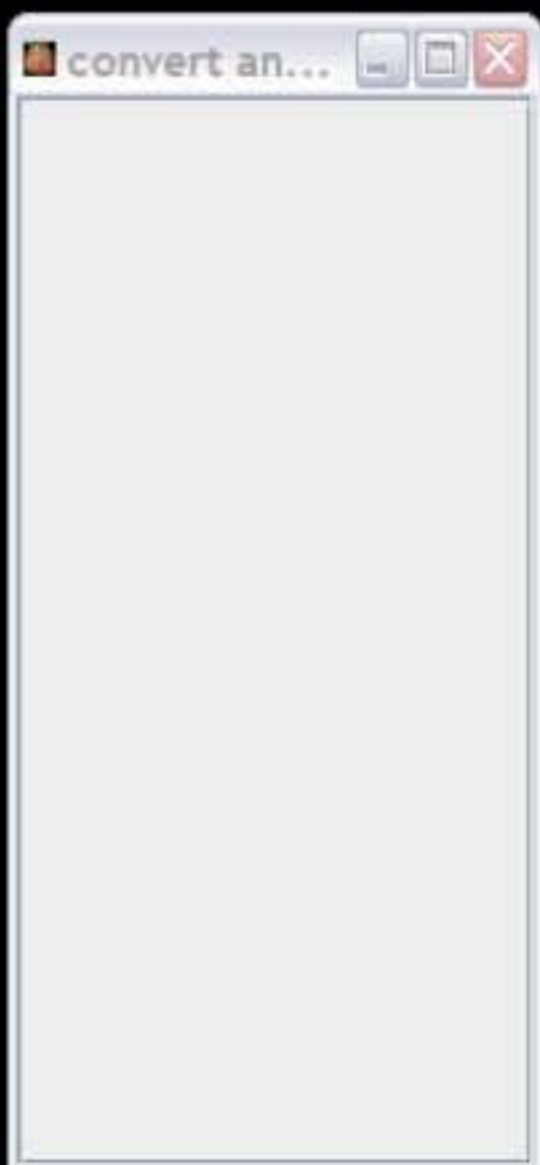
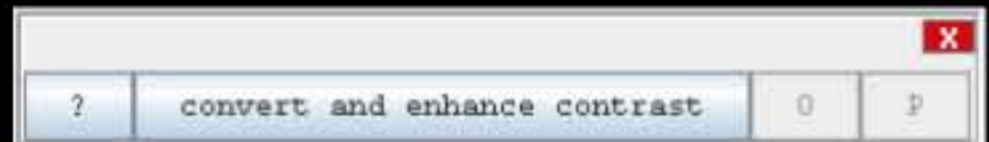
tile representation with buttons

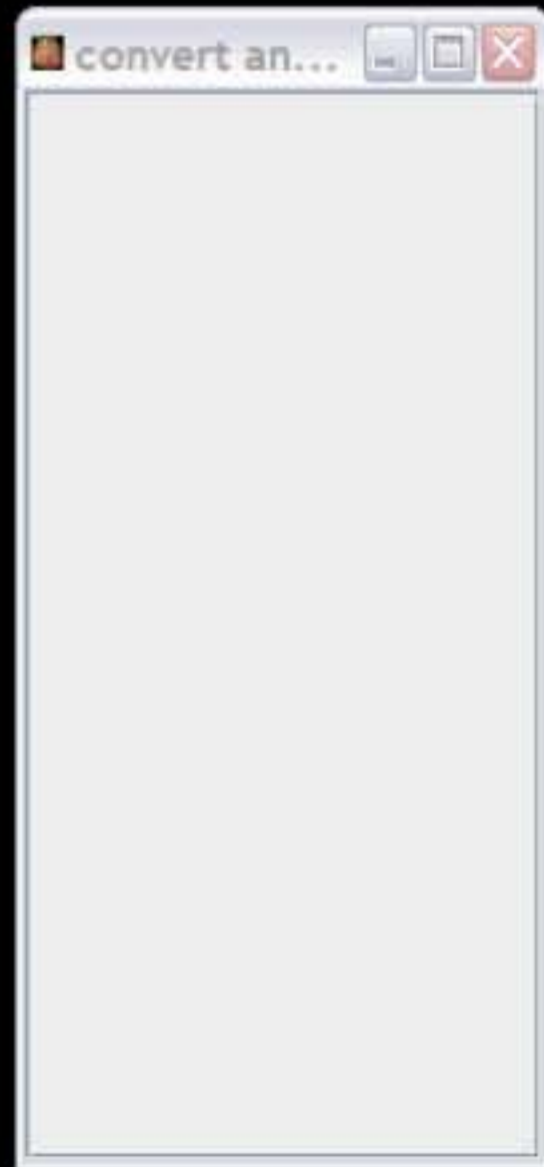
- help
- start application

box representation

- list of operations (empty)







to add operations, open

- collection of operations

or

- list of all operations

MRI Cell Image Analyzer

File Show Image View Channels Processing Stack Operations Applications About

8bit ▶ LUT

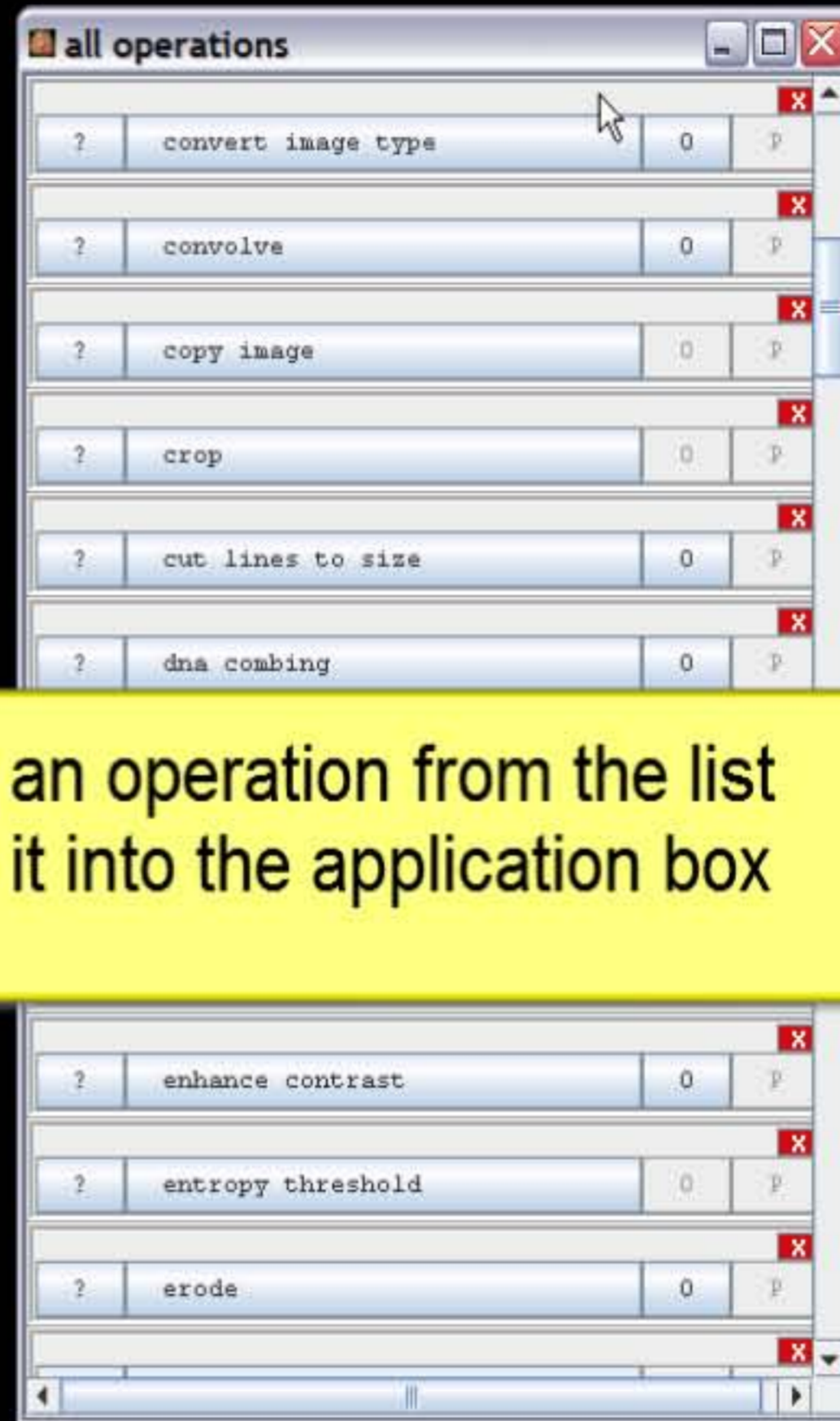
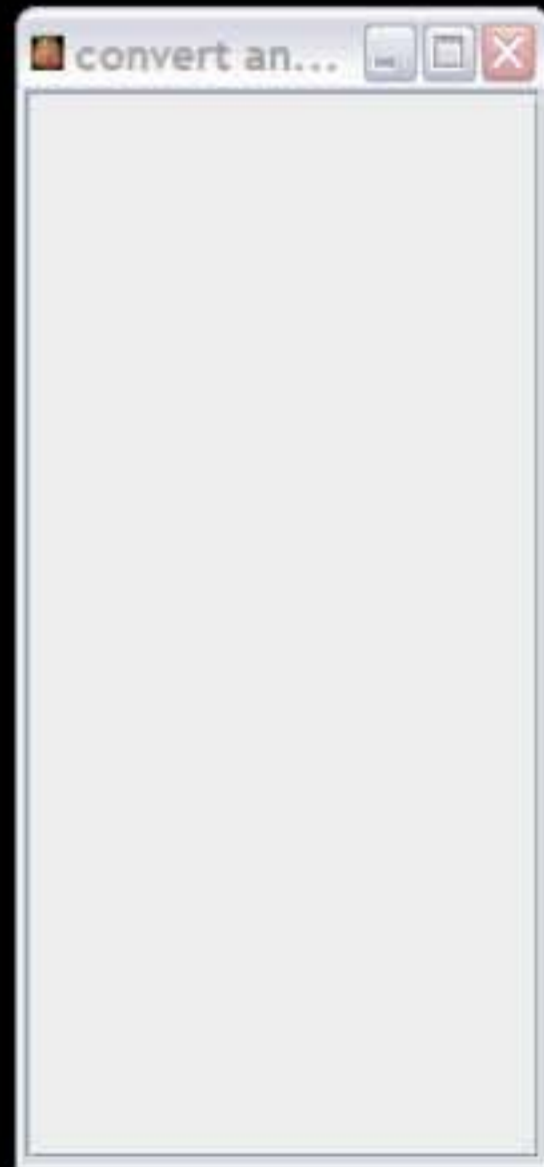
x=1224, y=468, value=21

convert and enhance contrast 0 P

convert an...

all operations

? accept or skip or exit	0	P
? adaptive erode	0	P
? apply lut	0	P
? auto threshold	0	P
? binary fill	0	P
? clear image	0	P
? clear	0	P
? close image	0	P
? close session	0	P
? combine images	0	P
? compute moments	0	P
? convert image type	0	P



- drag an operation from the list
- drop it into the application box

MRI Cell Image Analyzer

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x=1224, y=468, value=21

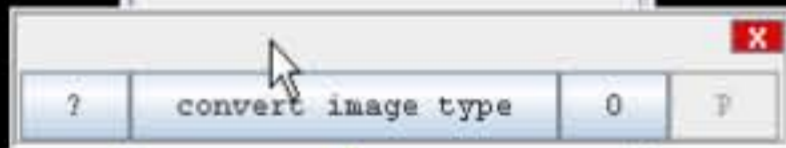
? convert and enhance contrast 0 P

convert an...

all operations

? convert image type			
? convolve	0	P	
? copy image	0	P	
? crop	0	P	
? cut lines to size	0	P	
? dna combing	0	P	
? dilate	0	P	
? draw	0	P	
? duplicate slice	0	P	
? enhance contrast	0	P	
? entropy threshold	0	P	
? erode	0	P	

? convert image type 0 P



tile representation of operation

- help page
- start operation
- open options dialog
- open parameter dialog



MRI Cell Image Analyzer

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x=1224, y=468, value=21

?

convert and enhance contrast

0

P

convert and enhance cont...

?

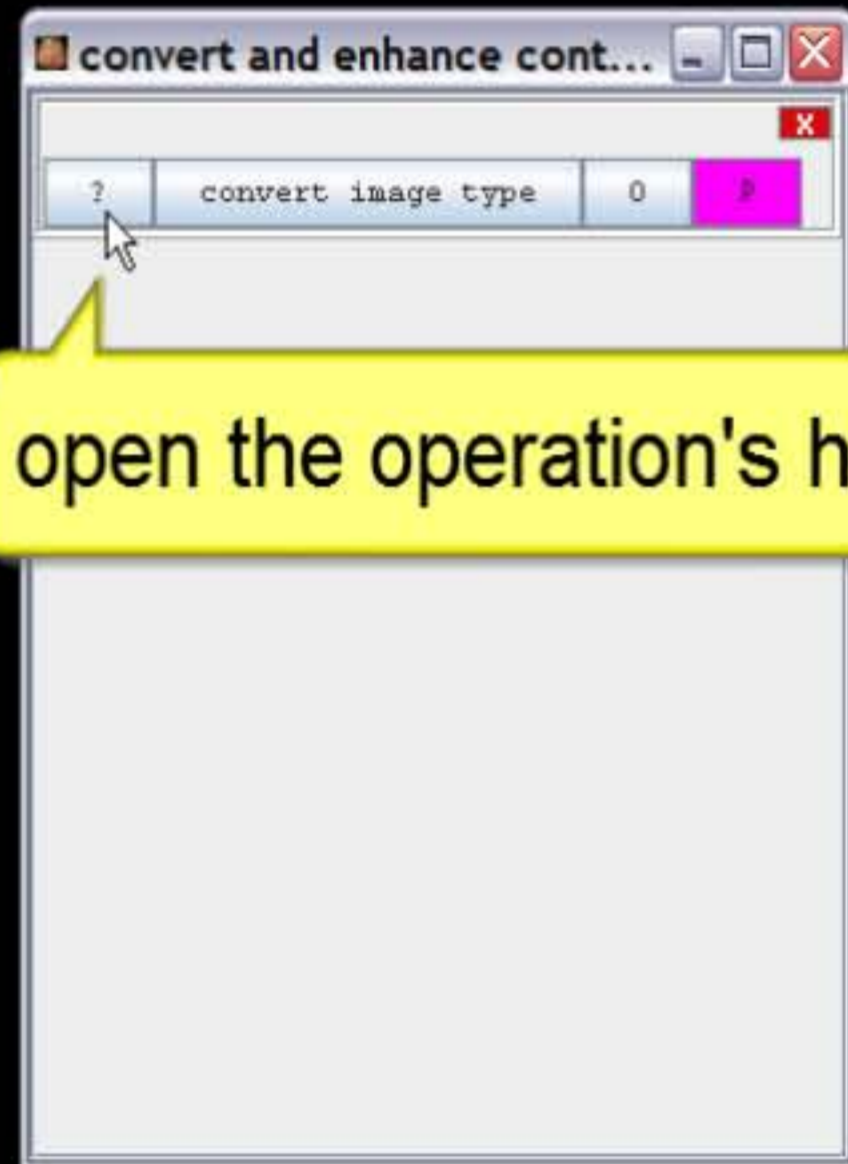
convert image type

0

P

all operations

?	convert image type	0	P
?	convolve	0	P
?	copy image	0	P
?	crop	0	P
?	cut lines to size	0	P
?	dna combing	0	P
?	dilate	0	P
?	draw	0	P
?	duplicate slice	0	P
?	enhance contrast	0	P
?	entropy threshold	0	P
?	erode	0	P



open the operation's help page

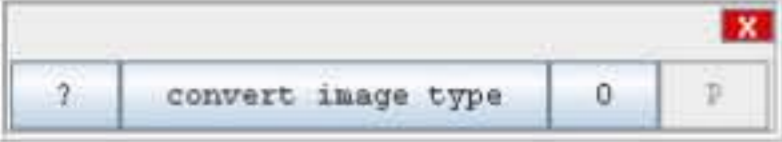


MRI Cell Image Analyzer Help - Operation: convert image type - Mozilla Firefox

Fichier Edition Affichage Aller à Marque-pages Outils ?

file:///C:/Programme/MRI%20cell%20image%20analyzer/_help/c

Operation: convert image type



Author:

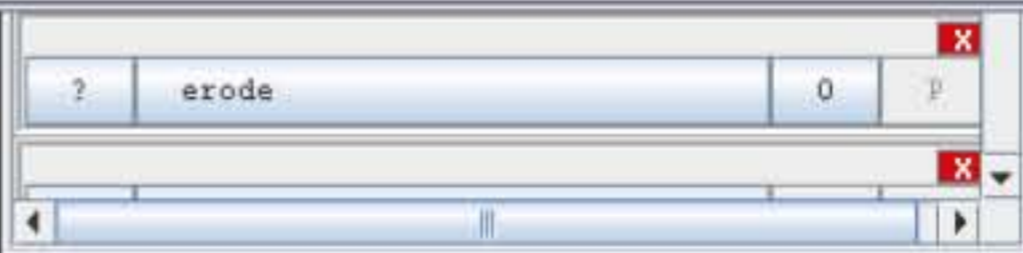
The operation uses the Class [Converter](#) from ImageJ.

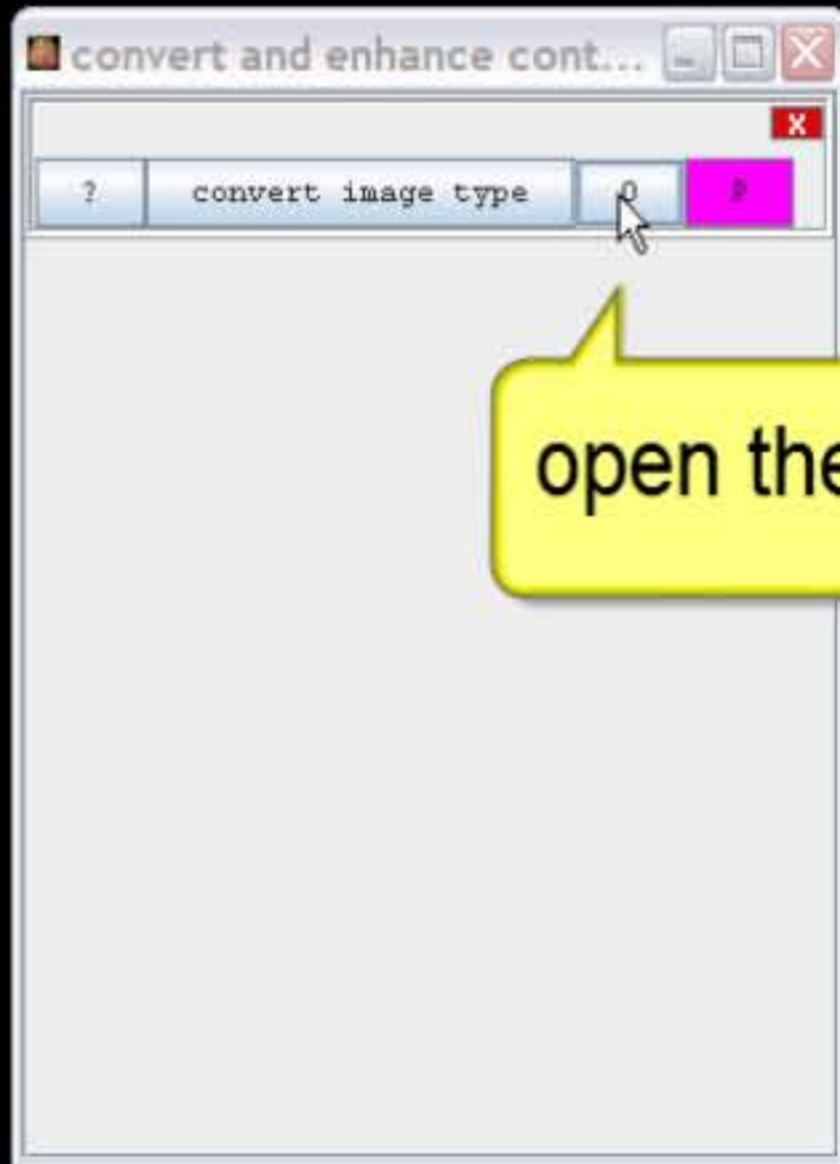
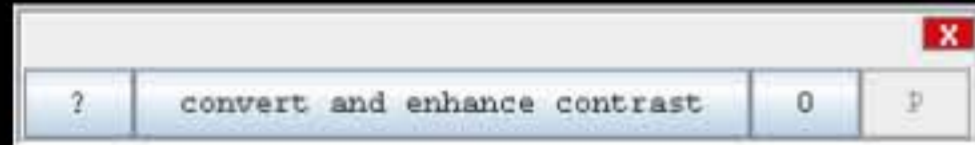
Description

Converts the image type of the input image to the type selected in the operation's options. Depending on the type of the input image not all conversions are possible. See [ImageJ documentation, menu "image"](#) for details.

Supported Conversions:

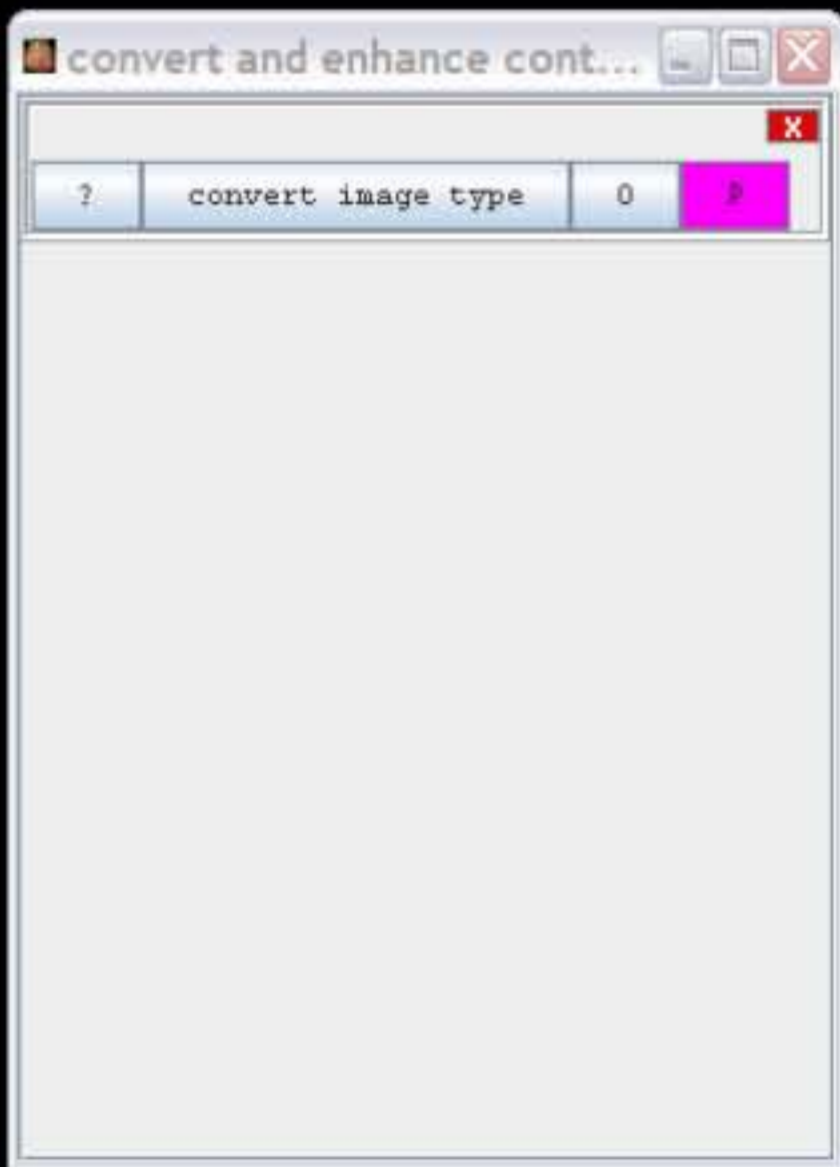
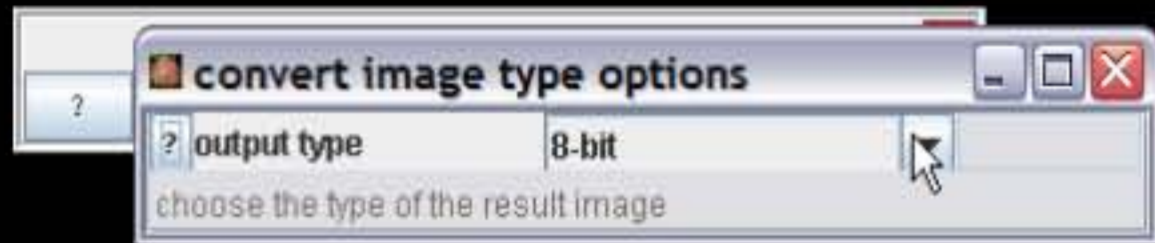
Terminé

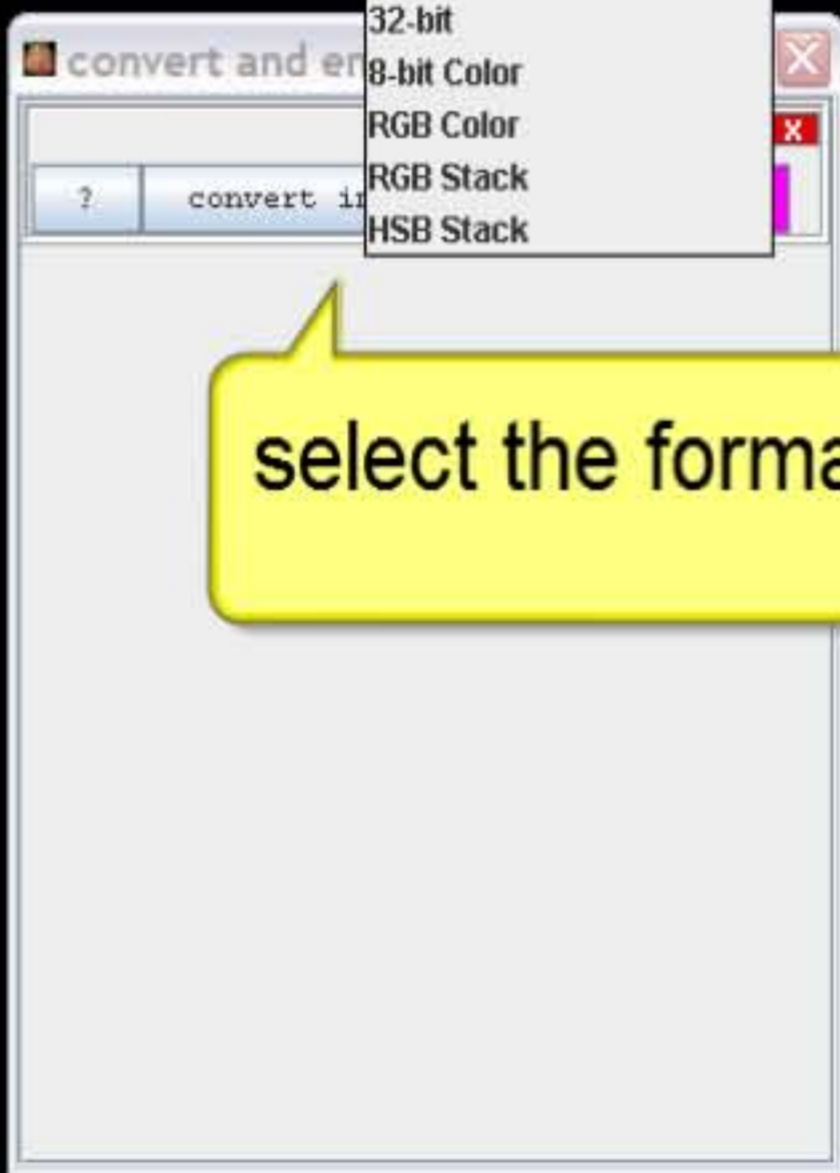
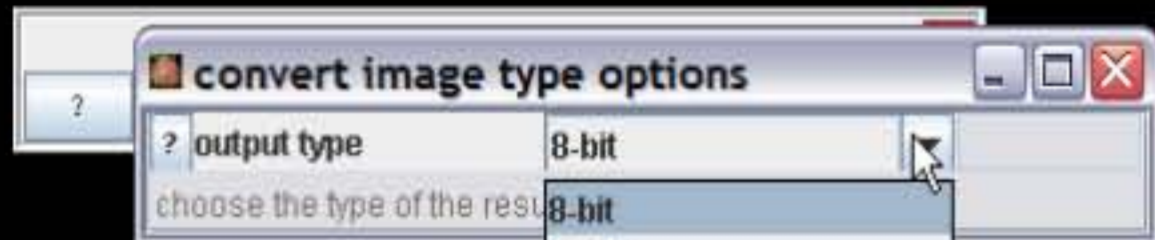




open the options dialog

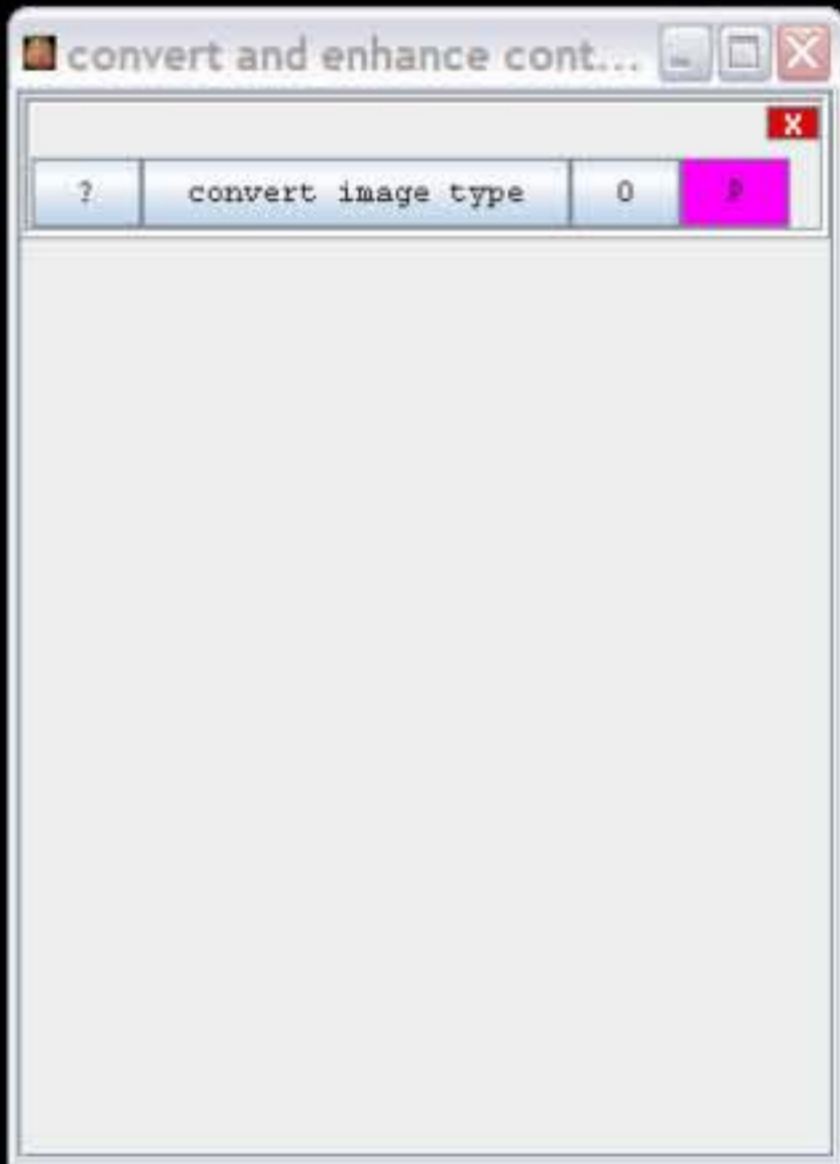
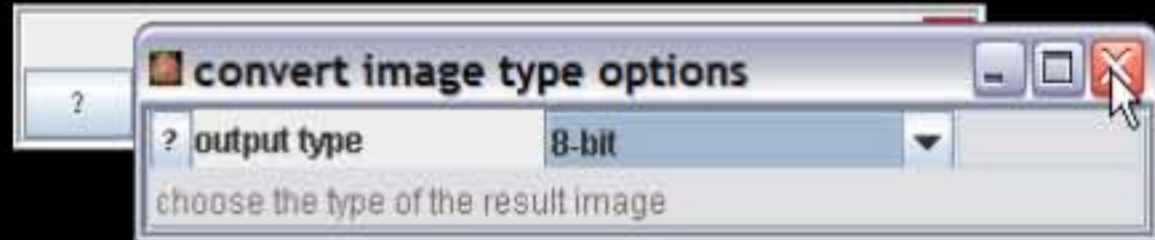


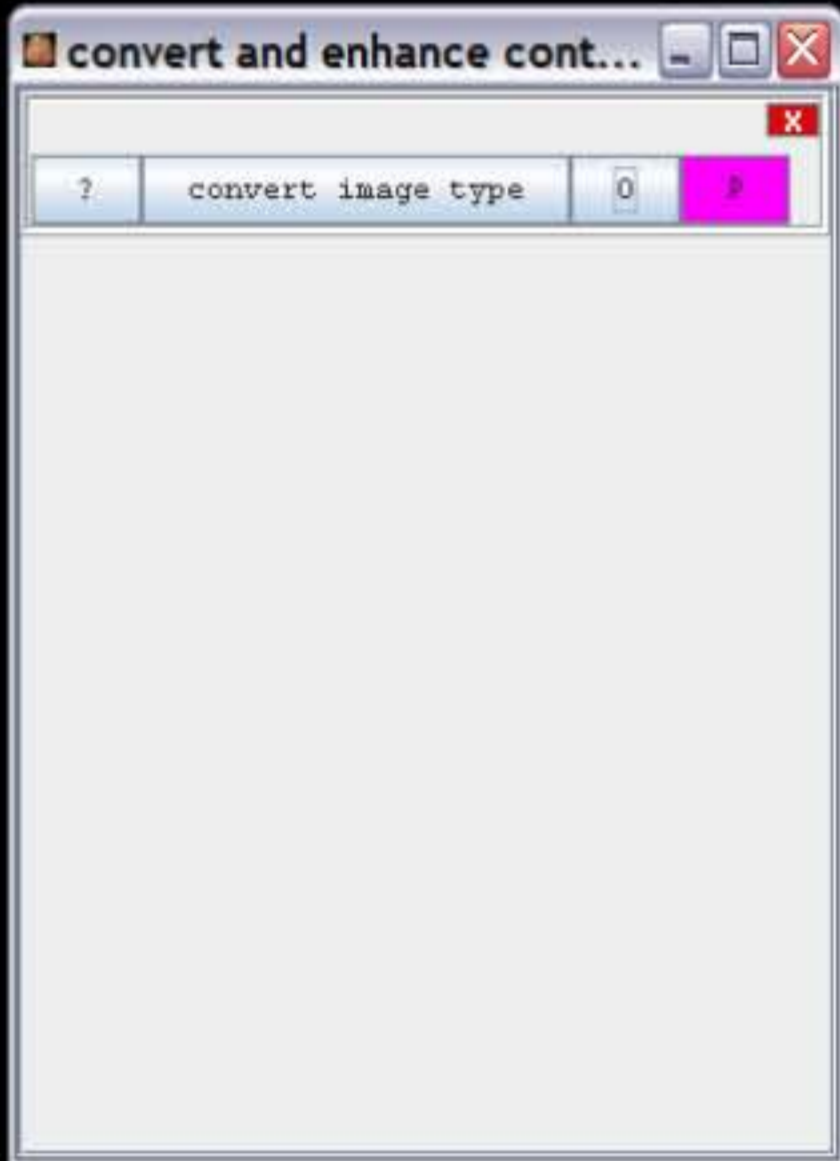
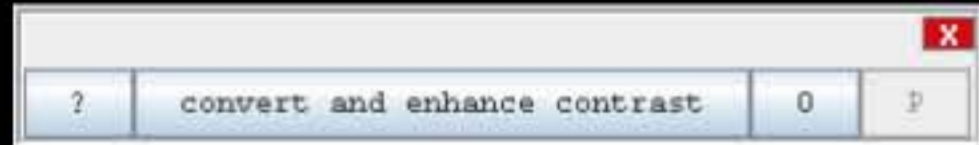
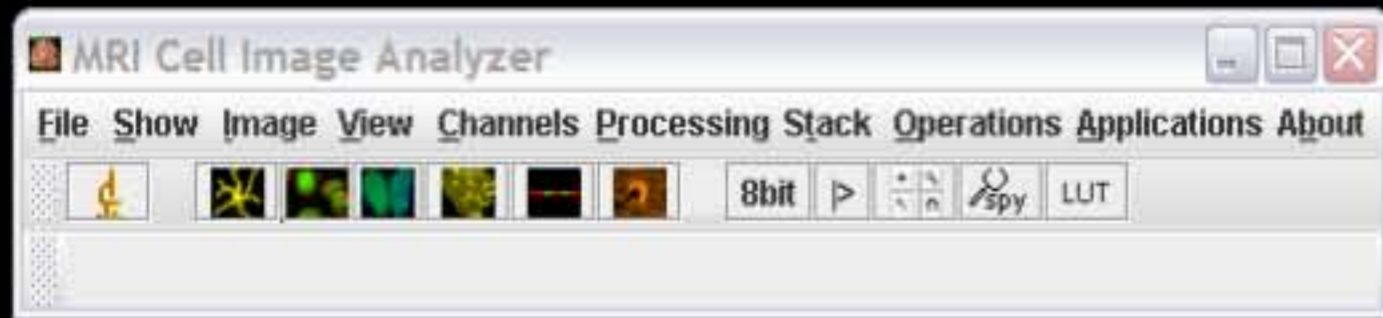


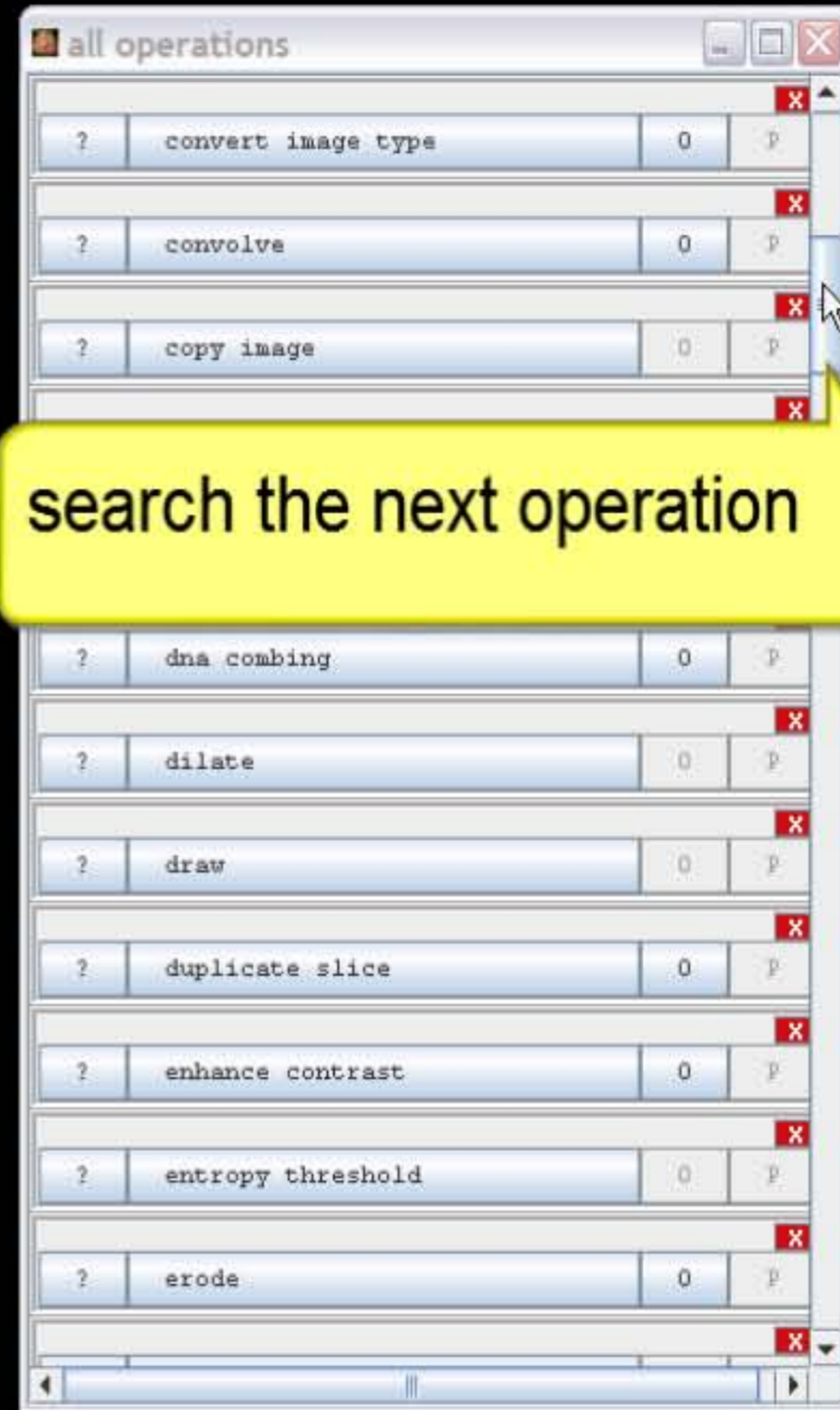
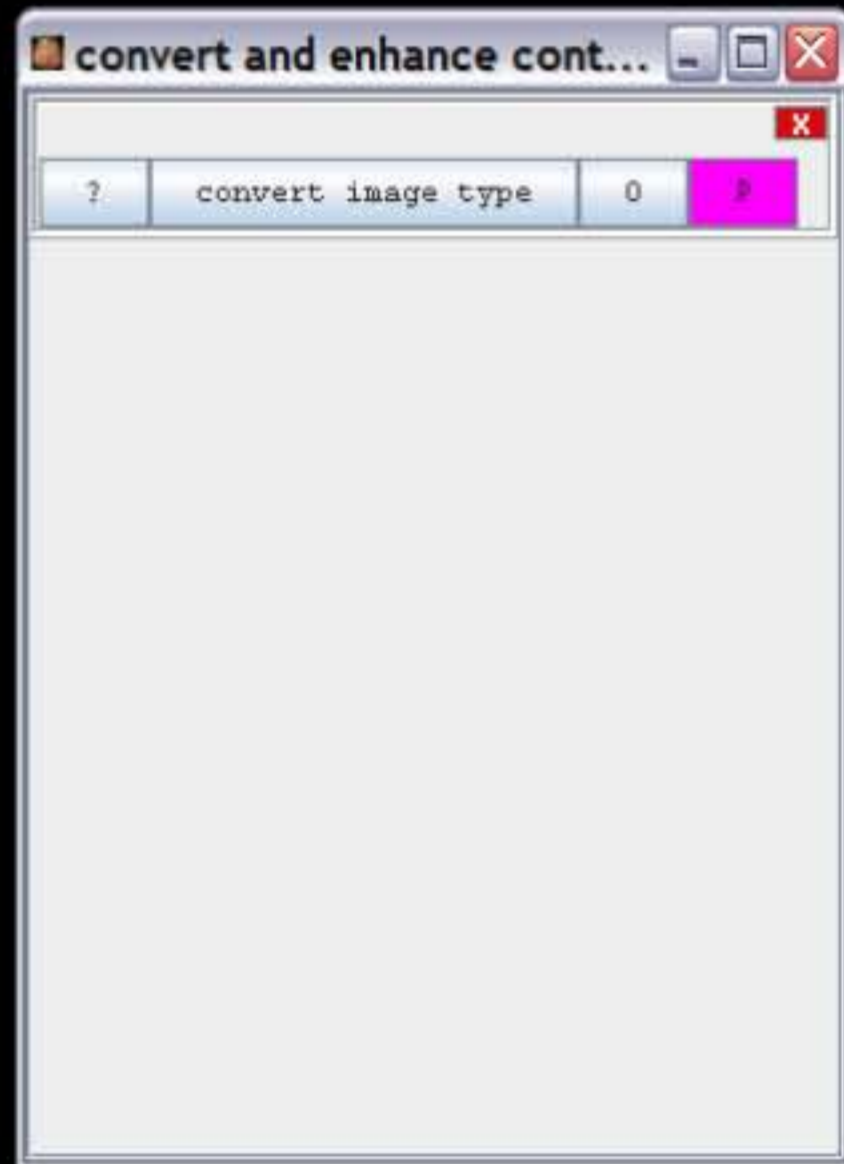


select the format to convert to

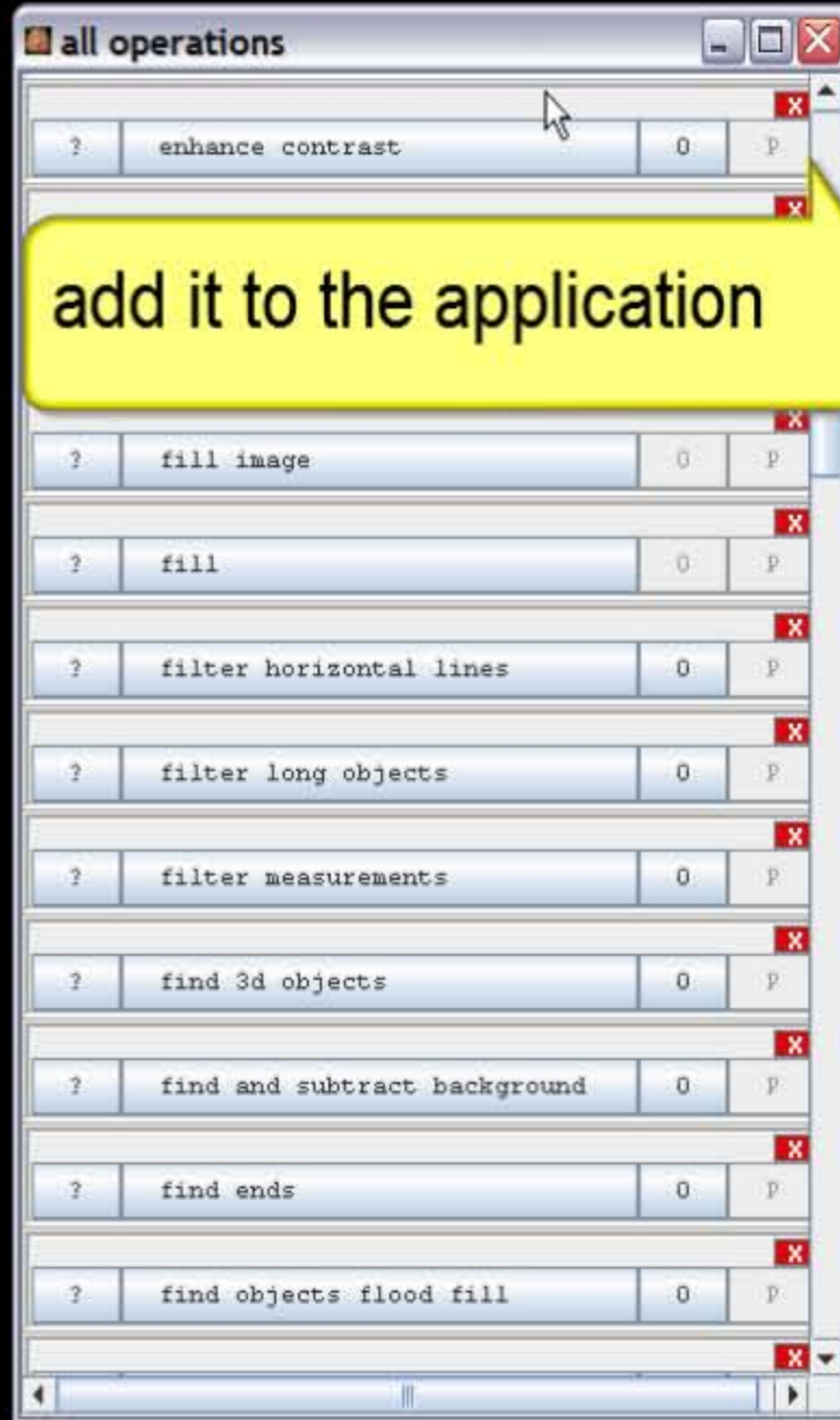
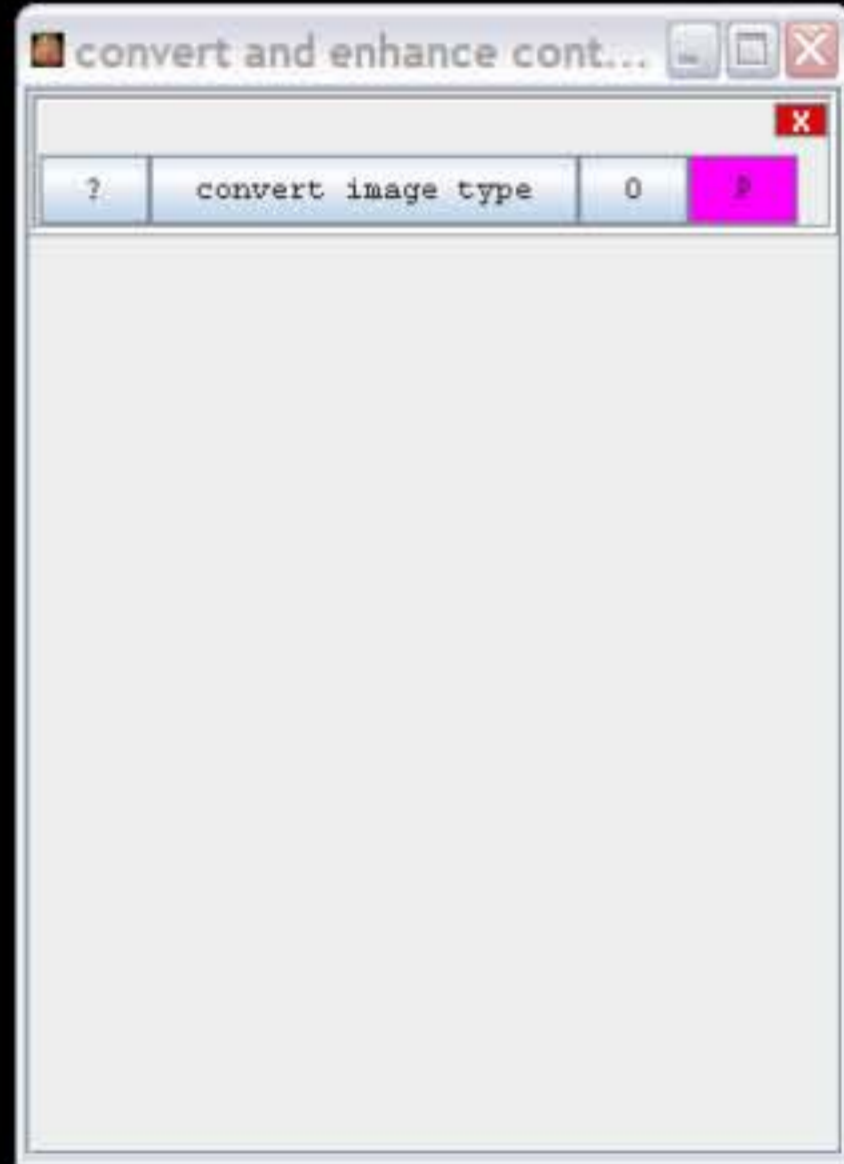








search the next operation



MRI Cell Image Analyzer

File Show Image View Channels Processing Stack Operations Applications About

8bit spy LUT

x=1224, y=468, value=21

convert and enhance contrast

0 P

convert and enhance cont...

convert image type 0 P

enhance contrast 0 P

all operations

?	enhance contrast	0	P
?	entropy threshold	0	P
?	erode	0	P
?	fill image	0	P
?	fill	0	P
?	filter horizontal lines	0	P
?	filter long objects	0	P
?	filter measurements	0	P
?	find 3d objects	0	P
?	find and subtract background	0	P
?	find ends	0	P
?	find objects flood fill	0	P

parameters for enhance contrast

(ImagePlus) input image = null

parameter	from operation	output
input image	convert image type	

apply

Help About

operations

enhance contrast	0	P
entropy threshold	0	P
erode	0	P
fill image	0	P
?	fill	0 P
?	filter horizontal lines	0 P
?	filter long objects	0 P
?	find and subtract background	0 P
?	find ends	0 P
?	find objects flood fill	0 P

convert and enhance cont...

?	convert image type	0	P
?	enhance contrast	0	P

open the parameter dialog

parameters for enhance contrast

(ImagePlus) input image = null

parameter	from operation	output
input image	convert image type	

apply

connect input slot with output slot of previous operation

convert and enhance cont...

?	convert image type	0	P
?	enhance contrast	0	P

?	contrast	0	P
?	threshold	0	P
?	erode	0	P
?	fill image	0	P
?	fill	0	P
?	filter horizontal lines	0	P
?	filter long objects	0	P
?	filter measurements	0	P
?	find 3d objects	0	P
?	find and subtract background	0	P
?	find ends	0	P
?	find objects flood fill	0	P

parameters for enhance contrast

(ImagePlus) input image = null

parameter	from operation	output
input image	convert image type	result

apply

operations

enhance contrast	0	P	X
entropy threshold	0	P	X
erode	0	P	X
fill image	0	P	X
fill	0	P	X
filter horizontal lines	0	P	X
filter long objects	0	P	X
filter measurements	0	P	X
find 3d objects	0	P	X
find and subtract background	0	P	X
find ends	0	P	X
find objects flood fill	0	P	X

convert and enhance cont...

convert image type	0	P	X
enhance contrast	0	P	X

parameters for enhance contrast

(ImagePlus) input image = null

parameter	from operation	output
input image	convert image type	result

apply

operations

- enhance contrast 0 P
- entropy threshold 0 P
- erode 0 P
- fill image 0 P
- fill ? 0 P
- filter horizontal lines ? 0 P
- filter long objects ? 0 P
- filter measurements ? 0 P
- find 3d objects ? 0 P
- find and subtract background ? 0 P
- find ends ? 0 P
- find objects flood fill ? 0 P

convert and enhance cont...

- ? convert image type 0 P
- ? enhance contrast 0 P

parameters for enhance contrast

(ImagePlus) input image = null

parameter	from operation	output
input image	convert image type	result

apply

operations

- enhance contrast 0 P
- entropy threshold 0 P
- erode 0 P
- fill image 0 P
- fill ? 0 P
- filter horizontal lines ? 0 P
- filter long objects ? 0 P
- filter measurements ? 0 P
- find 3d objects ? 0 P
- find and subtract background ? 0 P
- find ends ? 0 P
- find objects flood fill ? 0 P

convert and enhance cont...

- ? convert image type 0 P
- ? enhance contrast 0 P

parameters for enhance contrast

(ImagePlus) input image = convert image type Result

parameter	from operation	output
input image	convert image type	result

apply

operations

enhance contrast	0	P
entropy threshold	0	P
erode	0	P
fill image	0	P
fill	0	P
filter horizontal lines	0	P
filter long objects	0	P
filter measurements	0	P
find 3d objects	0	P
find and subtract background	0	P
find ends	0	P
find objects flood fill	0	P

convert and enhance cont...

convert image type	0	P
enhance contrast	0	P

parameters for enhance contrast

(ImagePlus) input image = convert image type Result

parameter	from operation	output
input image	convert image type	result

apply

operations

- enhance contrast 0 P
- entropy threshold 0 P
- erode 0 P
- fill image 0 P
- fill ? 0 P
- filter horizontal lines ? 0 P
- filter long objects ? 0 P
- filter measurements ? 0 P
- find 3d objects ? 0 P
- find and subtract background ? 0 P
- find ends ? 0 P
- find objects flood fill ? 0 P

convert and enhance cont...

- ? convert image type 0 P
- ? enhance contrast 0 P

MRI Cell Image Analyzer

File Show Image View Channels Processing Stack Operations Applications About

8bit P LUT

x=1224, y=468, value=21

convert and enhance contrast 0 P

convert and enhance cont...

convert image type 0 P

enhance contrast 0 P

all operations

? enhance contrast	0	P
? entropy threshold	0	P
? erode	0	P
? fill image	0	P
? fill	0	P
? filter horizontal lines	0	P
? filter long objects	0	P
? filter measurements	0	P
? find 3d objects	0	P
? find and subtract background	0	P
? find ends	0	P
? find objects flood fill	0	P

MRI Cell Image Analyzer

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8bit ▶ 🔍 spy LUT

x=1224, y=468, value=21

?

convert and enhance contrast 0 P

convert and enhance cont...

?

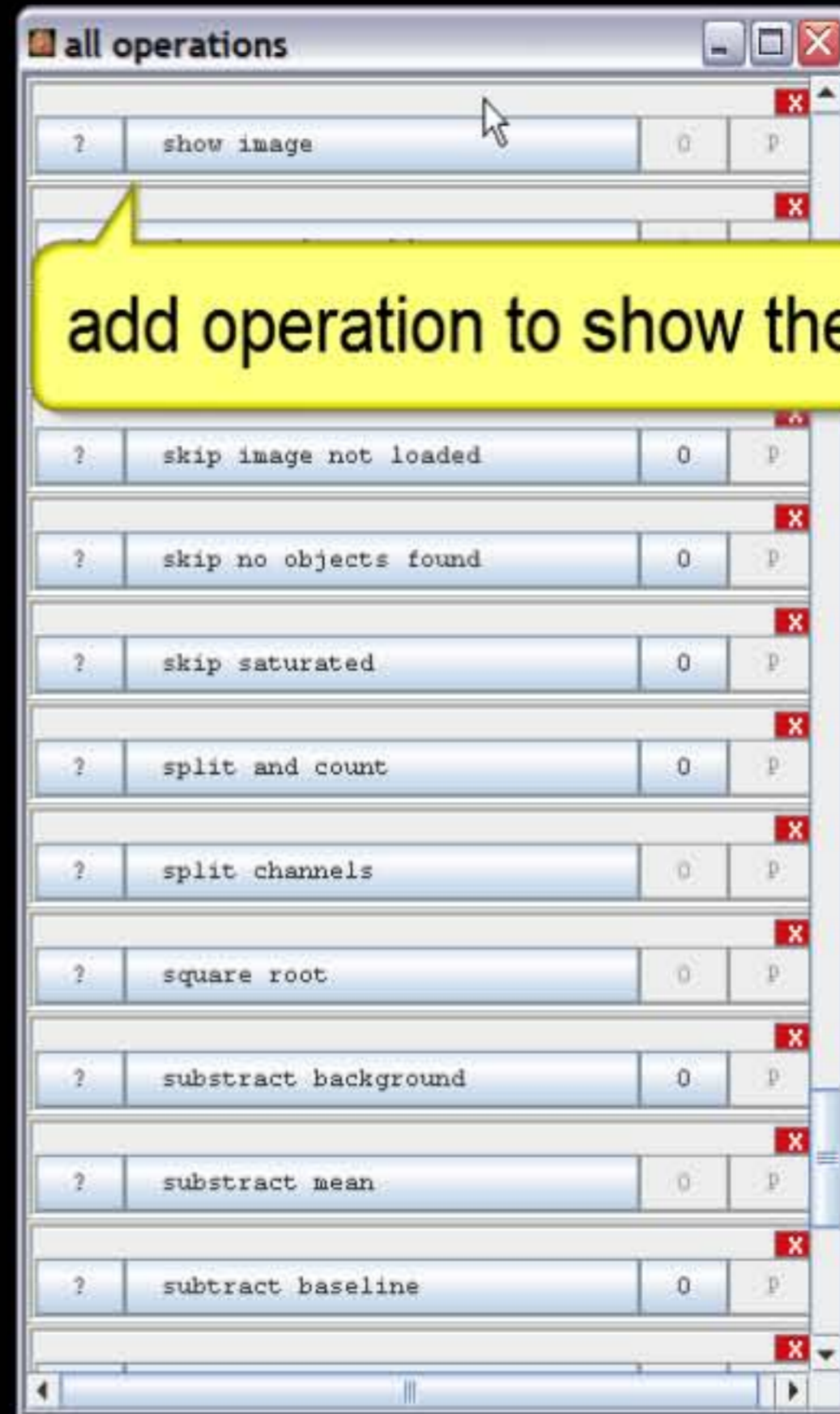
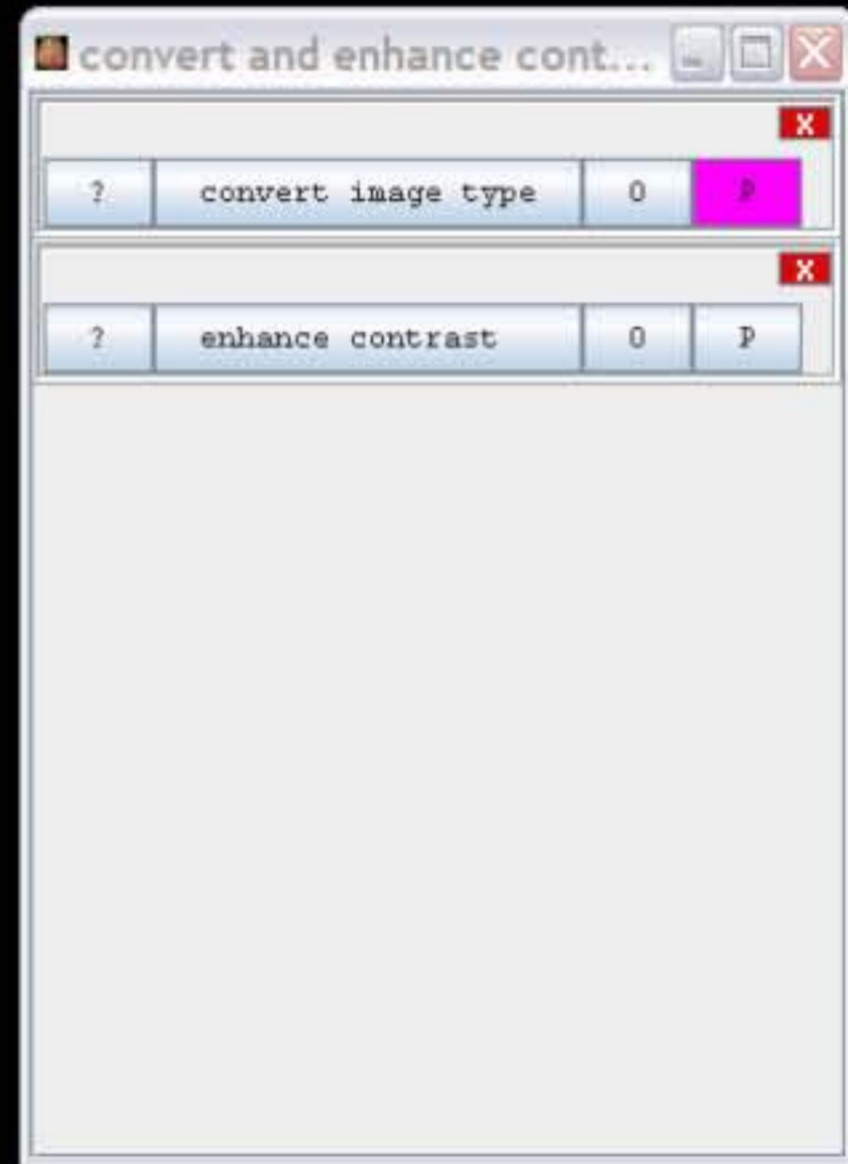
convert image type 0 P

?

enhance contrast 0 P

all operations

?	enhance contrast	0	P
?	entropy threshold	0	P
?	erode	0	P
?	fill image	0	P
?	fill	0	P
?	filter horizontal lines	0	P
?	filter long objects	0	P
?	filter measurements	0	P
?	find 3d objects	0	P
?	find and subtract background	0	P
?	find ends	0	P
?	find objects flood fill	0	P



add operation to show the result

MRI Cell Image Analyzer

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x=1224, y=468, value=21

? convert and enhance contrast 0 P

convert and enhance cont...

? convert image type 0 P

? enhance contrast 0 P

? show image 0 P

all operations

? show image	0	P
? show results table	0	P
? show text	0	P
? skip image not loaded	0	P
? skip no objects found	0	P
? skip saturated	0	P
? split and count	0	P
? split channels	0	P
? square root	0	P
? subtract background	0	P
? subtract mean	0	P
? subtract baseline	0	P

MRI Cell Image Analyzer

File Show Image View Channels Processing Stack Operations Applications About

8bit ▶ LUT

x=1224, y=468, value=21

? convert and enhance contrast 0 P

convert and enhance cont...

? convert image type 0 P

? enhance contrast 0 P

? show image 0 P

all operations

? show image	0	P
? show results table	0	P
? show text	0	P
? skip image not loaded	0	P
? skip no objects found	0	P
? skip saturated	0	P
? split and count	0	P
? split channels	0	P
? square root	0	P
? subtract background	0	P
? subtract mean	0	P
? subtract baseline	0	P

(ImagePlus) input image = null

parameter	from operation	output
input image	convert image type enhance contrast	

apply

- show image 0 P
- show results table 0 P
- show text 0 P
- skip image not loaded 0 P
- skip no objects found 0 P
- skip saturated 0 P
- split and count 0 P
- split channels 0 P
- square root 0 P
- subtract background 0 P
- subtract mean 0 P
- subtract baseline 0 P

- convert image type 0 P
- enhance contrast 0 P
- show image 0 P

(ImagePlus) input image = null

parameter	from operation	output
input image	convert image type enhance contrast	

apply

show image	0	P	X
show results table	0	P	X
show text	0	P	X
skip image not loaded	0	P	X
skip no objects found	0	P	X
skip saturated	0	P	X
split and count	0	P	X
split channels	0	P	X
square root	0	P	X
subtract background	0	P	X
subtract mean	0	P	X
subtract baseline	0	P	X

? convert image type	0	P	X
? enhance contrast	0	P	X
? show image	0	P	X

(ImagePlus) input image = null

parameter	from operation	output
input image	convert image type	result
	enhance contrast	

apply

show image	0	P	X
show results table	0	P	X
show text	0	P	X
skip image not loaded	0	P	X
skip no objects found	0	P	X
skip saturated	0	P	X
split and count	0	P	X
split channels	0	P	X
square root	0	P	X
subtract background	0	P	X
subtract mean	0	P	X
subtract baseline	0	P	X

? convert image type	0	P	X
? enhance contrast	0	P	X
? show image	0	P	X

(ImagePlus) Input image = null

parameter	from operation	output
input image	convert image type enhance contrast	result

apply

show image	0	P
show results table	0	P
show text	0	P
skip image not loaded	0	P
skip no objects found	0	P
skip saturated	0	P
split and count	0	P
split channels	0	P
square root	0	P
subtract background	0	P
subtract mean	0	P
subtract baseline	0	P

convert image type	0	P
enhance contrast	0	P
show image	0	P

(ImagePlus) input image = enhance contrast Result

parameter	from operation	output
input image	convert image type	result
	enhance contrast	

apply

show image	0	P
show results table	0	P
show text	0	P
skip image not loaded	0	P
skip no objects found	0	P
skip saturated	0	P
split and count	0	P
split channels	0	P
square root	0	P
subtract background	0	P
subtract mean	0	P
subtract baseline	0	P

?	convert image type	0	P
?	enhance contrast	0	P
?	show image	0	P

(ImagePlus) Input image = enhance contrast - Result

parameter	from operation	output
input image	convert image type	result
	enhance contrast	

apply

show image	0	P
show results table	0	P
show text	0	P
skip image not loaded	0	P
skip no objects found	0	P
skip saturated	0	P
split and count	0	P
split channels	0	P
square root	0	P
subtract background	0	P
subtract mean	0	P
subtract baseline	0	P

?	convert image type	0	P
?	enhance contrast	0	P
?	show image	0	P

MRI Cell Image Analyzer

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8bit ▶ LUT

x=1224, y=468, value=21

?

convert and enhance contrast 0 P

convert and enhance cont...

?

convert image type 0 P

?

enhance contrast 0 P

?

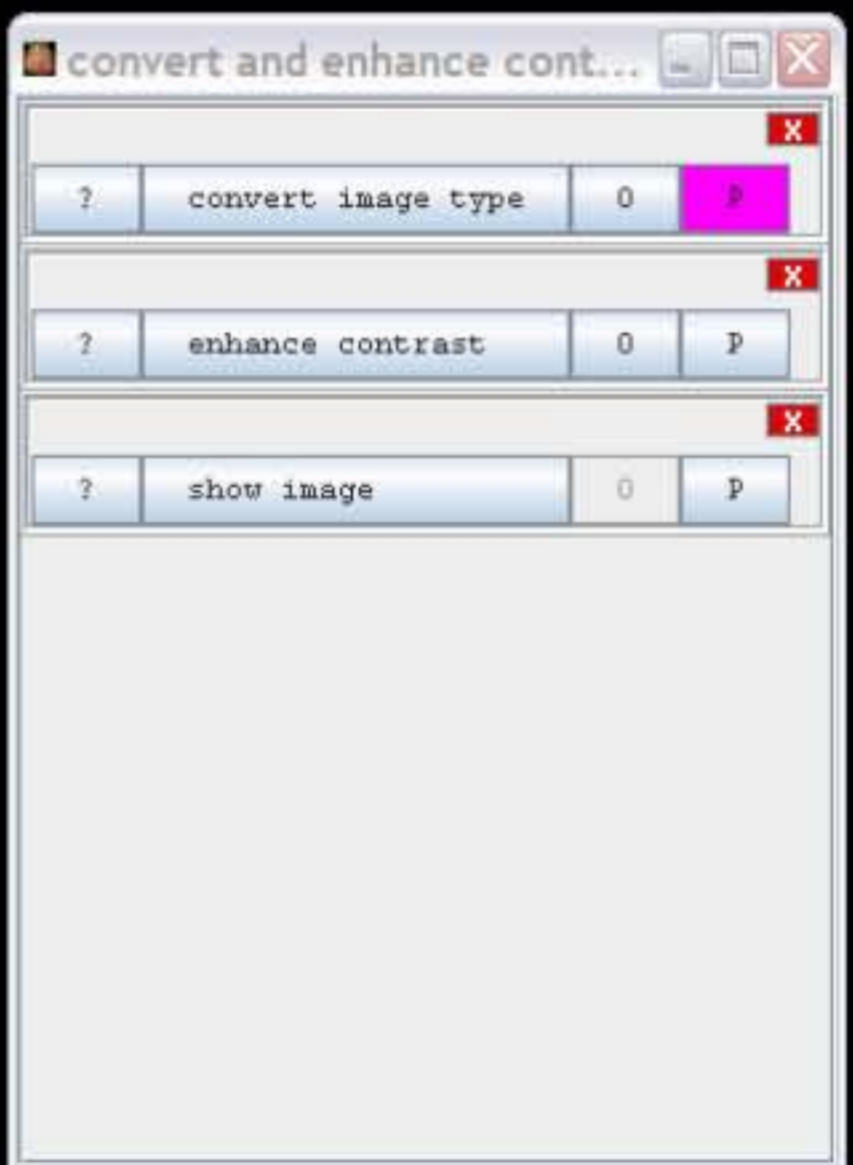
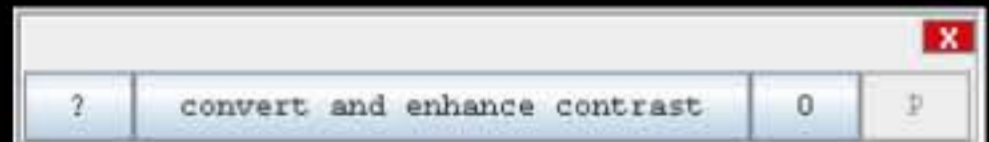
show image 0 P

all operations

?	show image	0	P
?	show results table	0	P
?	show text	0	P
?	skip image not loaded	0	P
?	skip no objects found	0	P
?	skip saturated	0	P
?	split and count	0	P
?	split channels	0	P
?	square root	0	P
?	subtract background	0	P
?	subtract mean	0	P
?	subtract baseline	0	P



open an image



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Open... Ctrl-O  8bit    LUT

Save As... Ctrl-S value=21

Quit

convert and enhance cont...

? convert and enhance contrast 0 P

convert and enhance cont...

? convert image type 0 P

? enhance contrast 0 P

? show image 0 P

all operations

? show image 0 P

? show results table 0 P

? show text 0 P

? skip image not loaded 0 P

? skip no objects found 0 P

? skip saturated 0 P

? split and count 0 P

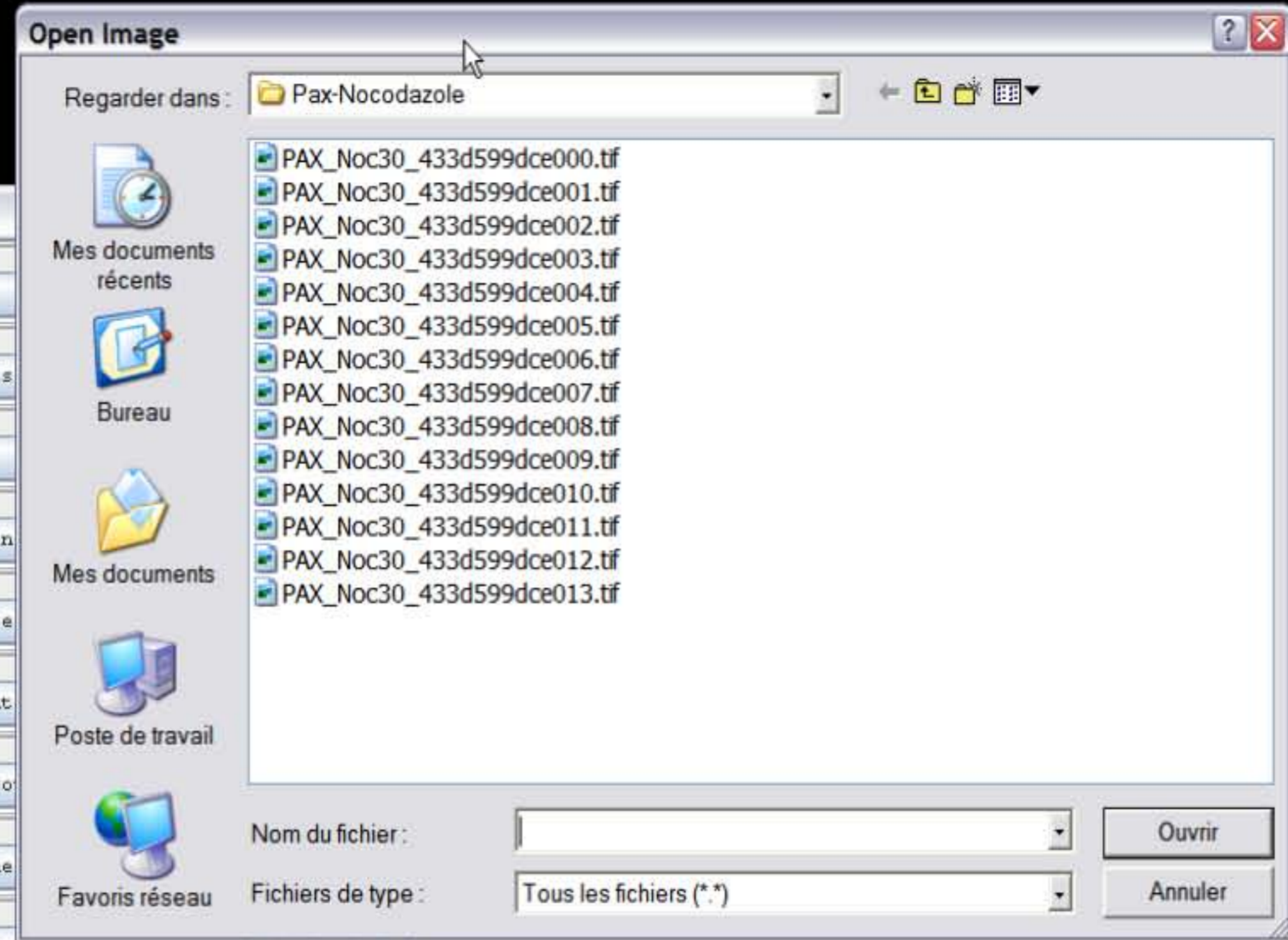
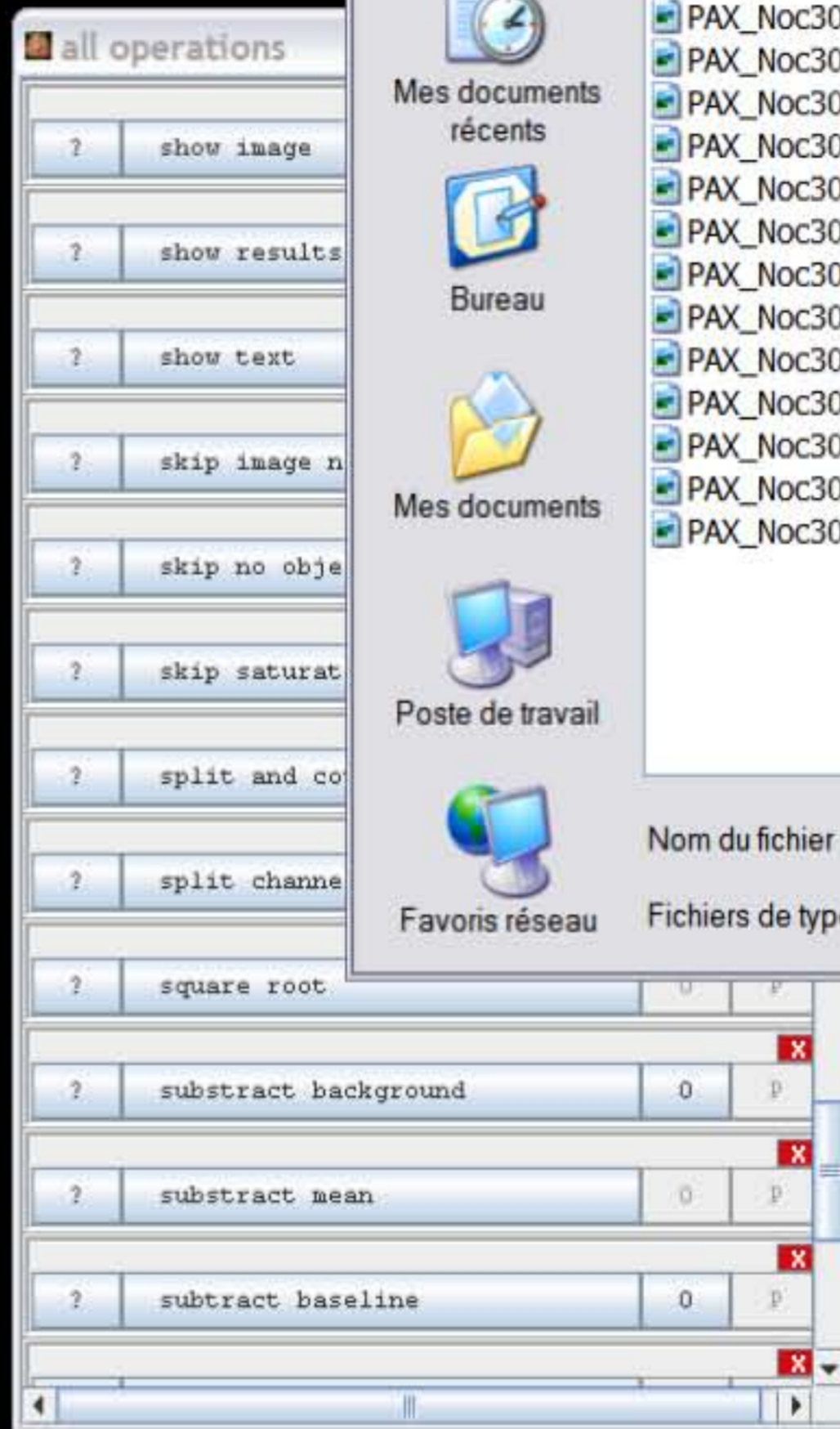
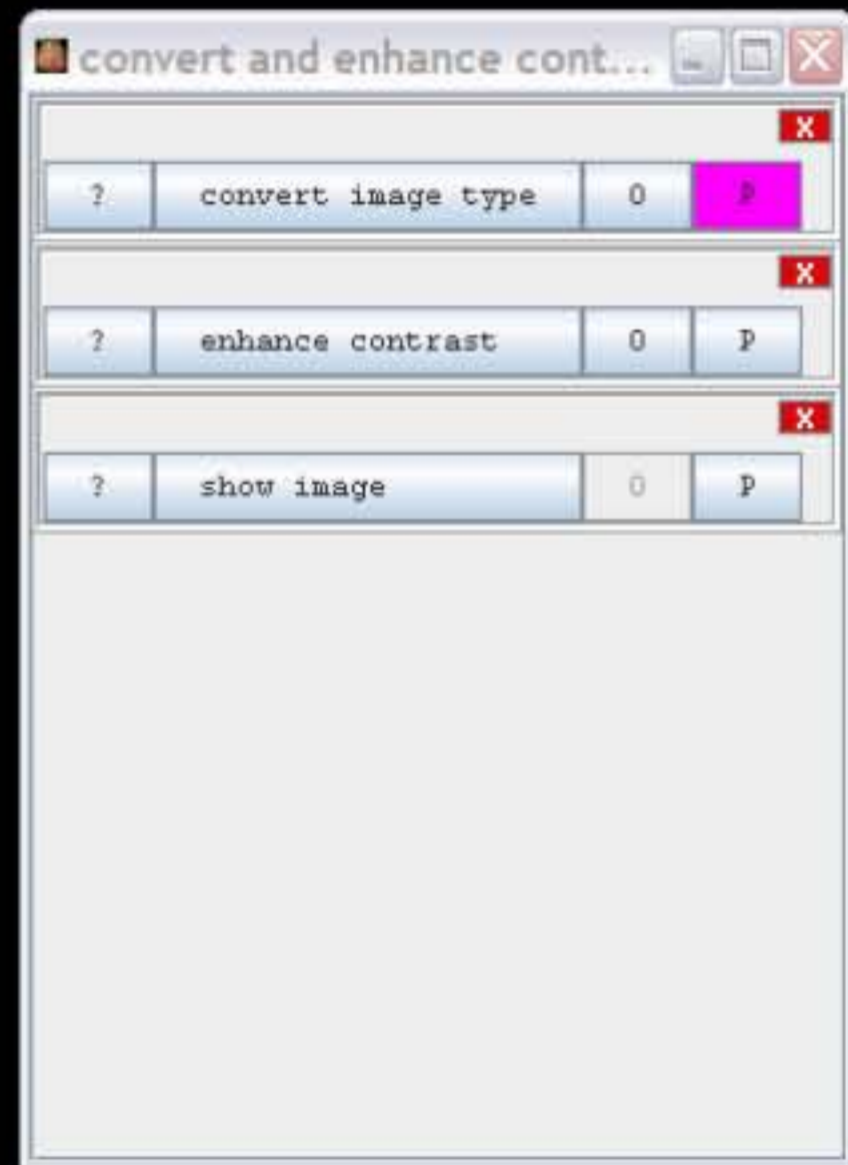
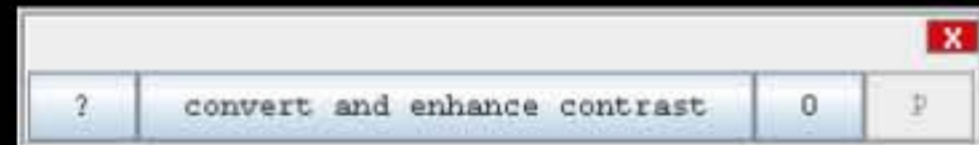
? split channels 0 P

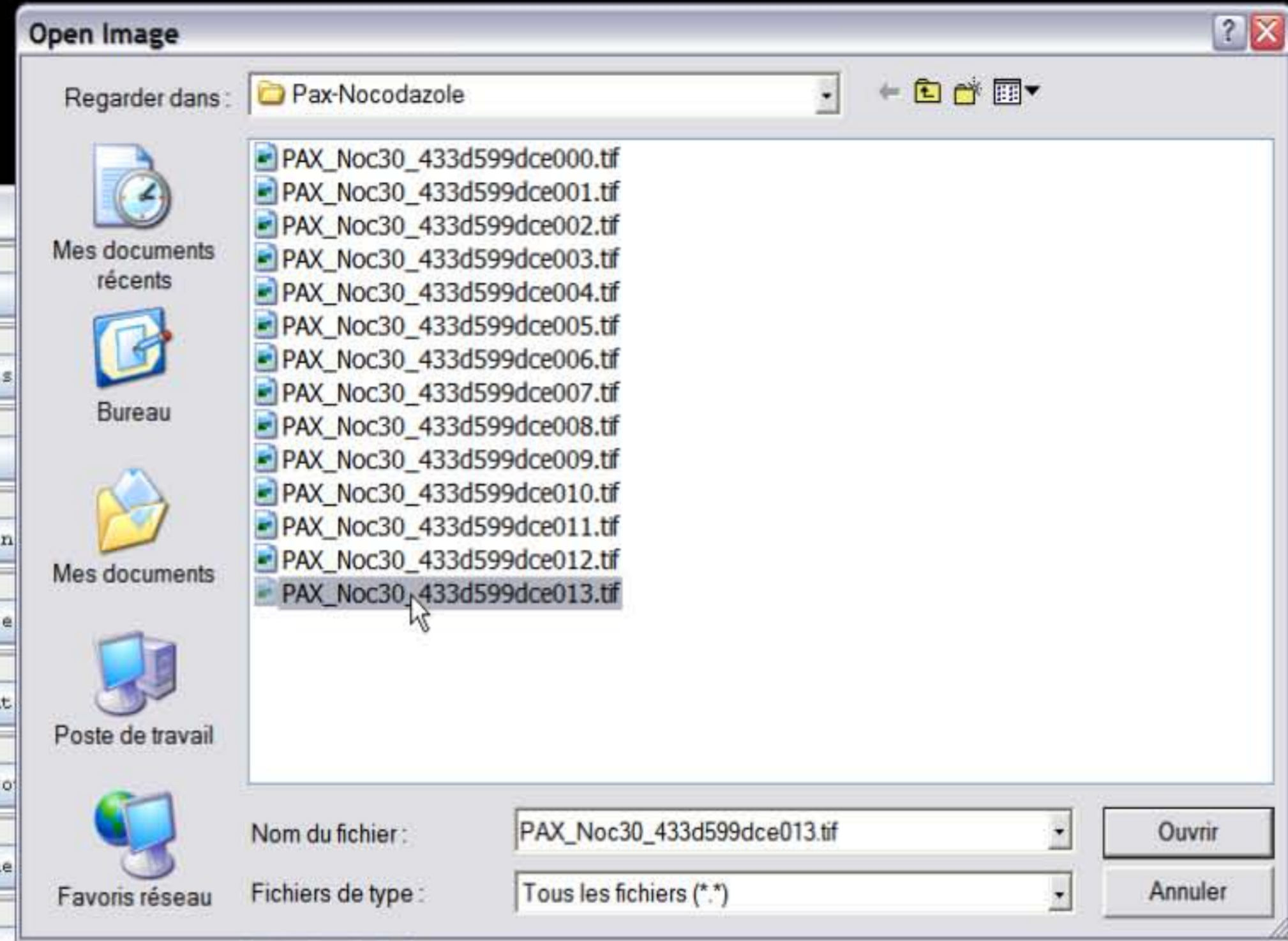
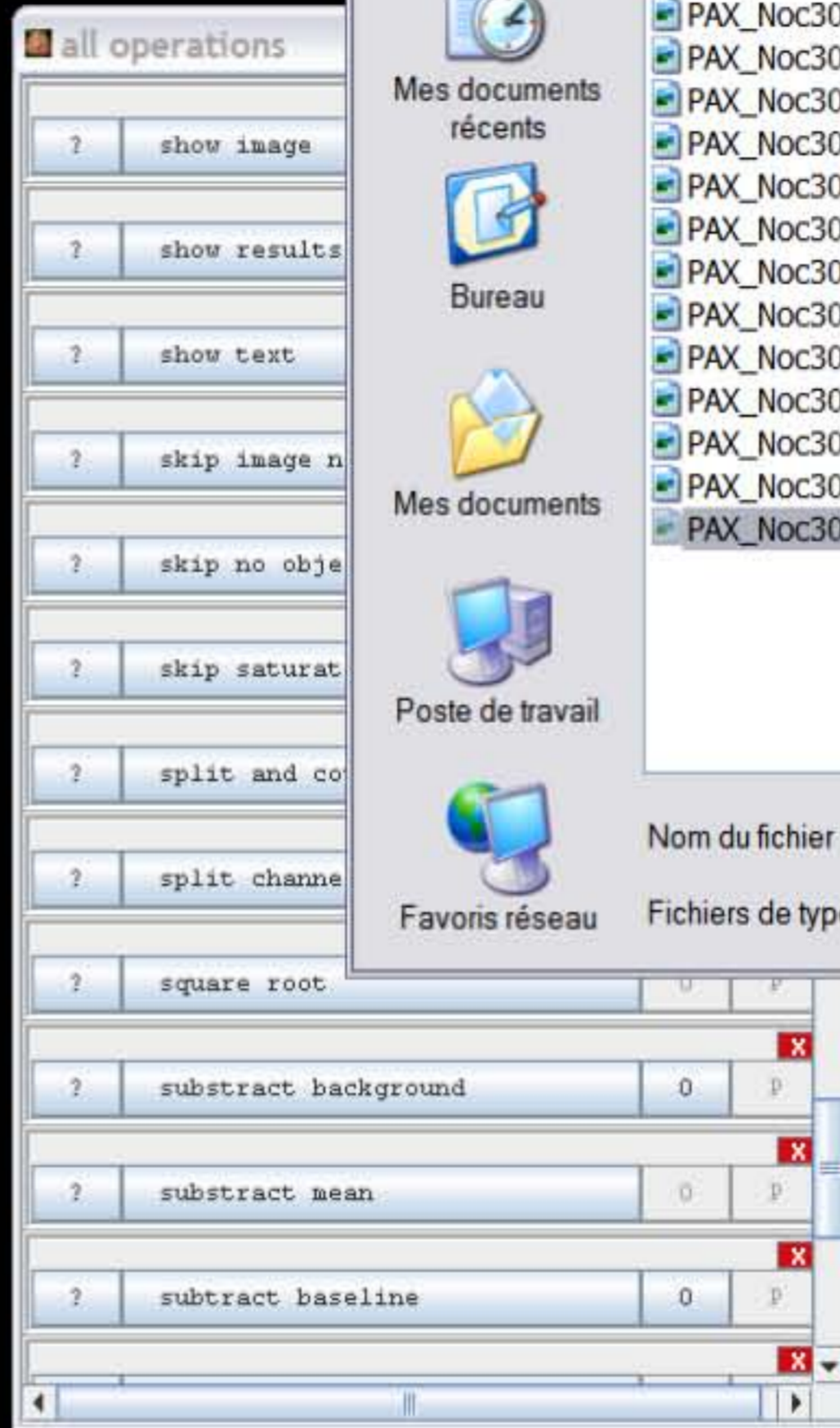
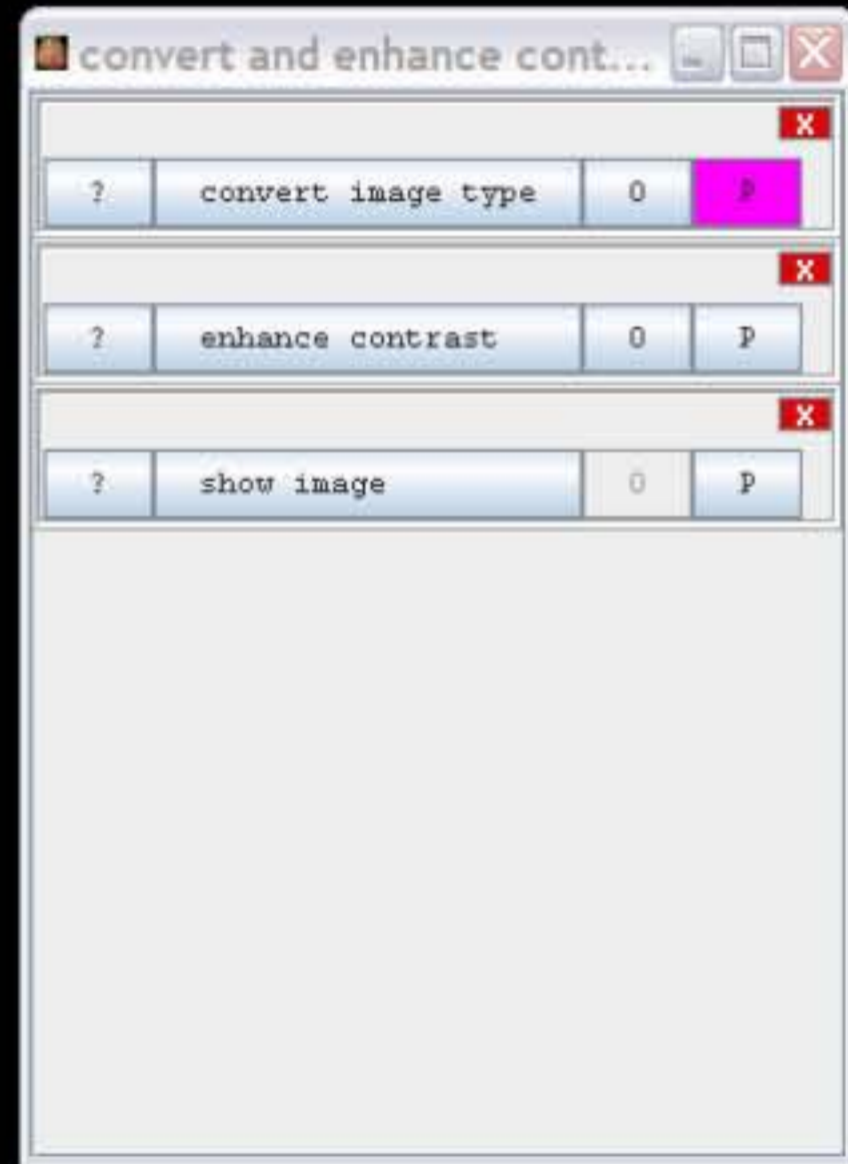
? square root 0 P

? subtract background 0 P

? subtract mean 0 P

? subtract baseline 0 P





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8bit spy LUT

convert and enhance contrast 0 P

convert and enhance cont...

convert image type 0 P

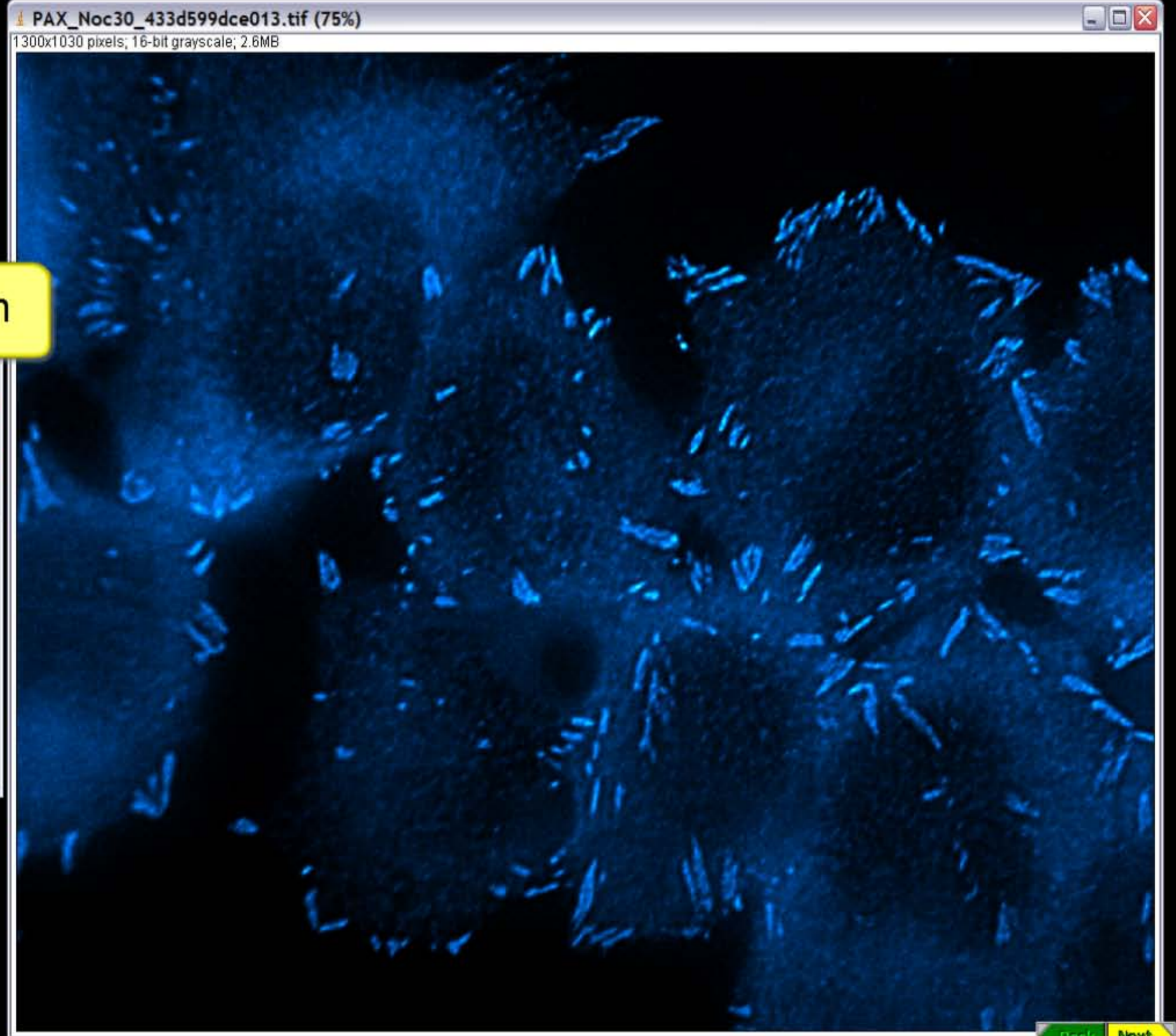
enhance contrast 0 P

show image 0 P

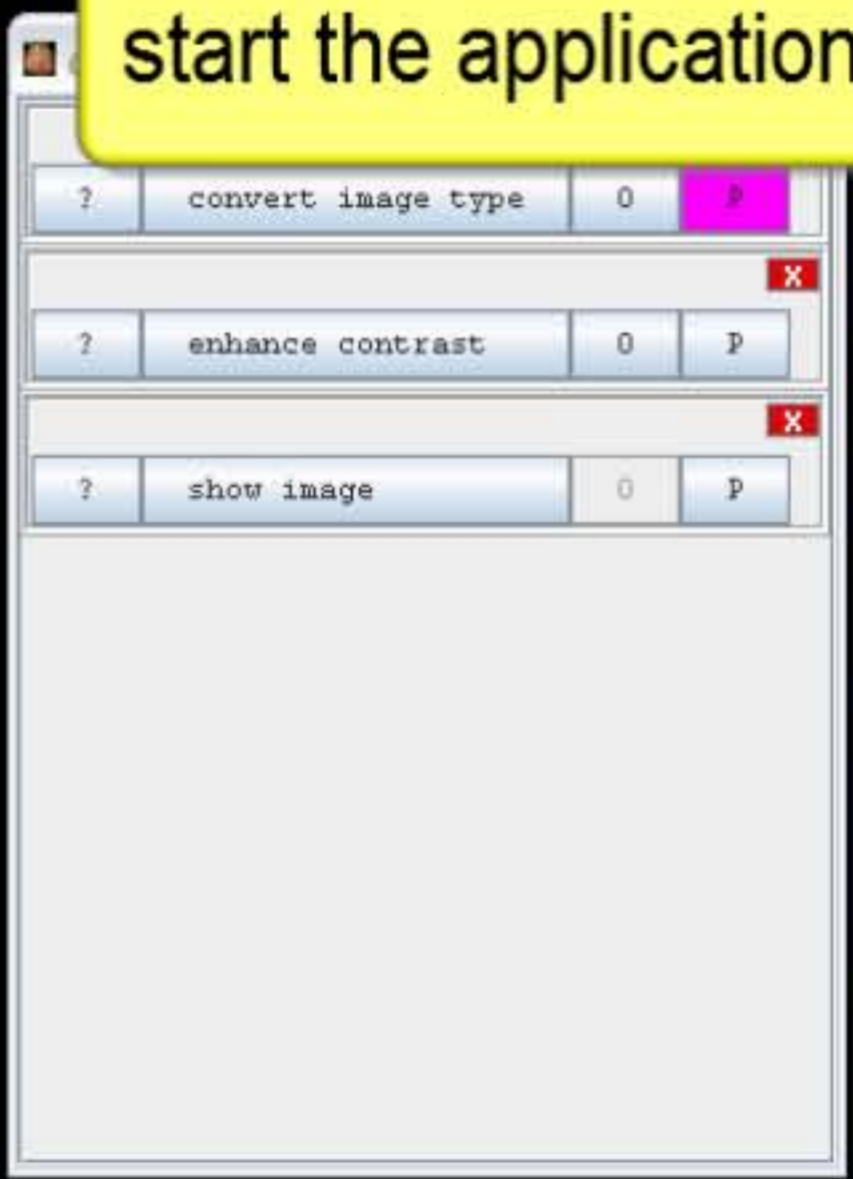
PAX_Noc30_433d599dqe013.tif (75%)

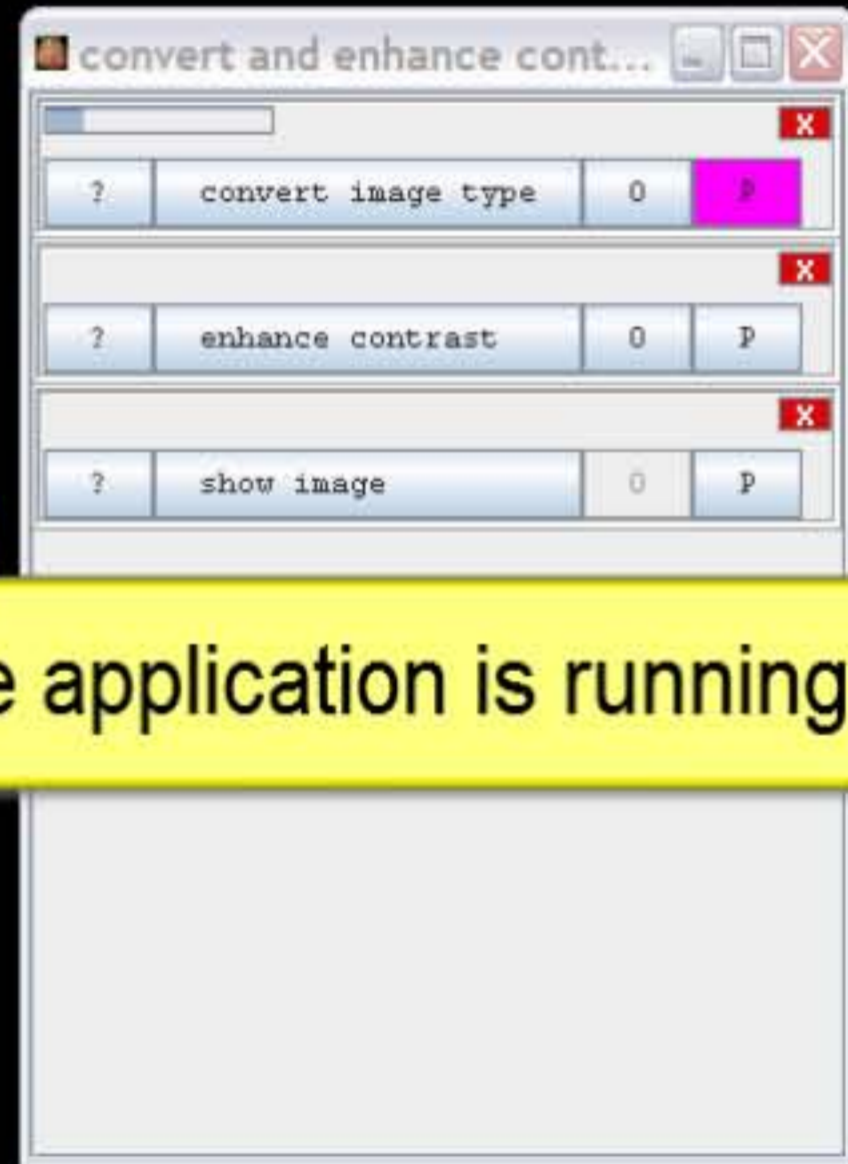
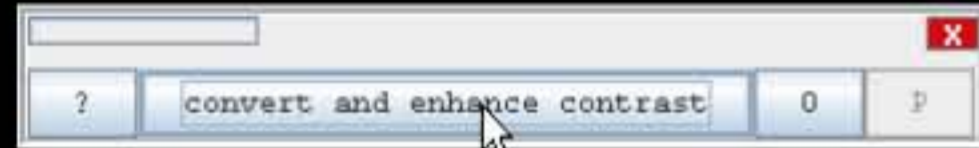
1300x1030 pixels; 16-bit grayscale; 2.6MB

Back Next

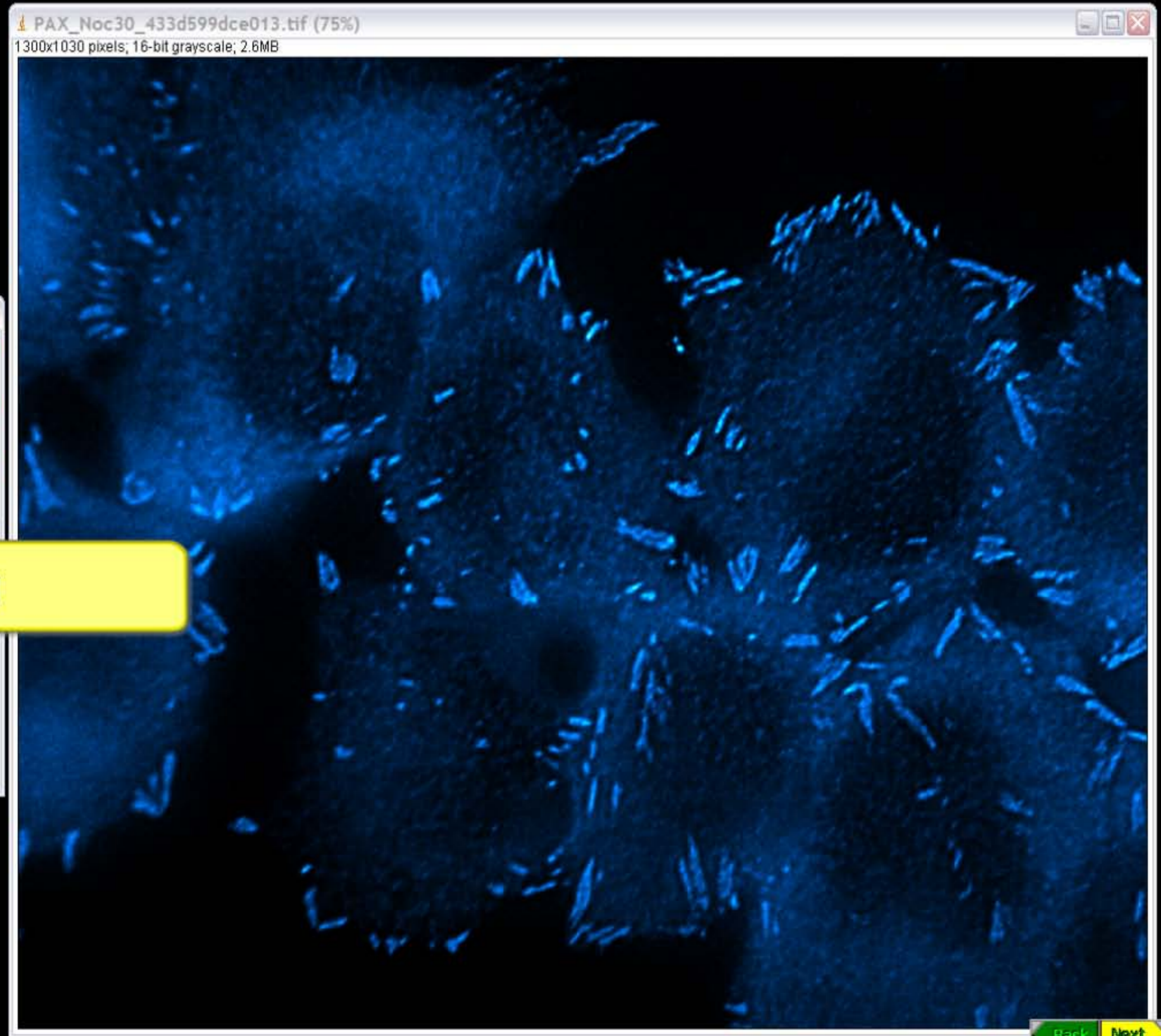


start the application





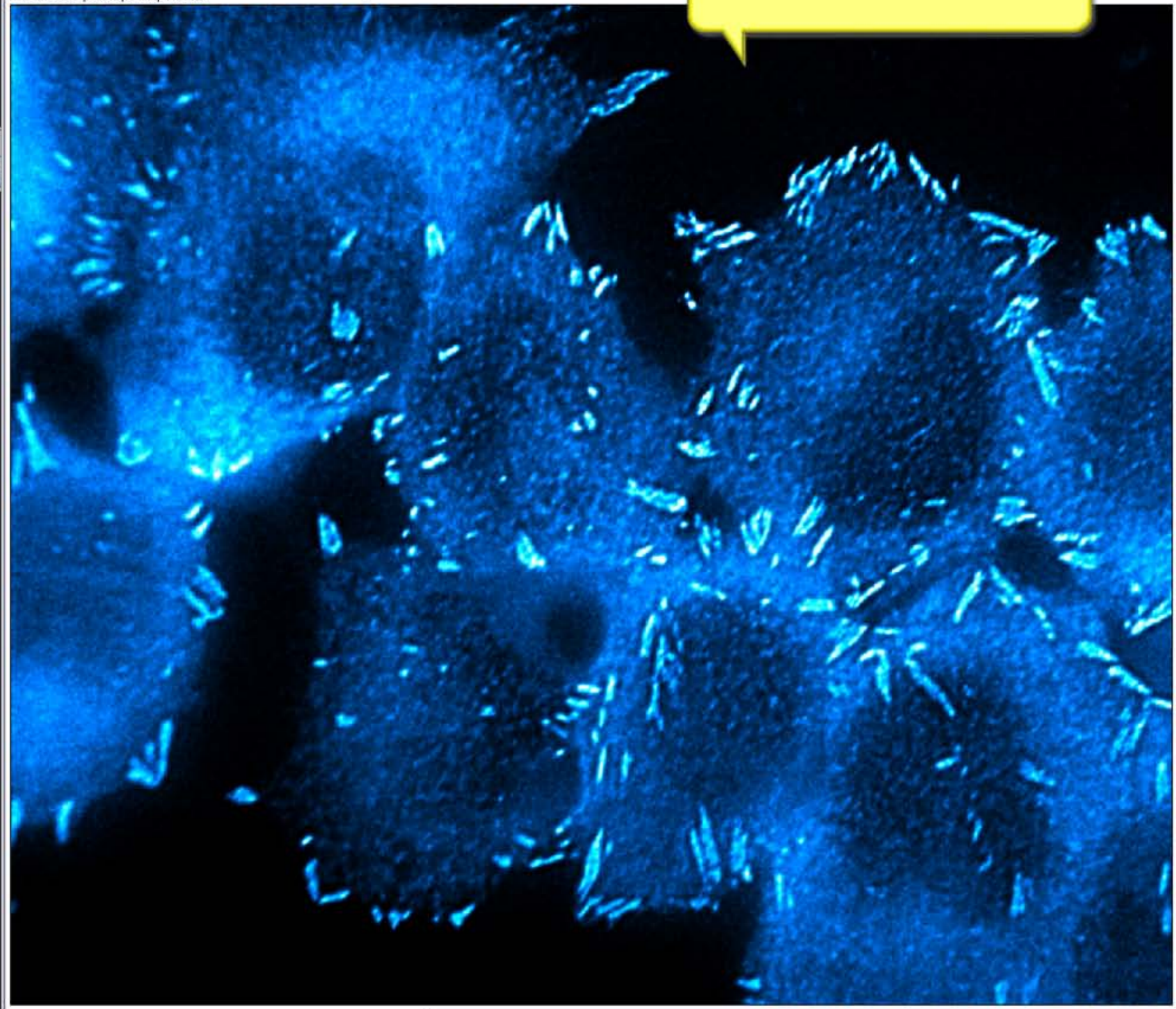
the application is running



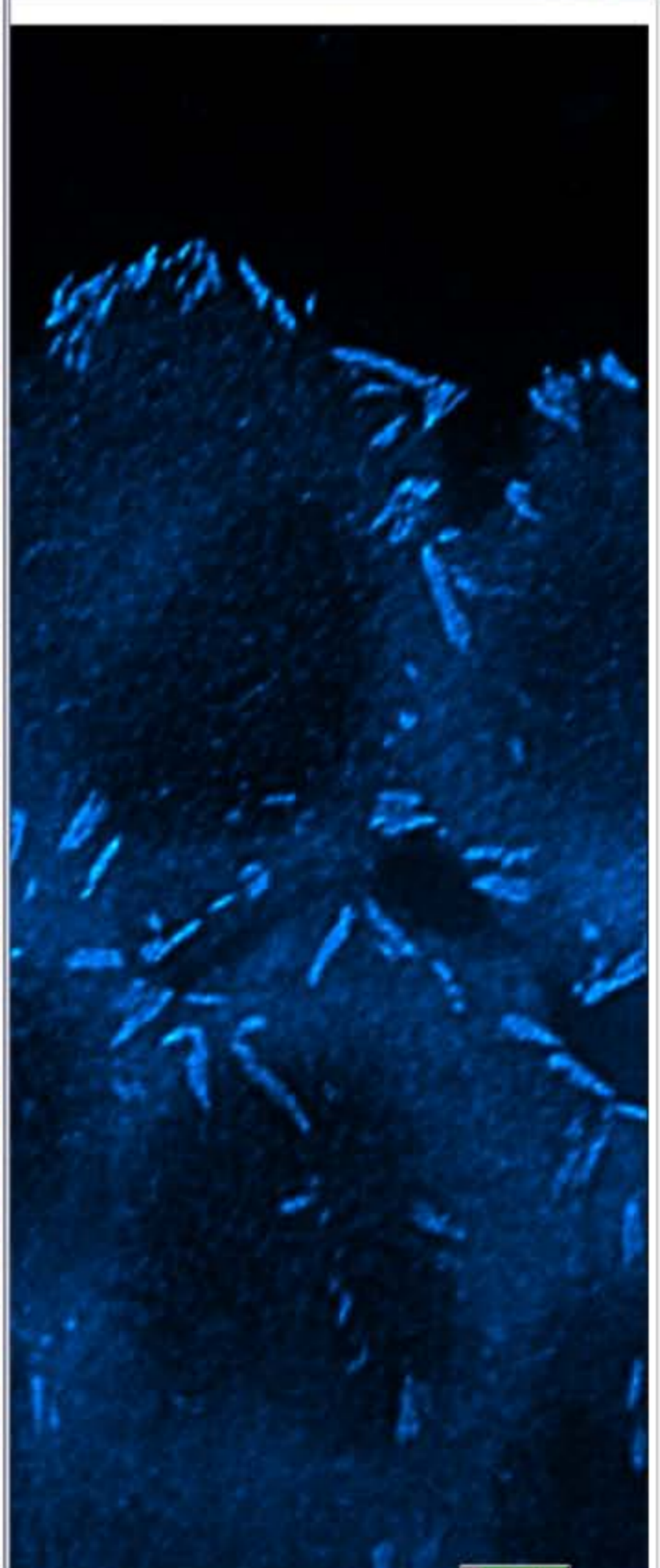


the result
- 8 bit image
- enhanced contrast

PAX_Noc30_433d599dce013.tif (75%)
1300x1030 pixels; 8-bit; 1.3MB



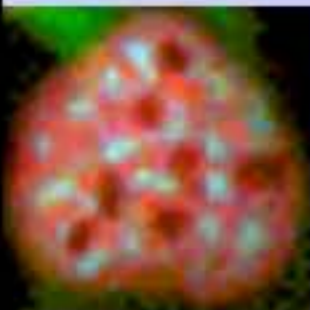
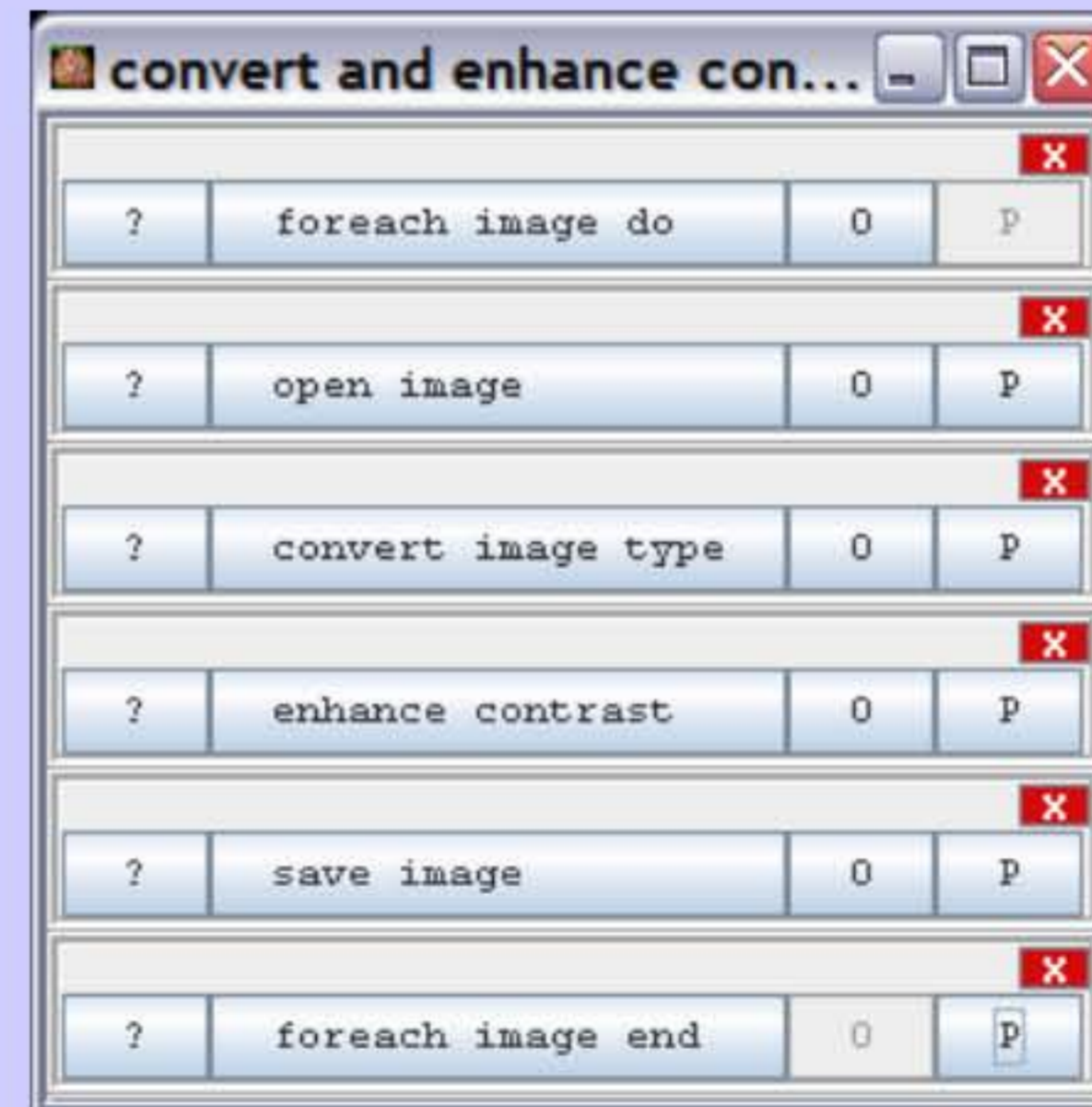
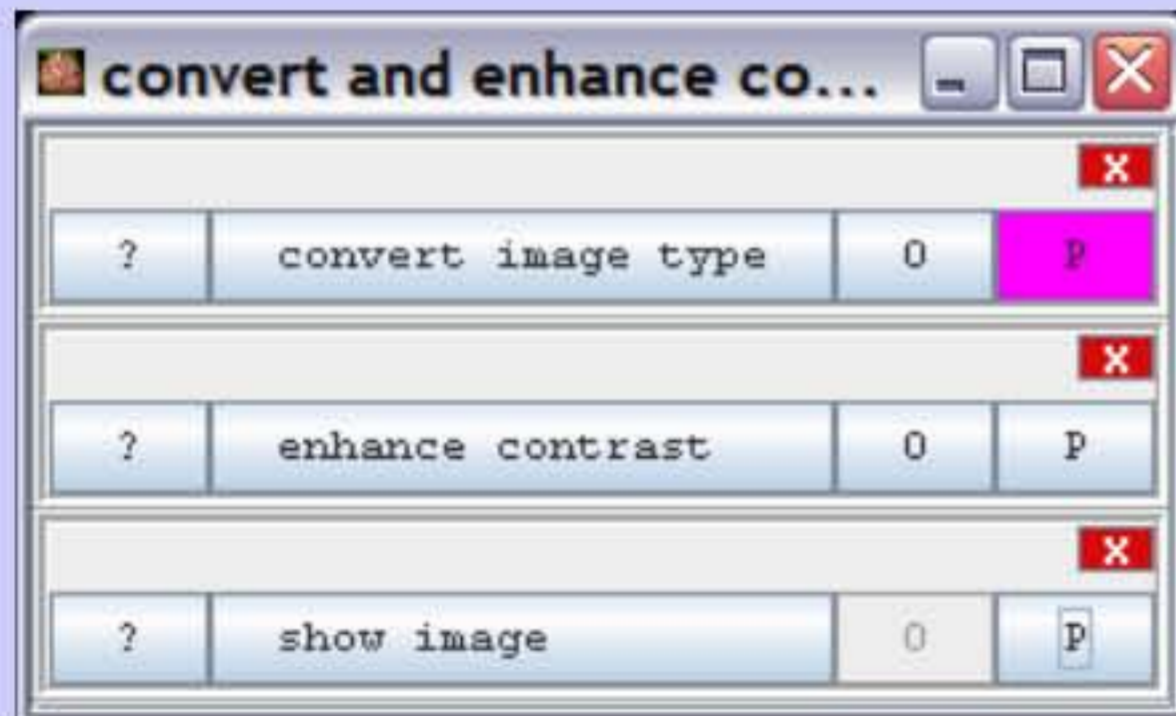
1300x1030 pixels; 8-bit; 1.3MB

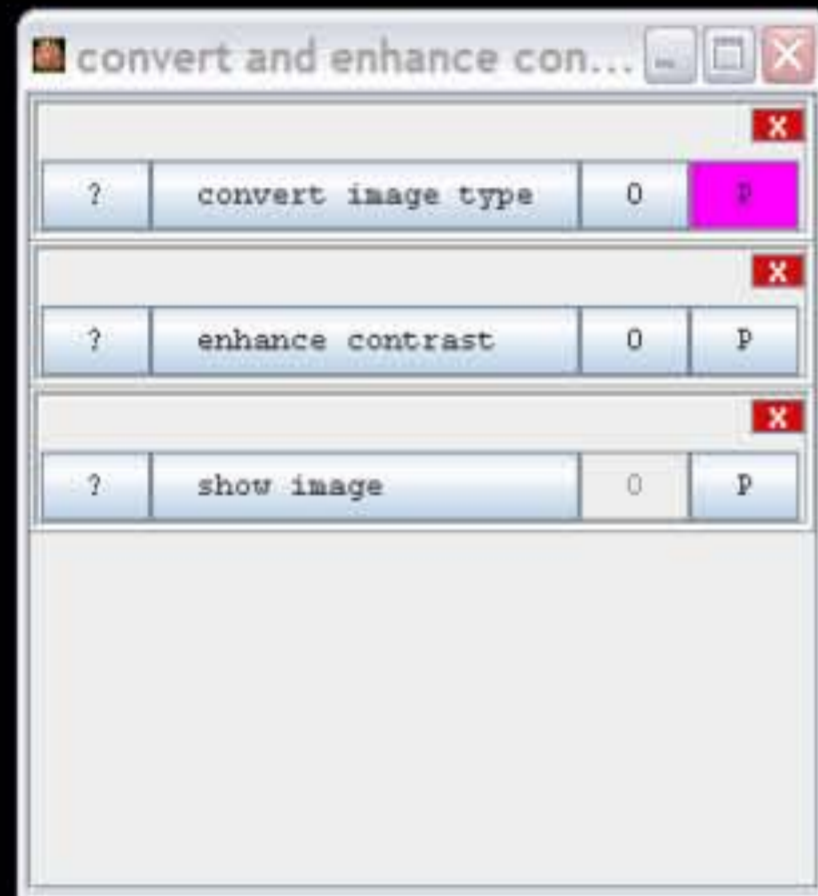
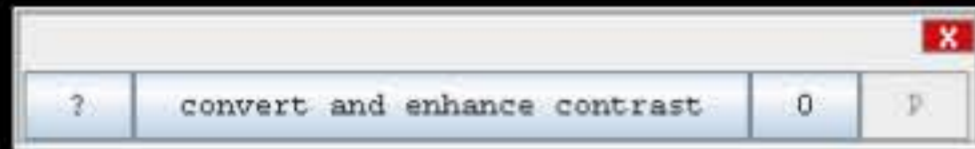


Back

Using visual scripting

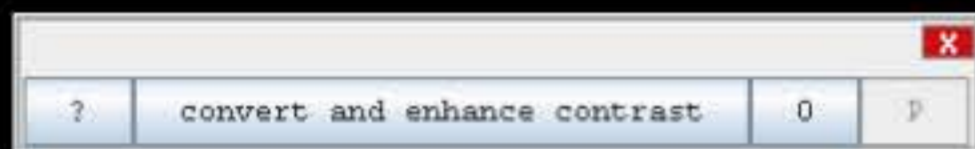
- create a batch application

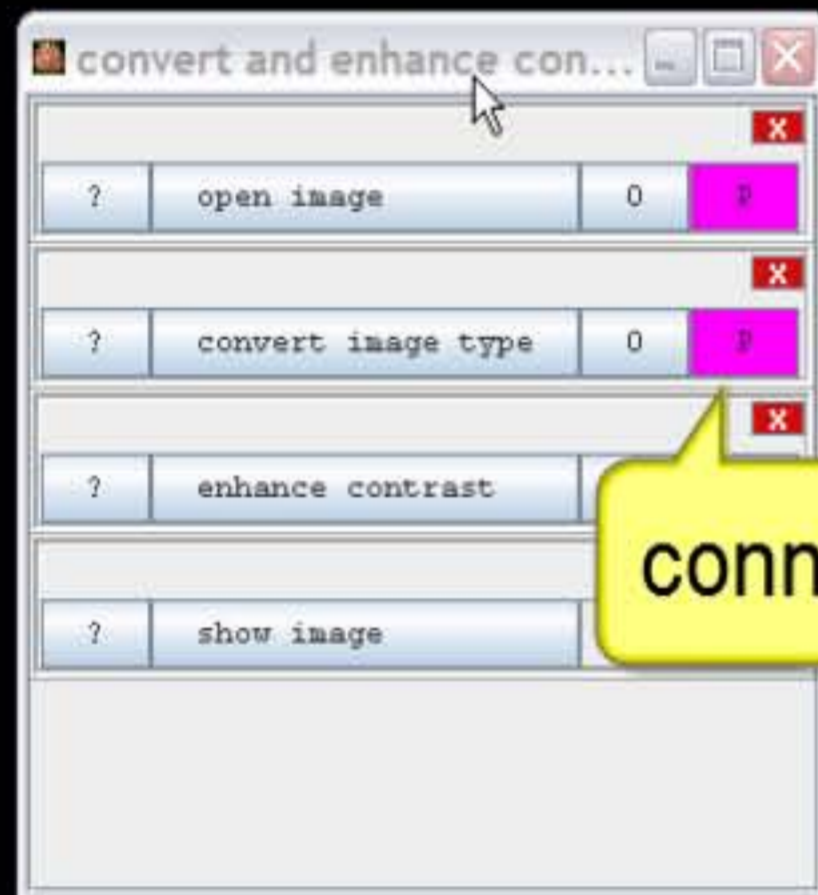
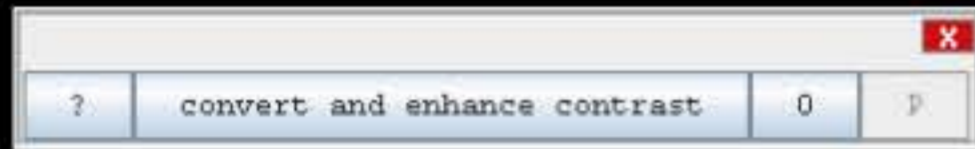




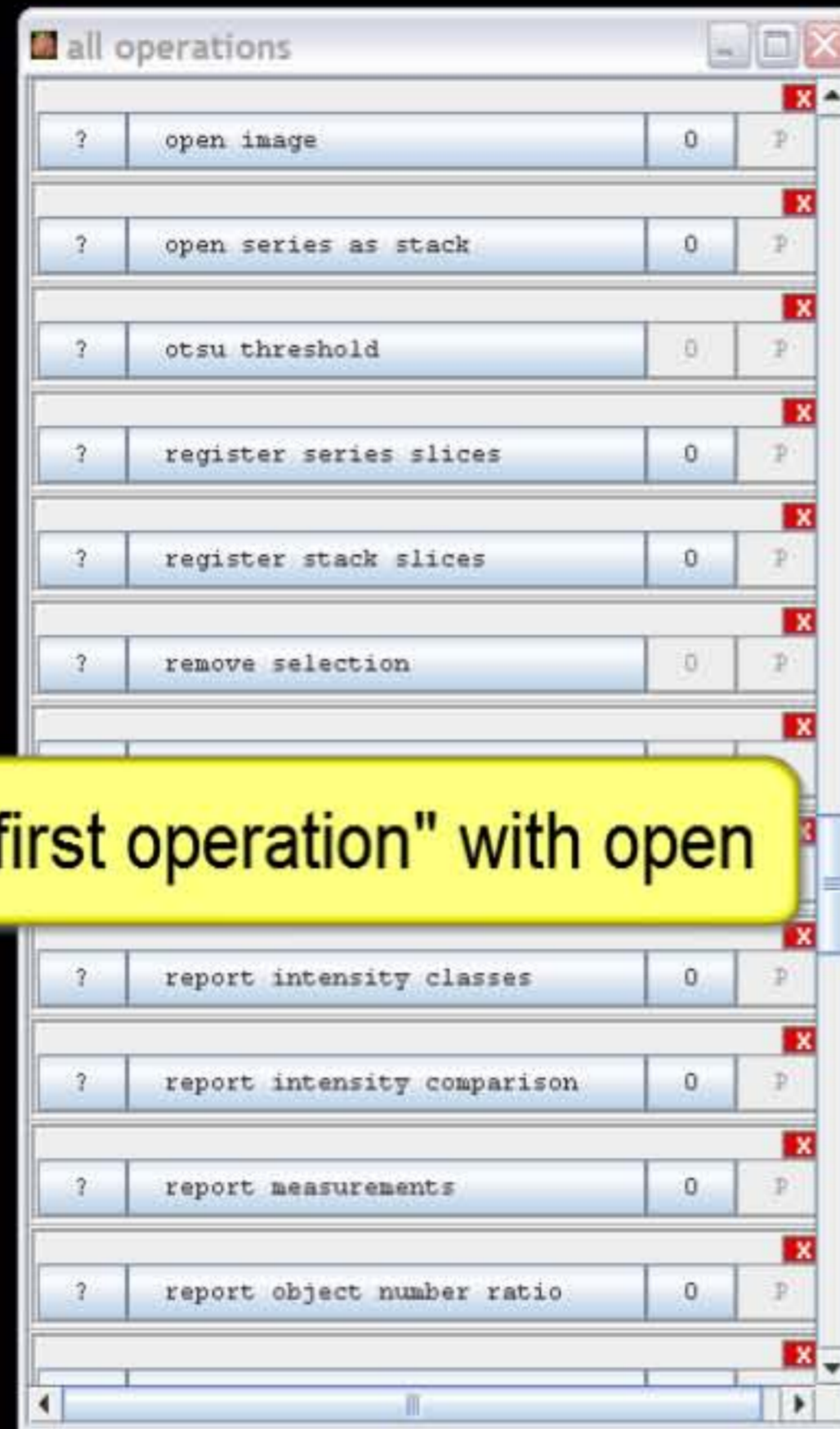
need to open image from disk now

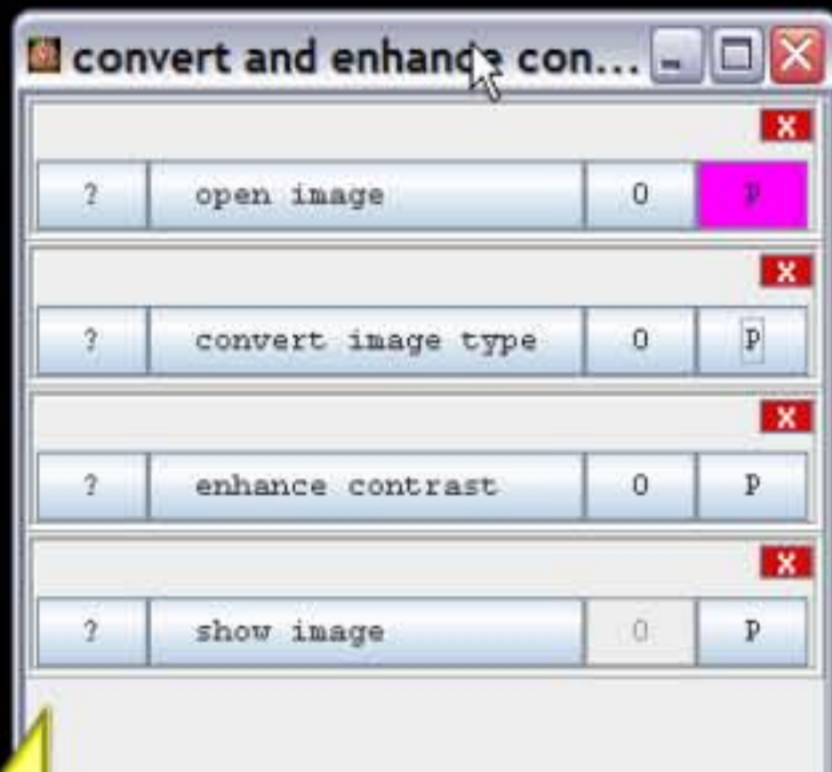
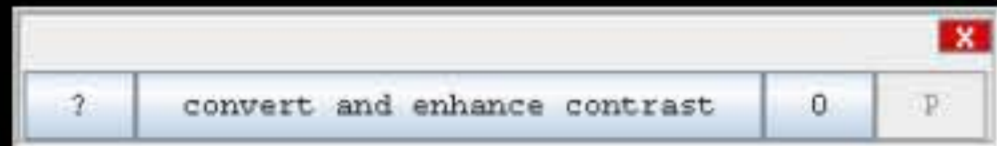






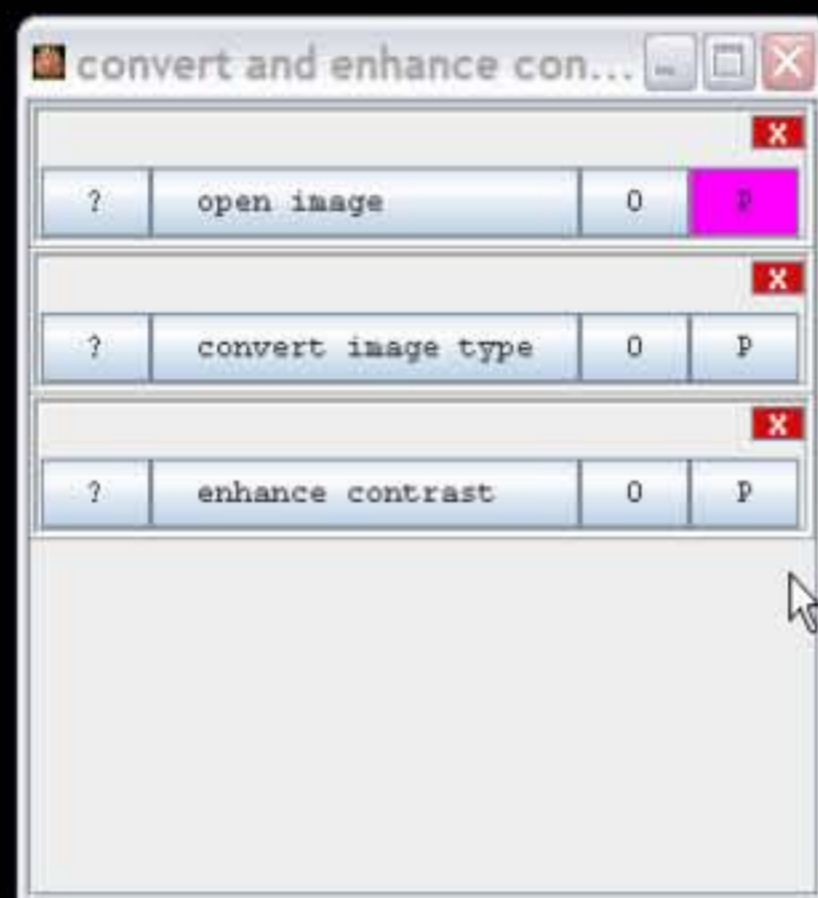
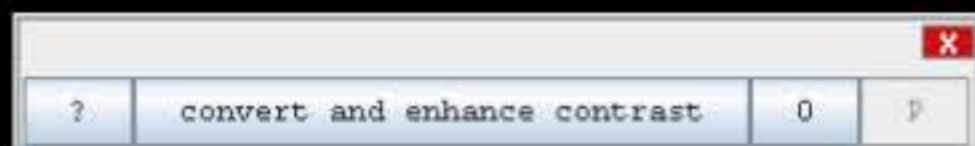
connect "first operation" with open

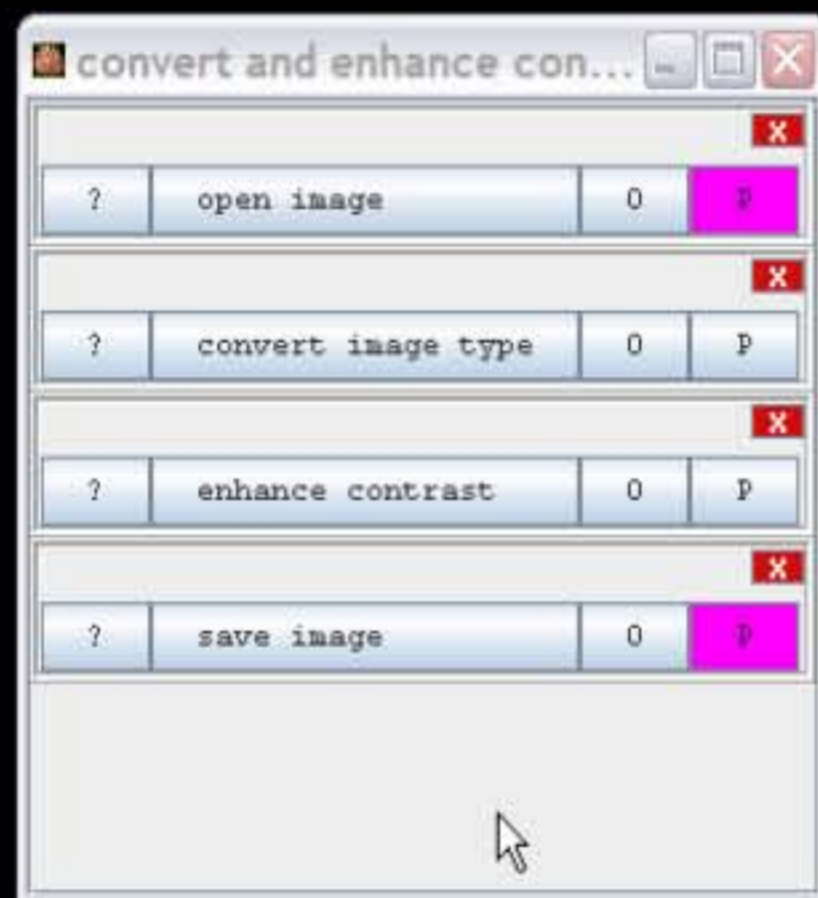
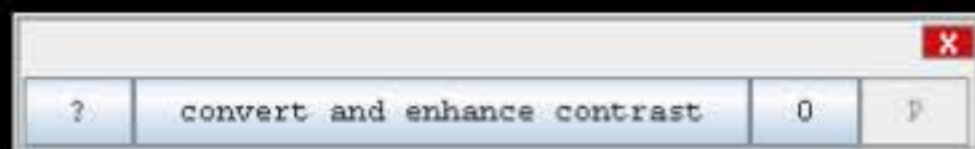




do not show the image







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parameters for save image

(ImagePlus) input image = open image | Result
 (String) path = open image | AbsoluteFilename

parameter	from operation	output
input image	open image	
path	convert image type enhance contrast	

apply

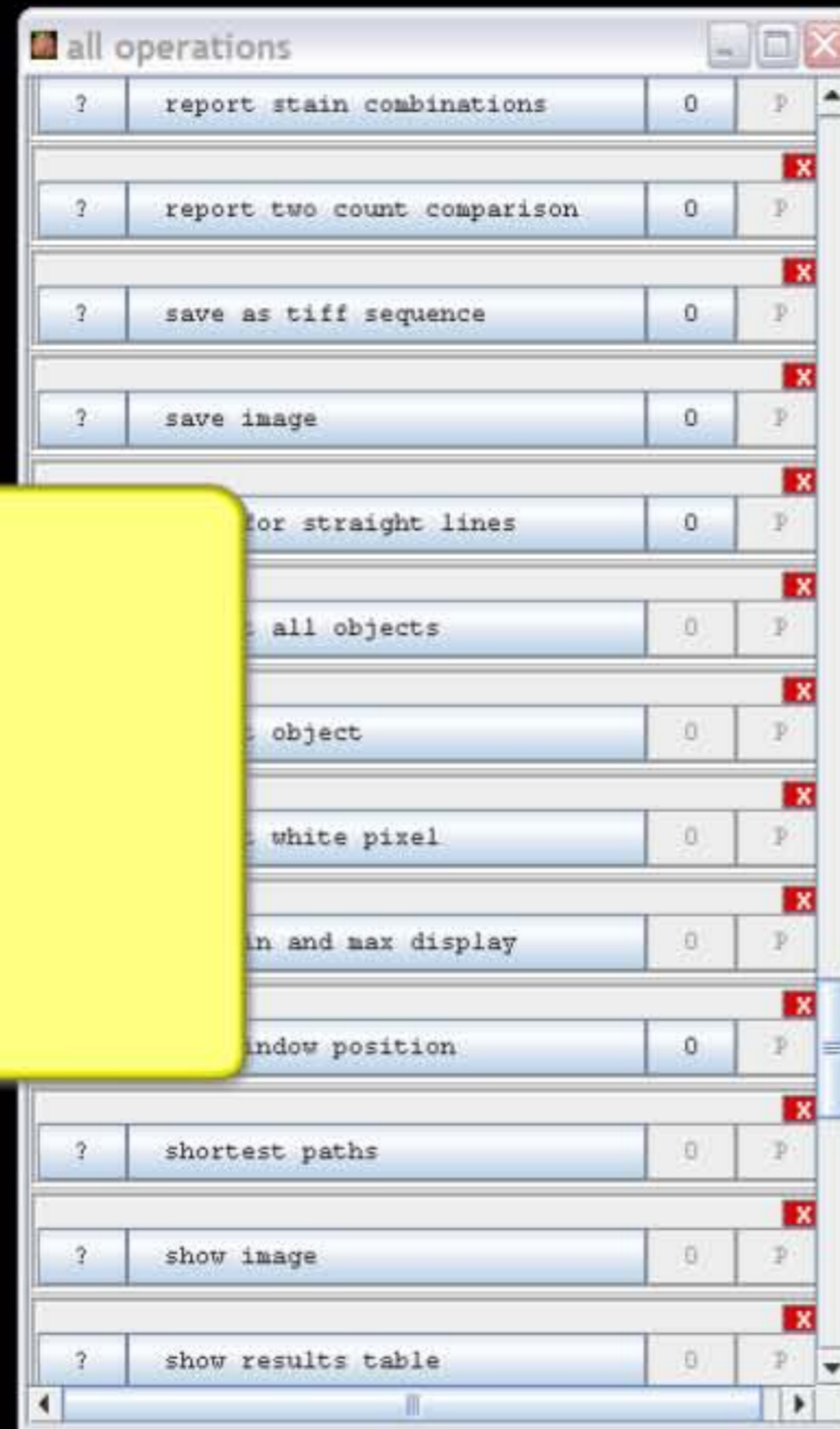
input parameter of save:
 - image
 - filename

enhance contrast 0 P

save image 0 P

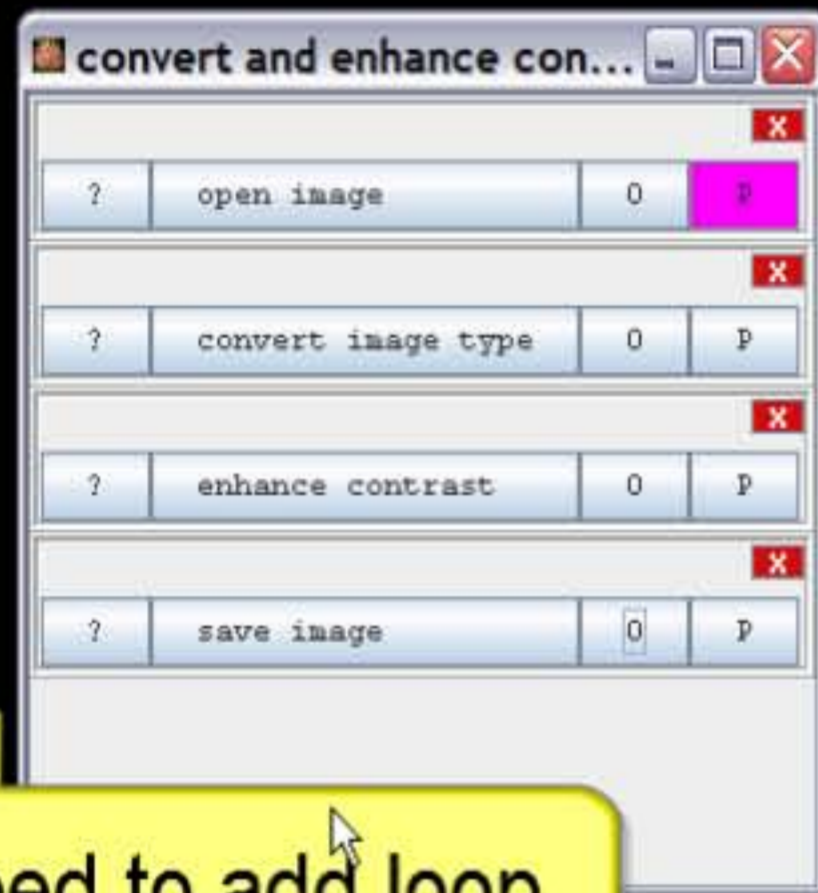
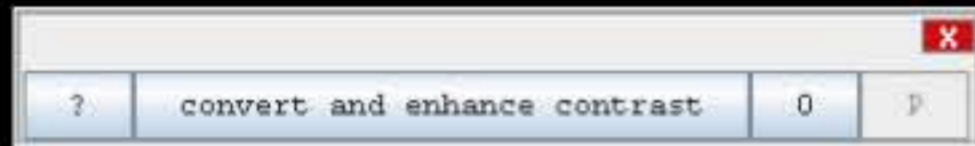
all operations

? report stain combinations 0 P
? report two count comparison 0 P
? save as tiff sequence 0 P
? save image 0 P
? scan for straight lines 0 P
? select all objects 0 P
? select object 0 P
? select white pixel 0 P
? set min and max display 0 P
? set window position 0 P
? shortest paths 0 P
? show image 0 P
? show results table 0 P



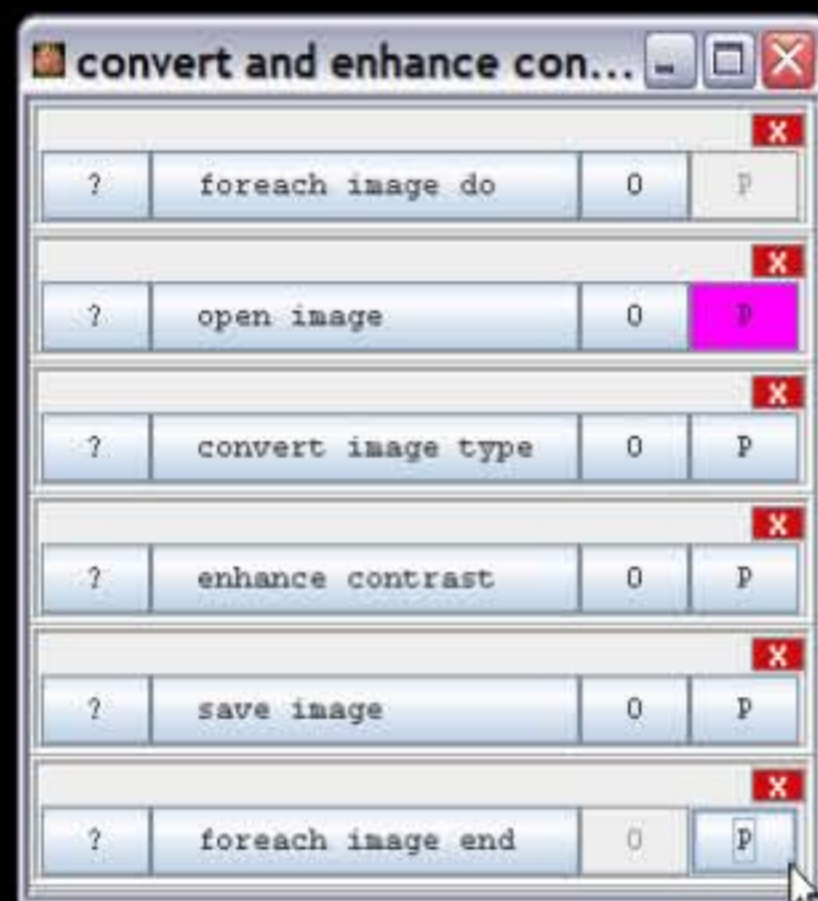
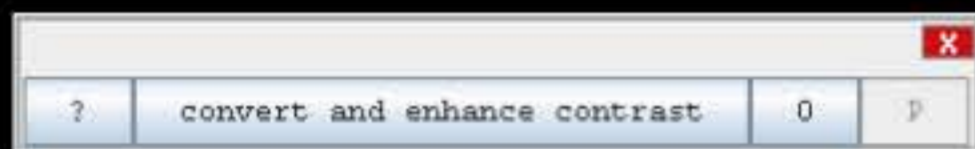
result location:

- absolute path
- path relative to input folder
- add string to filename
- add loop index to filename



need to add loop





parameters for open image

(String) path = null

parameter	from operation	output
path	foreach image do	current filename

apply

the input filename of open is the result from "forach image do"

?	open image	0	P
?	convert image type	0	P
?	enhance contrast	0	P
?	save image	0	P
?	foreach image end	0	P

all operations

?	and ends	0	P
?	and objects flood fill	0	P
?	find objects	0	P
?	find objects redirecting	0	P
?	find straight lines	0	P
?	foreach image do	0	P
?	foreach image end	0	P
?	foreach object do	0	P
?	foreach object end	0	P
?	gamma adjust	0	P
?	gaussian blur	0	P
?	get current image	0	P

parameters for open image

(String) path = foreach image do : CurrentFilename

parameter	from operation	output
path	foreach image do	current filename

apply

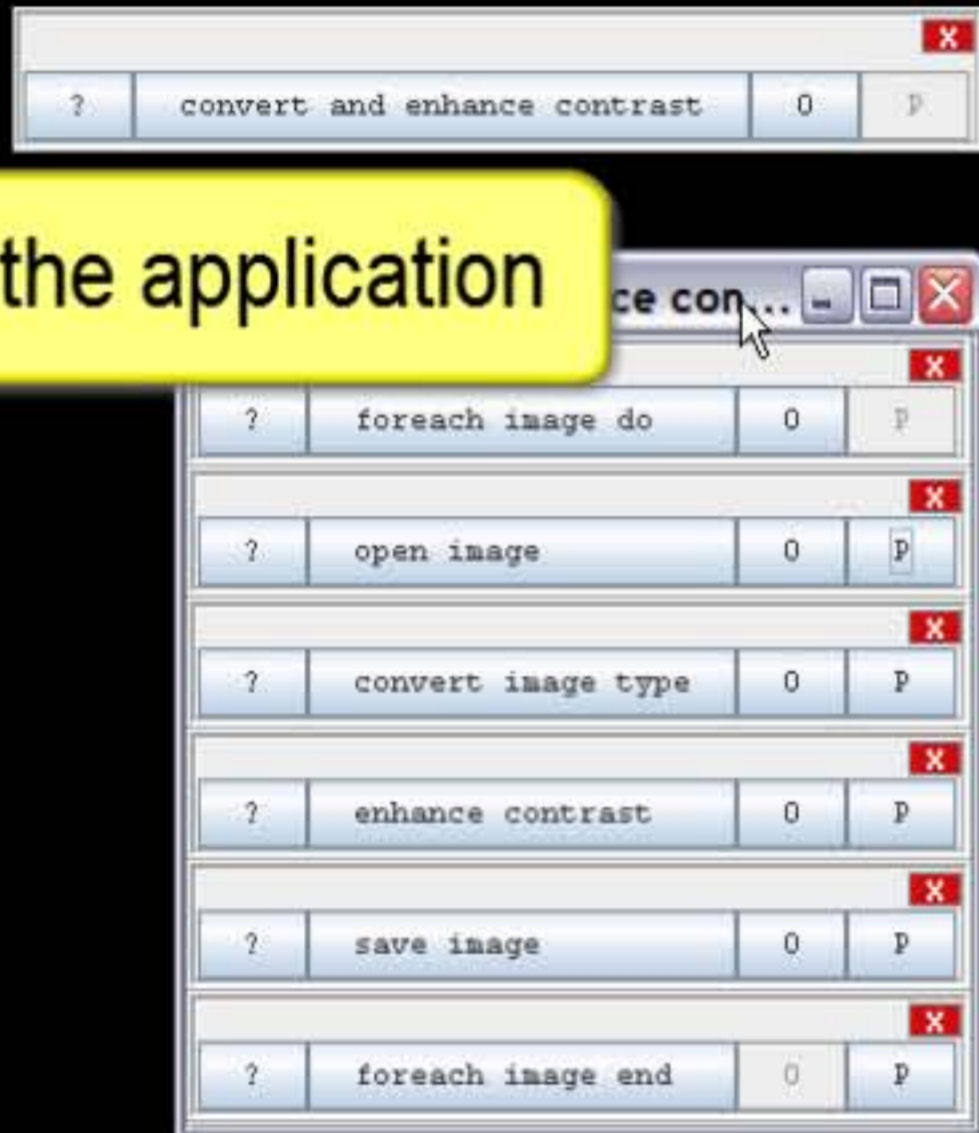
?	open image	0	P
?	convert image type	0	P
?	enhance contrast	0	P
?	save image	0	P
?	foreach image end	0	P

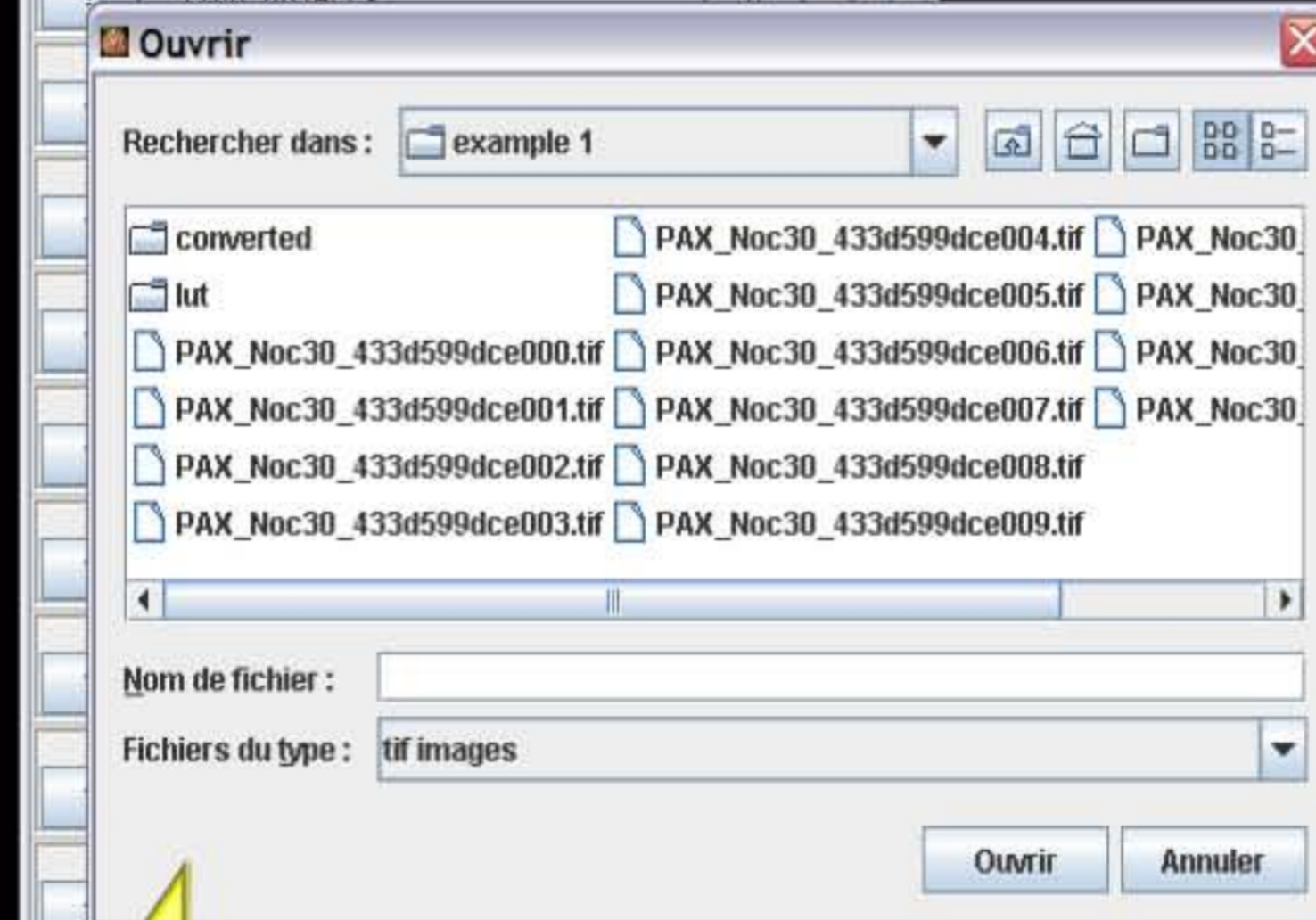
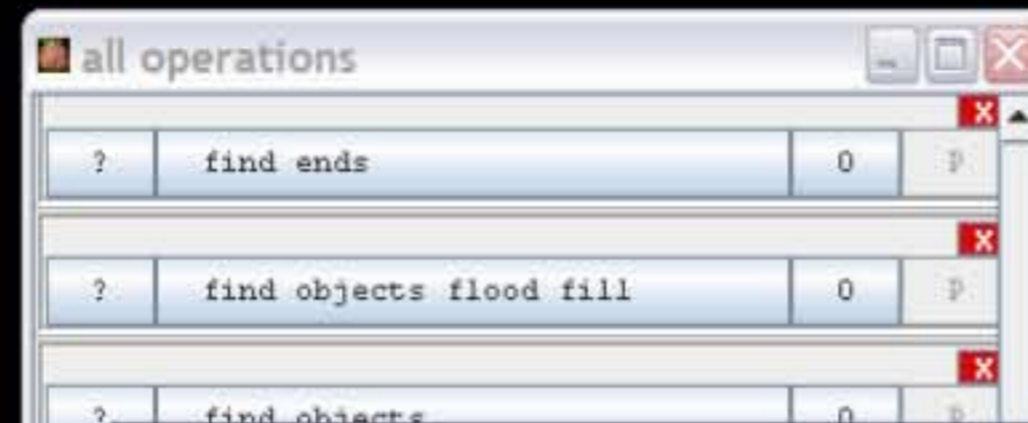
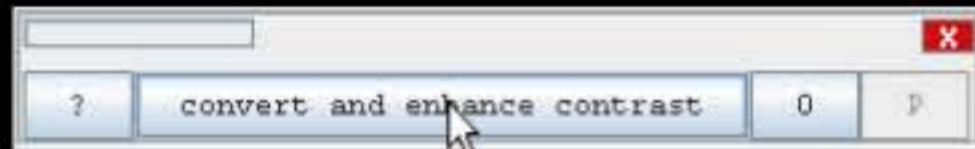
all operations

?	find ends	0	P
?	find objects flood fill	0	P
?	find objects	0	P
?	find objects redirecting	0	P
?	find straight lines	0	P
?	foreach image do	0	P
?	foreach image end	0	P
?	foreach object do	0	P
?	foreach object end	0	P
?	gamma adjust	0	P
?	gaussian blur	0	P
?	get current image	0	P



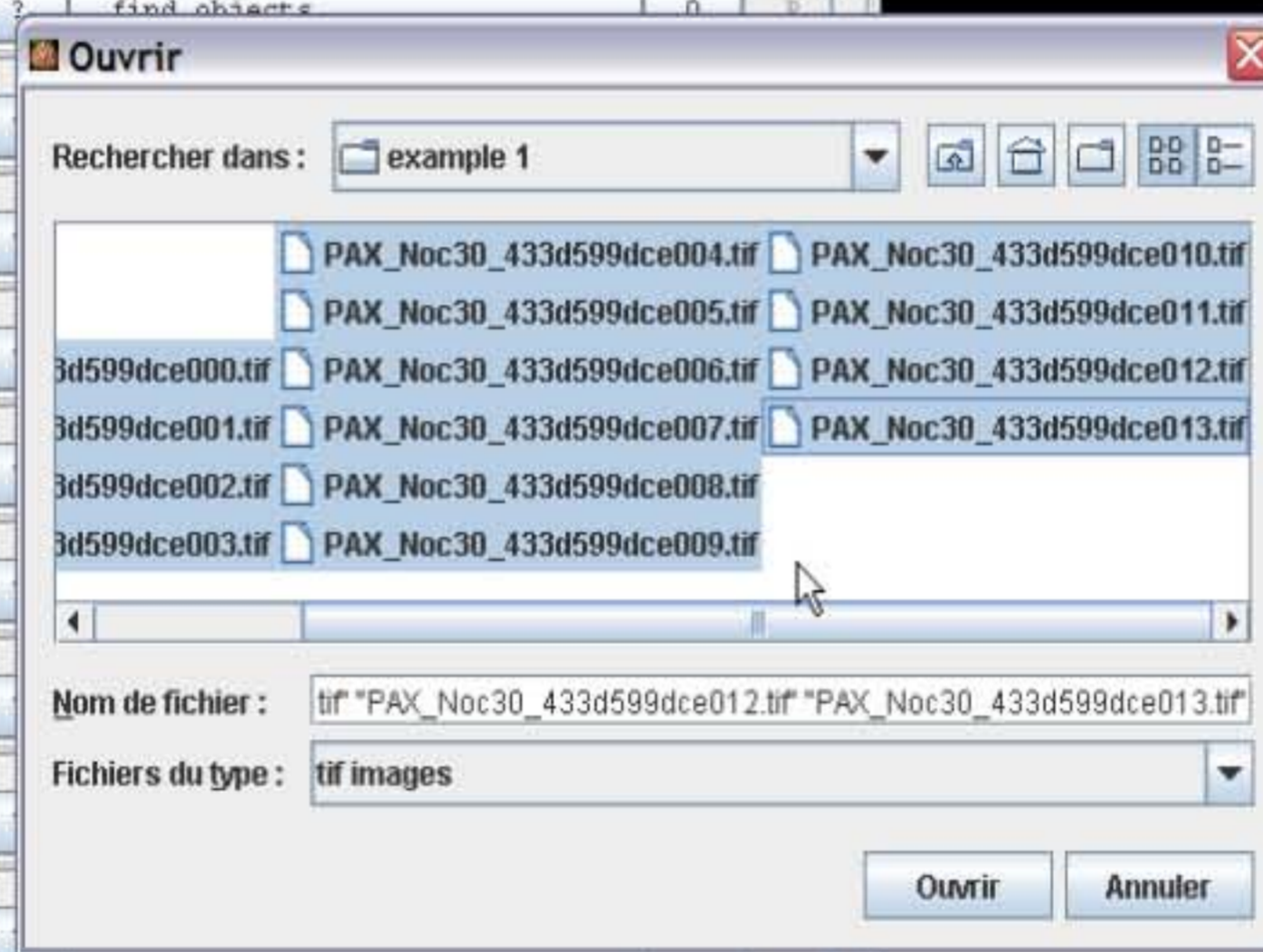
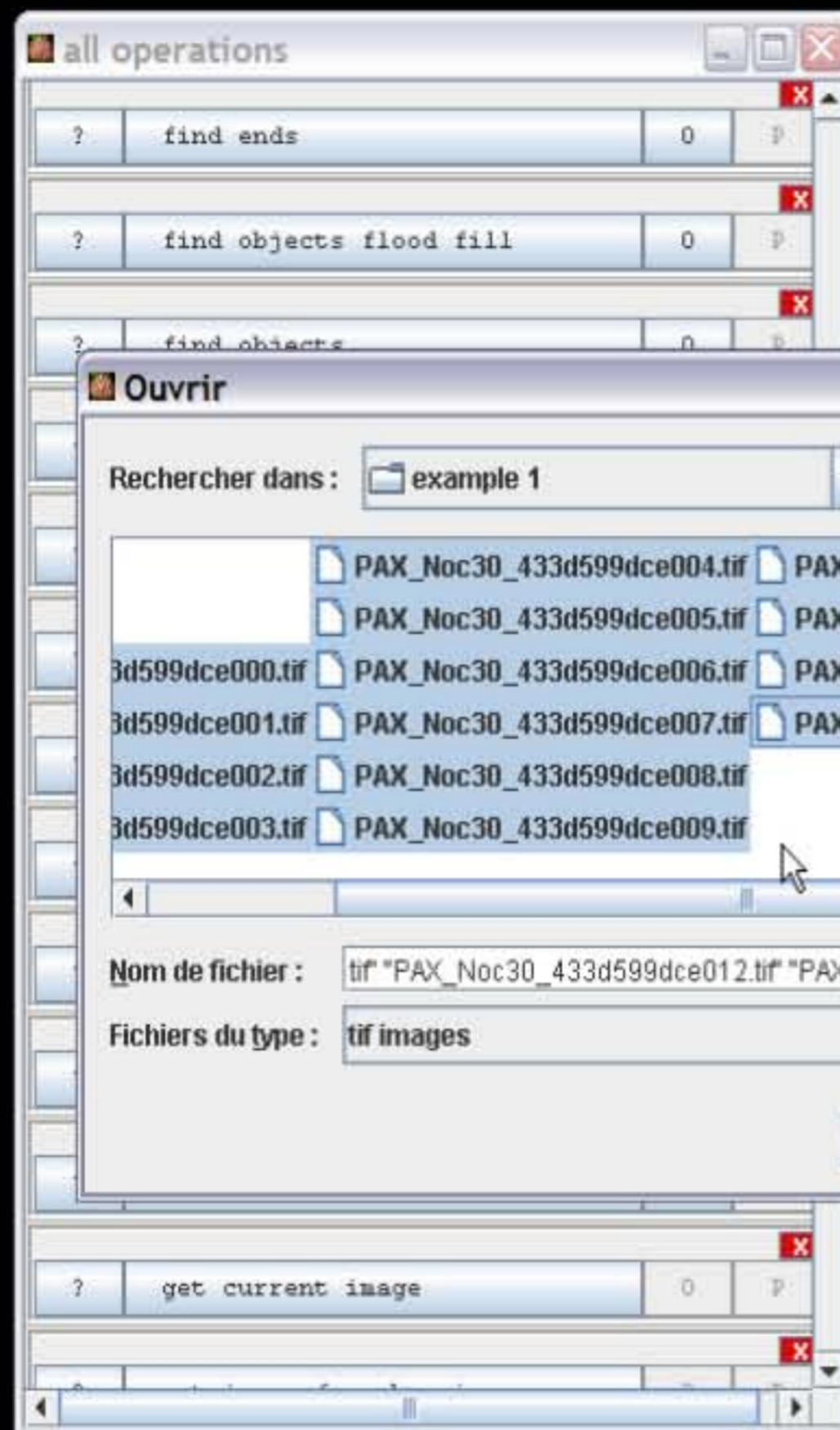
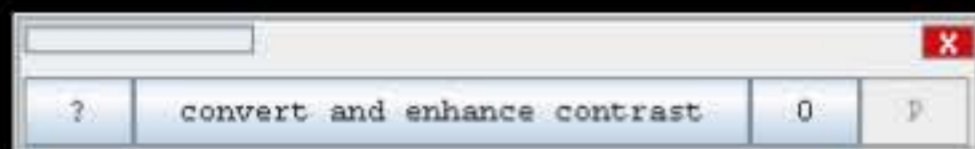
run the application

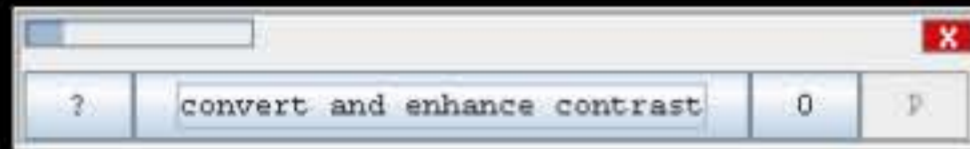




select

- a list of input images
 - all images are treated
- one folder
 - all images in the folder and in all sub-folders are treated





the application is running



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8bit > Spy LUT

0.093 seconds, 14.4 million pixels/second

convert and enhance contrast

0 P

convert and enhance con...

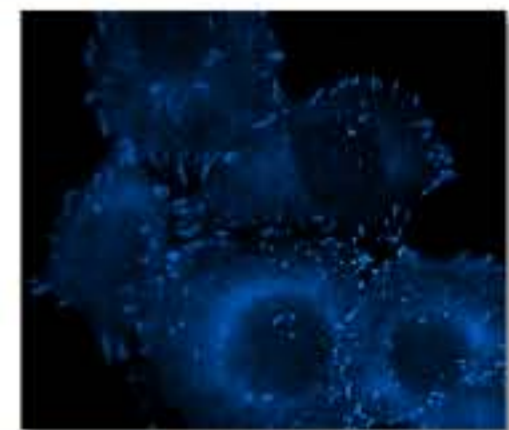
?	foreach image do	0	P
?	open image	0	P
?	convert image type	0	P
?	enhance contrast	0	P
?	save image	0	P
?	foreach image end	0	P

all operations

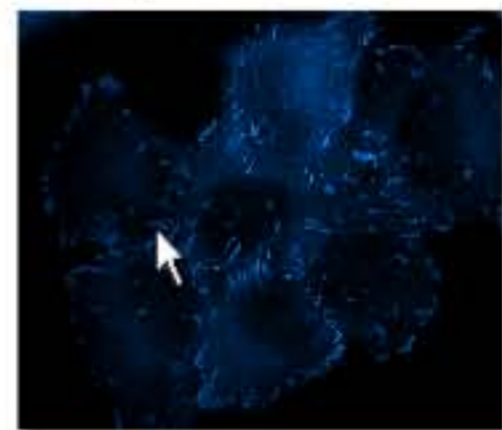
?	find ends	0	P
?	find objects flood fill	0	P
?	find objects	0	P
?	find objects redirecting	0	P
?	find straight lines	0	P
?	foreach image do	0	P
?	foreach image end	0	P
?	foreach object do	0	P
?	foreach object end	0	P
?	gamma adjust	0	P
?	gaussian blur	0	P
?	get current image	0	P



C:\Documents and Settings\Volker Bäcker\Mes documents\images\example 1\converted\



PAX_Noc30_433d599dce000.tif



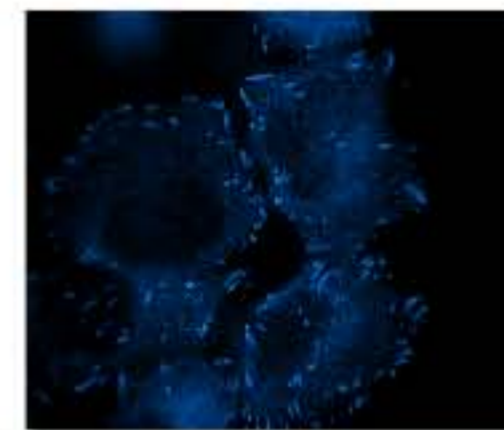
PAX_Noc30_433d599dce001.tif



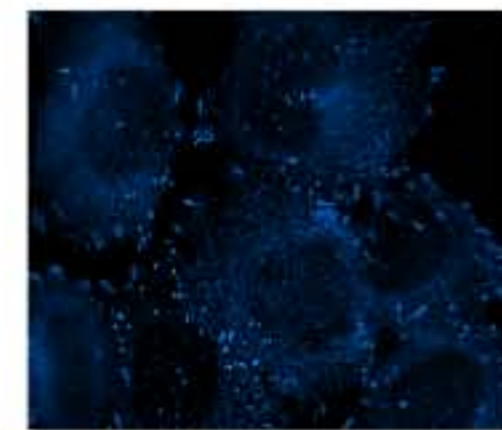
PAX_Noc30_433d599dce002.tif



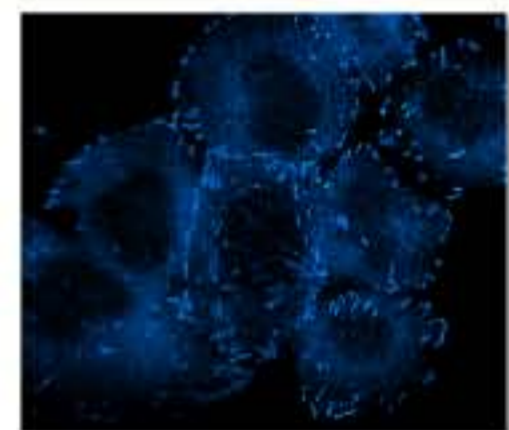
PAX_Noc30_433d599dce003.tif



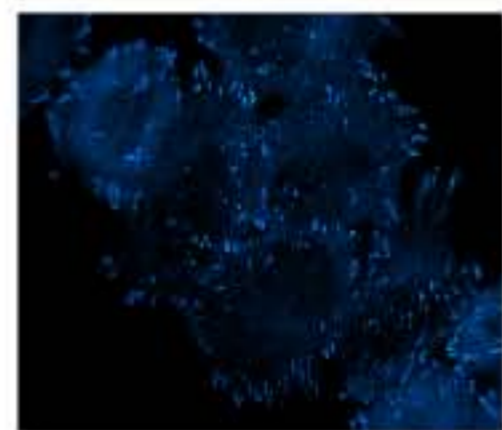
PAX_Noc30_433d599dce004.tif



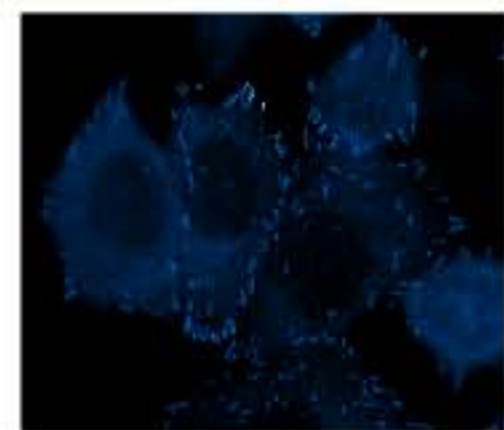
PAX_Noc30_433d599dce005.tif



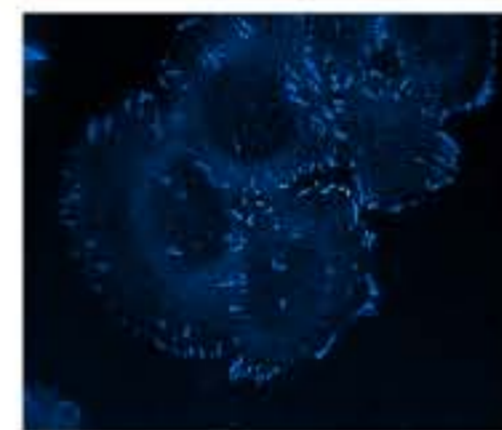
PAX_Noc30_433d599dce006.tif



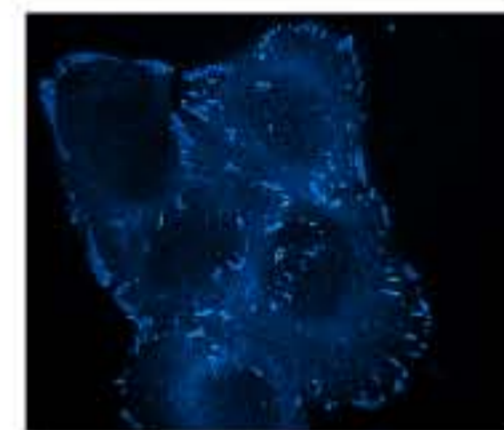
PAX_Noc30_433d599dce007.tif



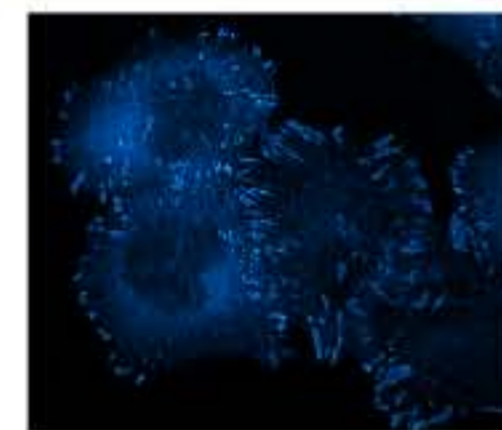
PAX_Noc30_433d599dce008.tif



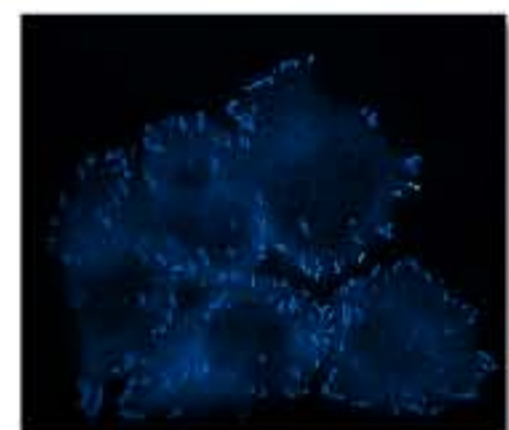
PAX_Noc30_433d599dce009.tif



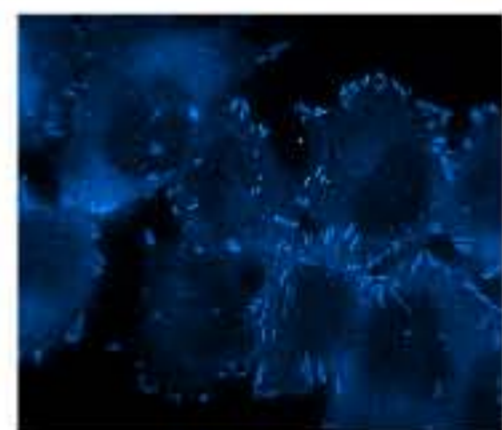
PAX_Noc30_433d599dce010.tif



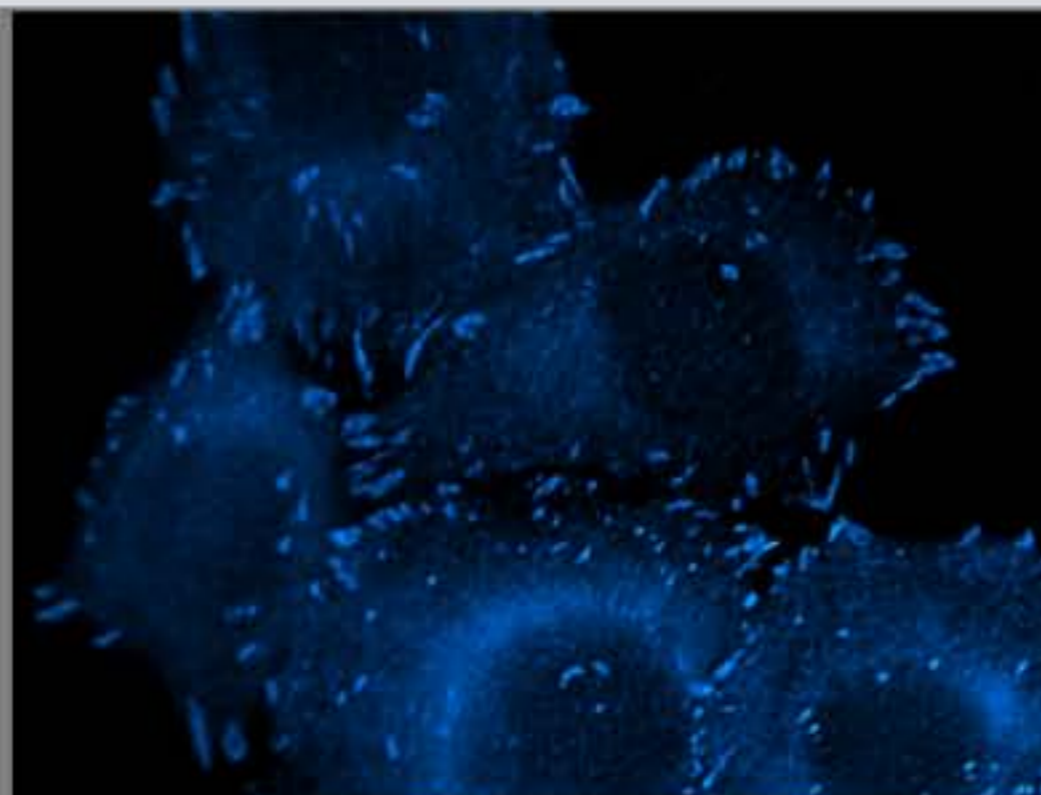
PAX_Noc30_433d599dce011.tif

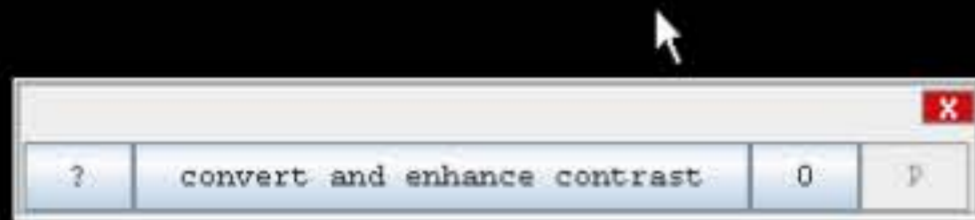


PAX_Noc30_433d599dce012.tif



PAX_Noc30_433d599dce013.tif

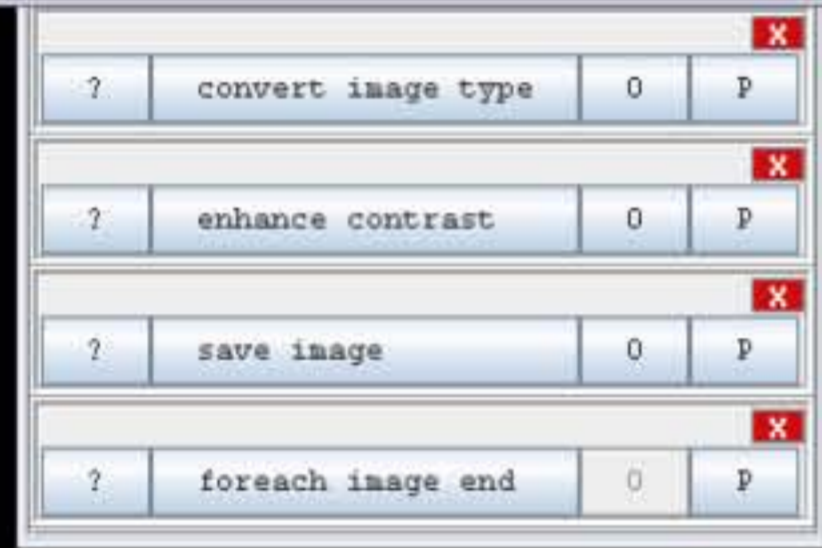
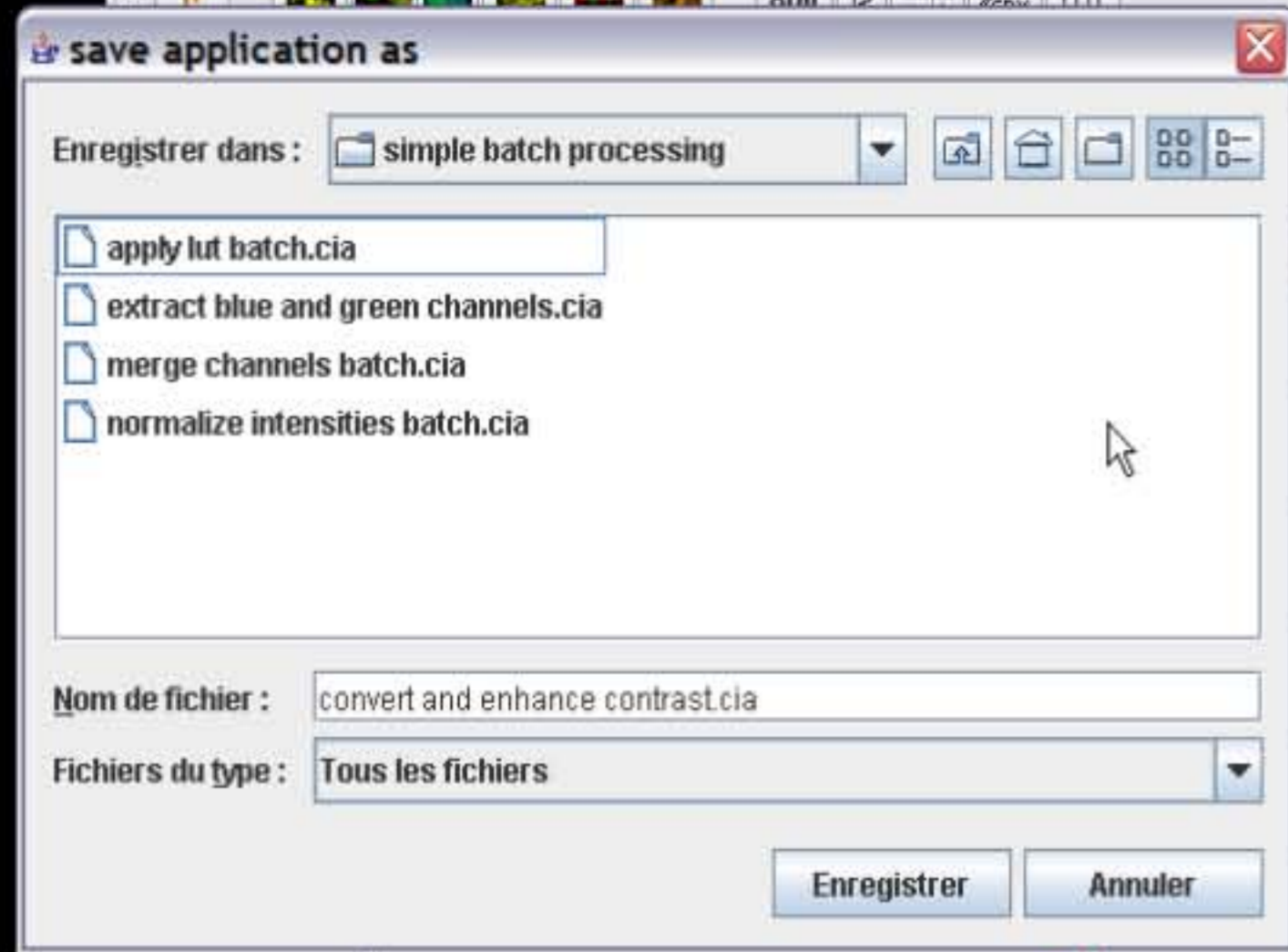


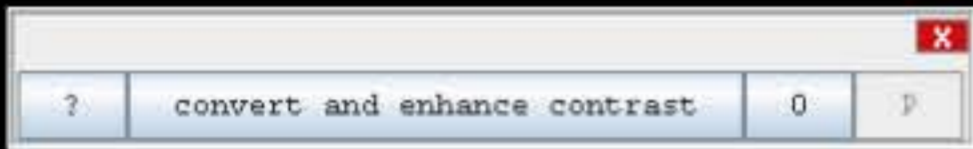
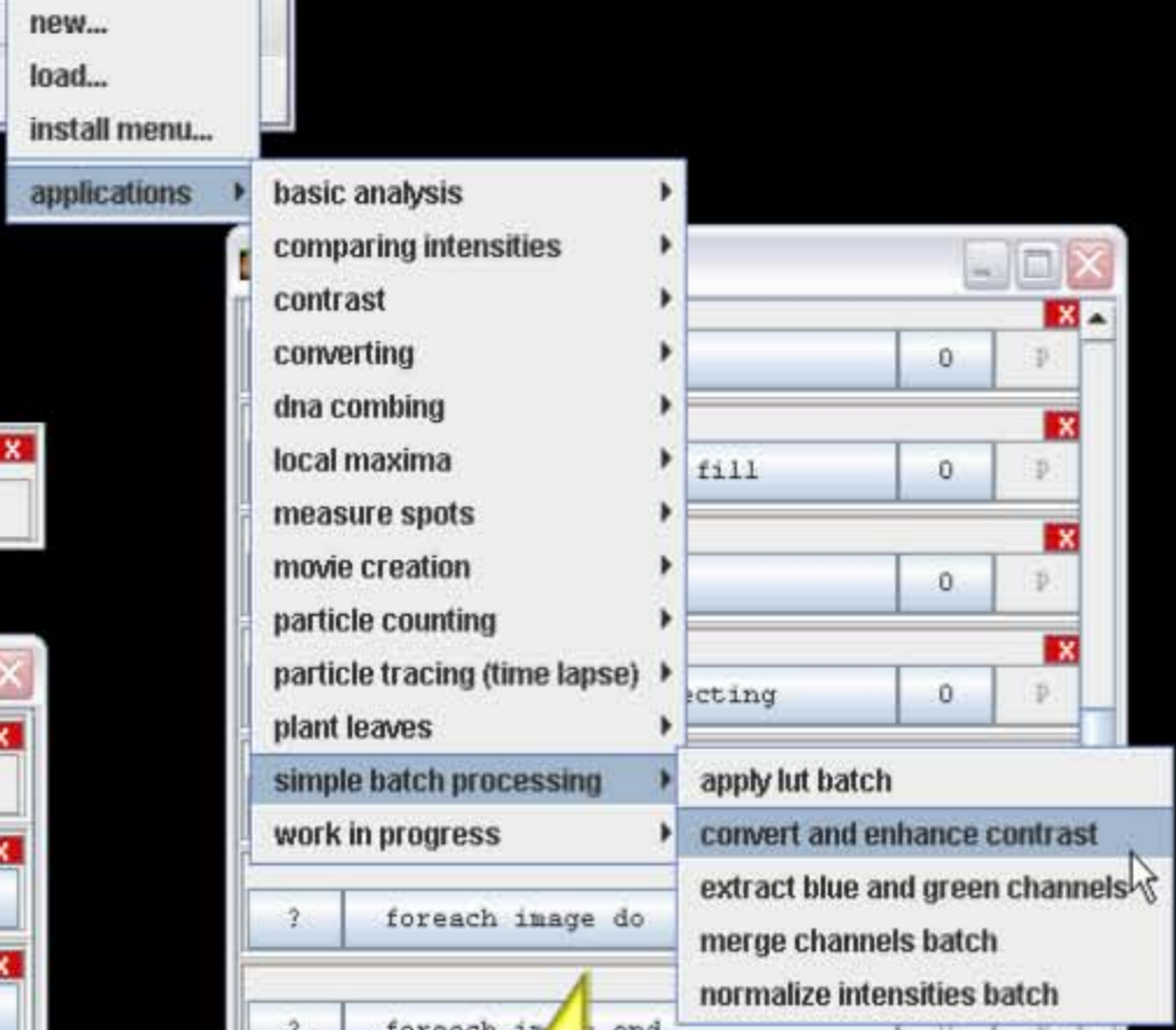


save the application





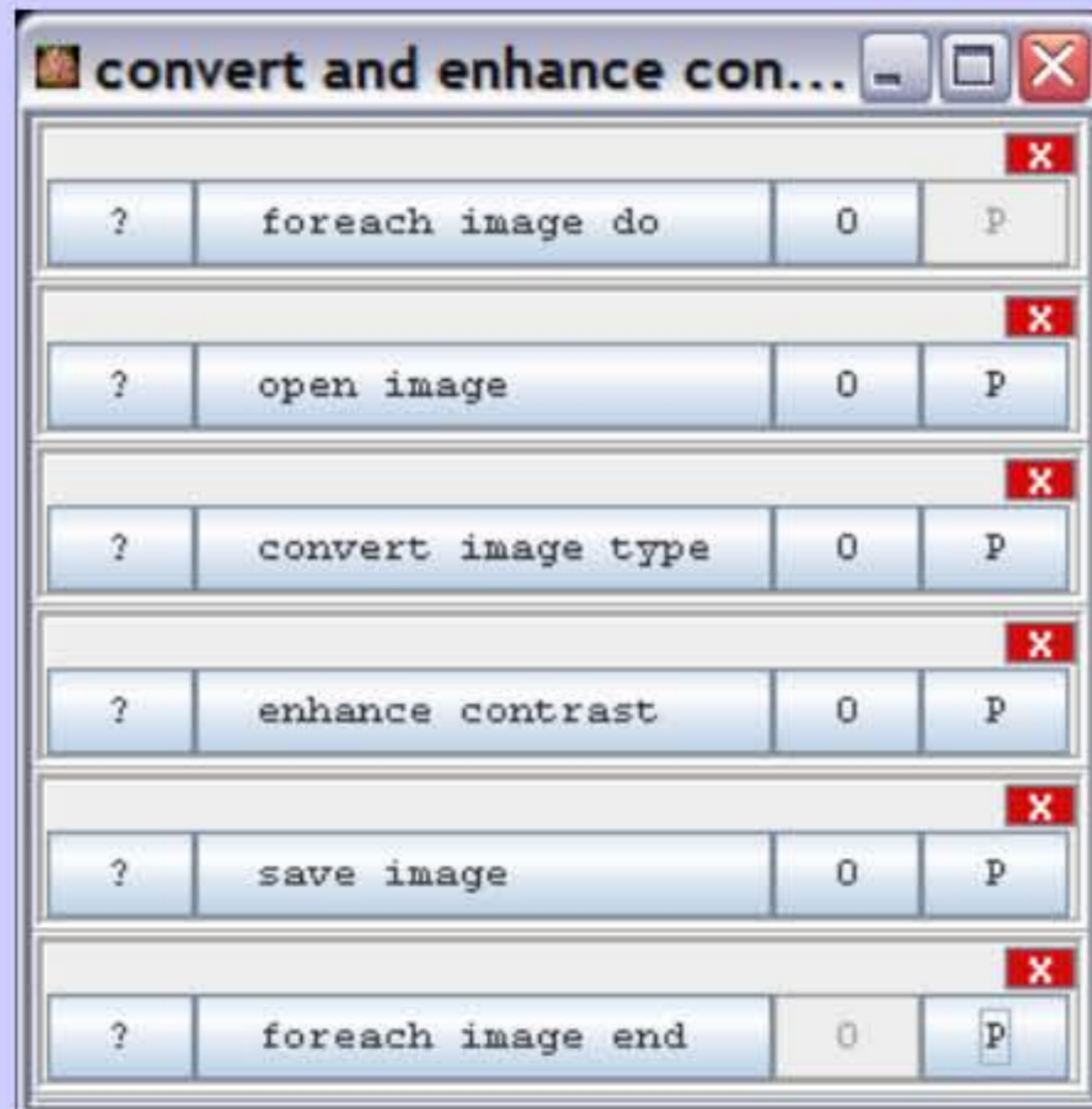




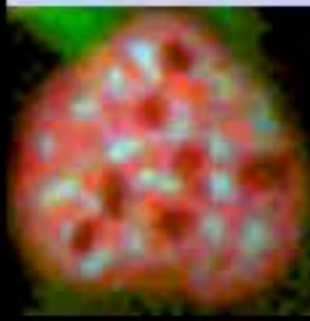
the application can be opened from the applications menu



Comparison visual script / macro

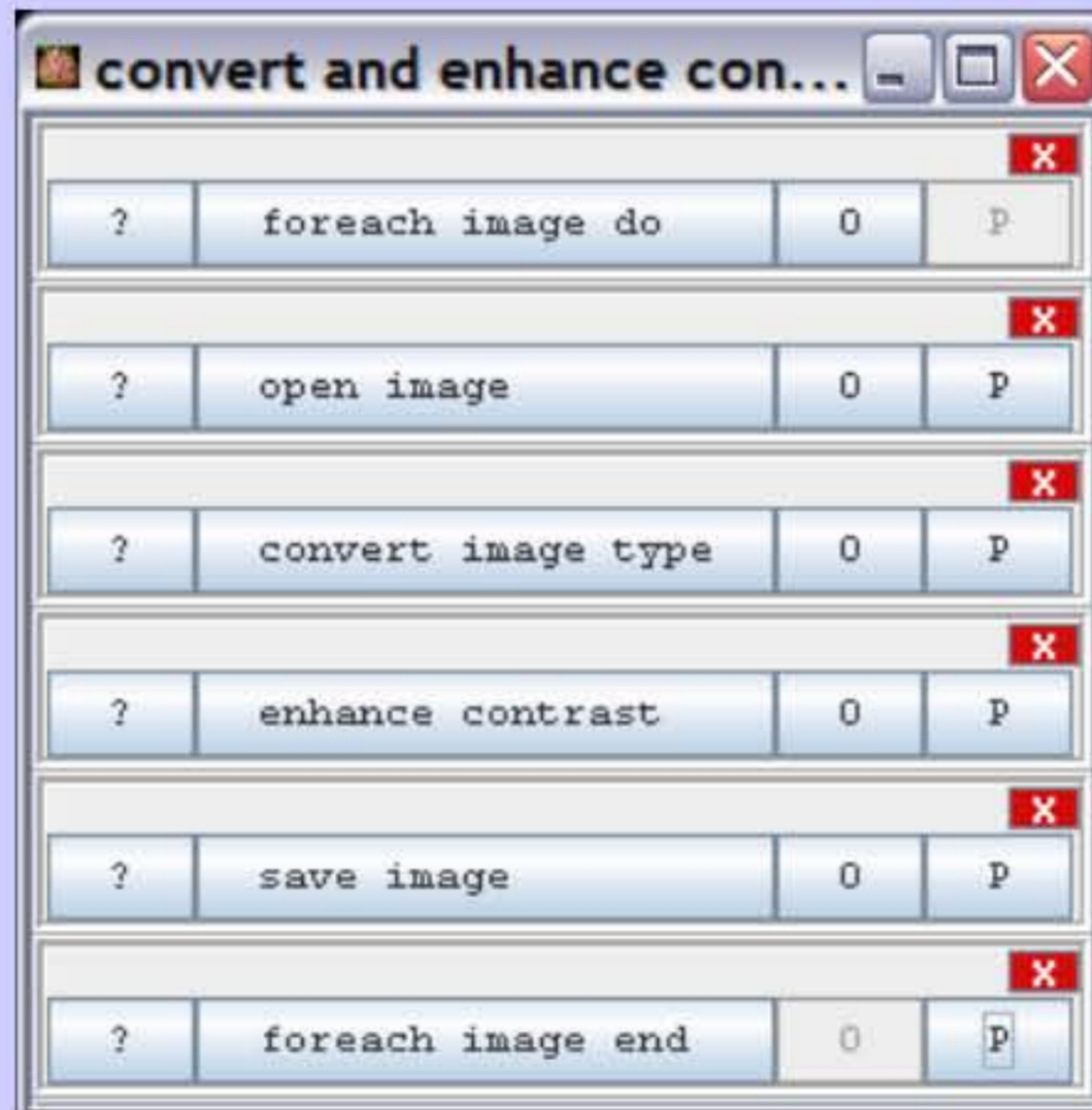


```
dir1 = getDirectory("Choose Source Directory ");
dir2 = getDirectory("Choose Destination Directory ");
list = getFileList(dir1);
Dialog.create("enhance contrast options");
Dialog.addNumber("percent saturated", 0.5);
Dialog.addCheckbox("equalize", false);
Dialog.addCheckbox("normalize", true);
Dialog.addCheckbox("use stack histogram", false);
Dialog.show();
saturated=Dialog.getNumber();
equalize = Dialog.getCheckbox();
normalize = Dialog.getCheckbox();
useStackHistogram = equalize = Dialog.getCheckbox();
setBatchMode(true);
for (i=0; i<list.length; i++) {
    showProgress(i+1, list.length);
    if (endsWith(list[i], ".tif")) {
        open(dir1+list[i]);
        run("8-bit");
        run("Enhance Contrast", "saturated=" + saturated + "
normalize");
        saveAs("Tiff", dir2+list[i]);
    }
}
```



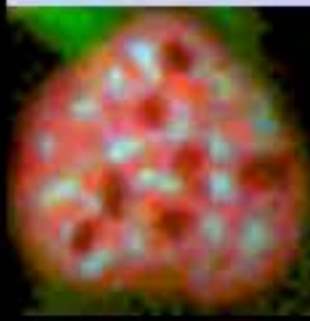


Comparison visual script / macro



```
dir1 = getDirectory("Choose Source Directory ");  
dir2 = getDirectory("Choose Destination Directory ");  
list = getFileList(dir1);
```

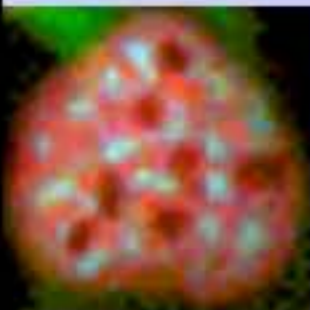
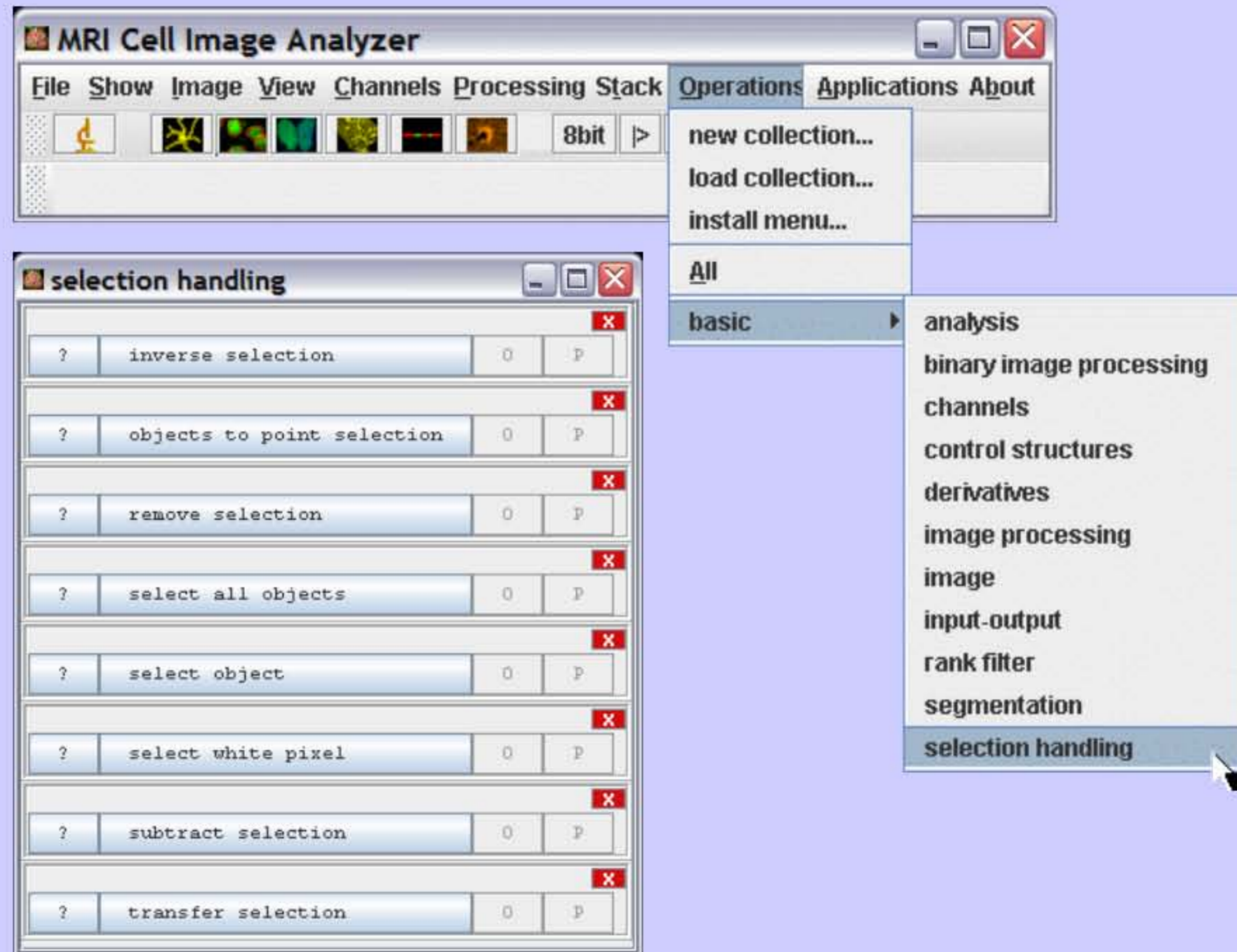
```
setBatchMode(true);  
for (i=0; i<list.length; i++) {  
    showProgress(i+1, list.length);  
    if (endsWith(list[i], ".tif")) {  
        open(dir1+list[i]);  
        run("8-bit");  
        run("Enhance Contrast", "saturated=" + saturated + "  
normalize");  
        saveAs("Tiff", dir2+list[i]);  
    }  
}
```





Operation collections

- processing
- analysis
- control structures
- selection handling
- input / output
- reporting



Operation collections

- processing
- analysis
- control structures

foreach image do
 foreach object do
 accept or skip or exit
 wait for user
 skip no objects found
 skip saturated
 ...
end

The screenshot shows the 'MRI Cell Image Analyzer' window with the 'Operations' menu open. The menu items are: new collection..., load collection..., install menu..., All, basic, analysis, binary image processing, channels, control structures, derivatives, image processing, image, input-output, rank filter, segmentation, and selection handling. The 'selection handling' dialog box is also open, displaying a list of operations with checkboxes for 'O' and 'P'.

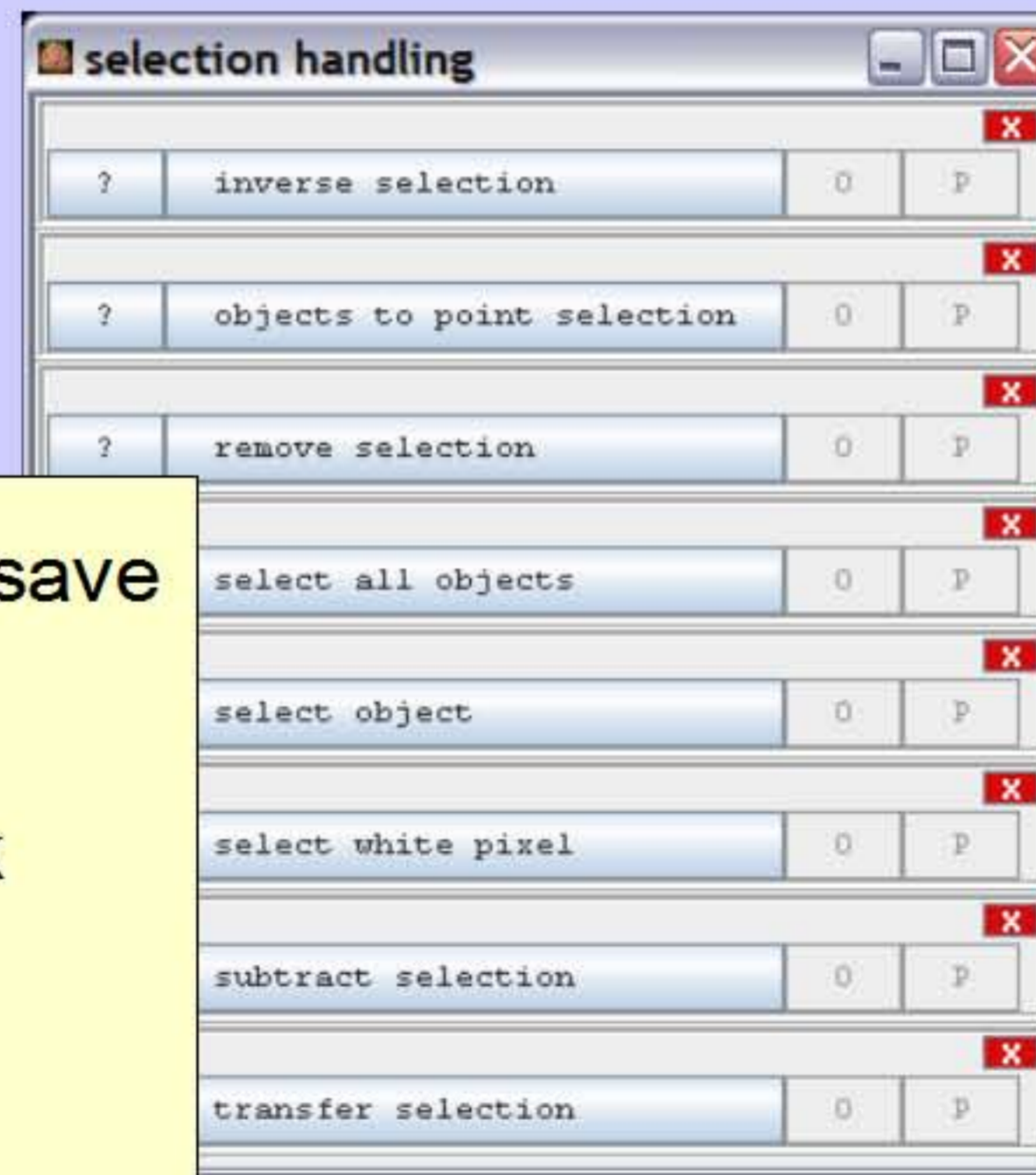
Operation	O	P
inverse selection	<input type="checkbox"/>	<input type="checkbox"/>
objects to point selection	<input type="checkbox"/>	<input type="checkbox"/>
remove selection	<input type="checkbox"/>	<input type="checkbox"/>
select all objects	<input type="checkbox"/>	<input type="checkbox"/>
select object	<input type="checkbox"/>	<input type="checkbox"/>
select white pixel	<input type="checkbox"/>	<input type="checkbox"/>
subtract selection	<input type="checkbox"/>	<input type="checkbox"/>
transfer selection	<input type="checkbox"/>	<input type="checkbox"/>



Operation collections

- processing
- analysis
- control structures
- selection handling
- input / output

open / show / hide / save
/ close image
show results table
open series as stack
save as tiff series

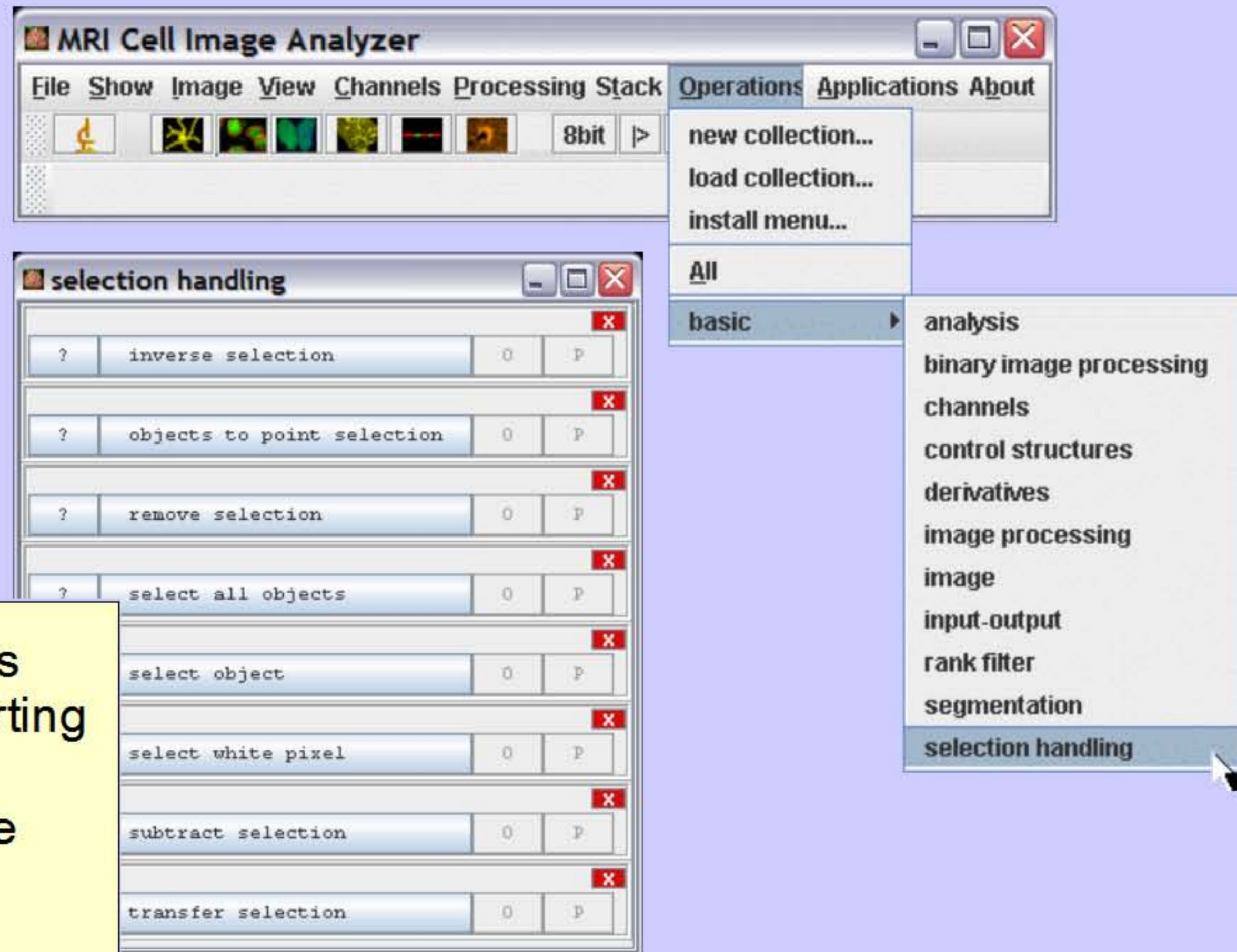


- analysis
- binary image processing
- channels
- control structures
- derivatives
- image processing
- image
- input-output
- rank filter
- segmentation
- selection handling

Operation collections

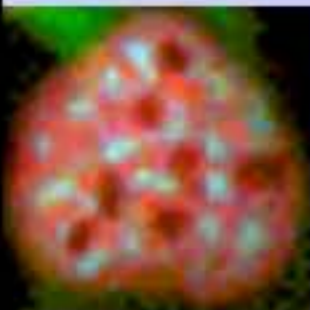
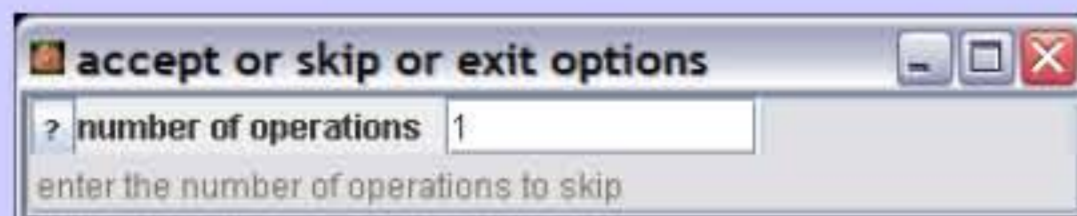
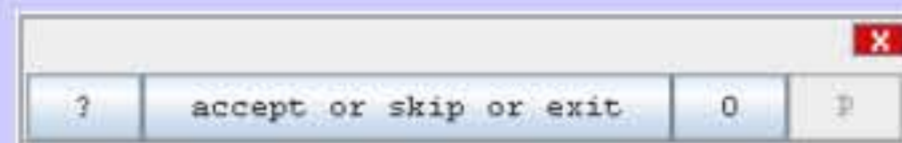
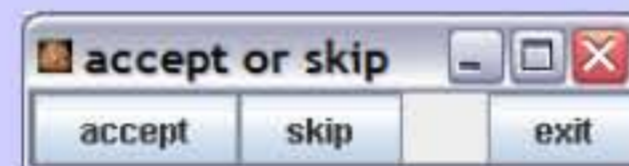
- processing
- analysis
- control structures
- selection handling
- input / output
- reporting

report measurements
project specific reporting
operations
write spreadsheet file

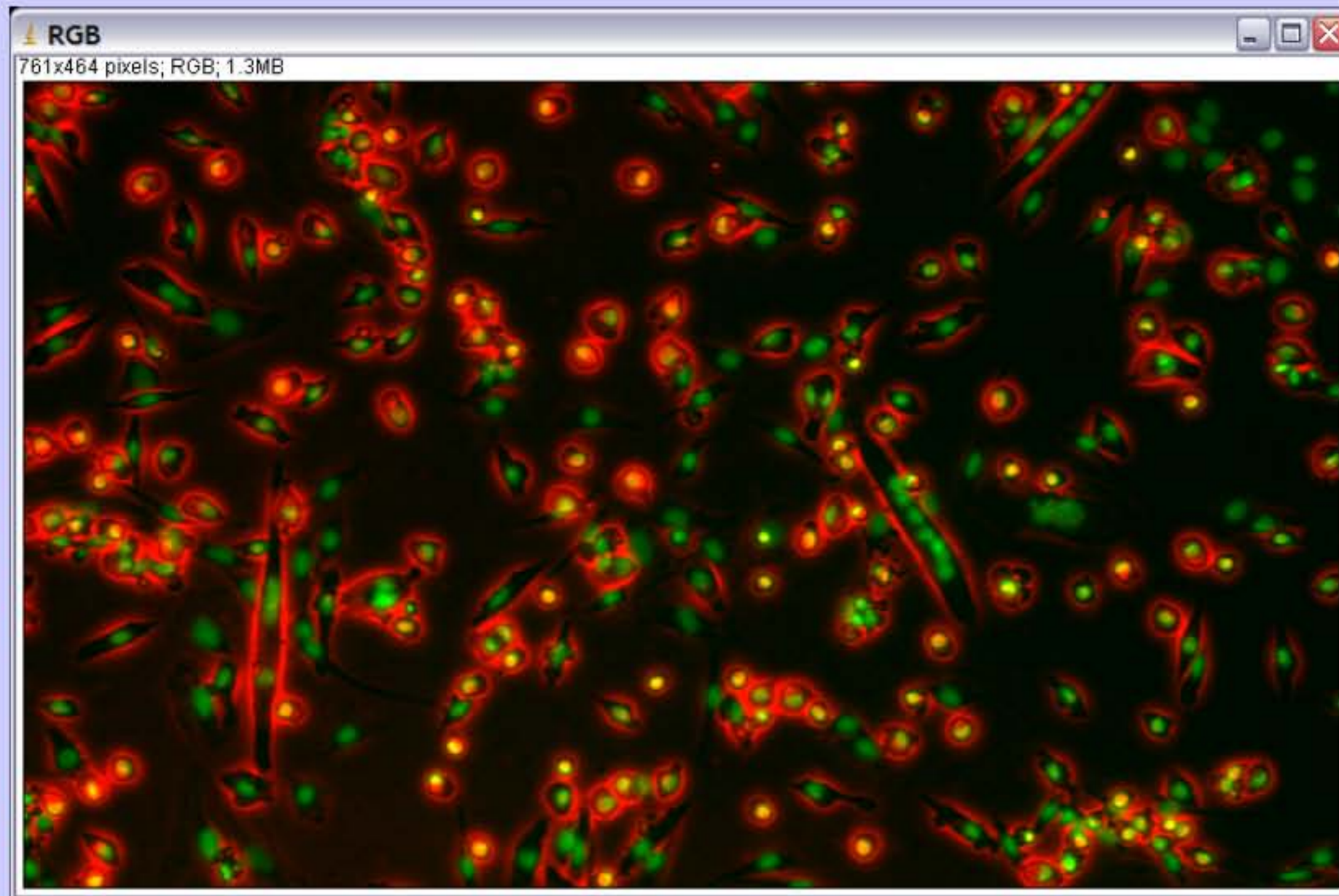


Interactive applications

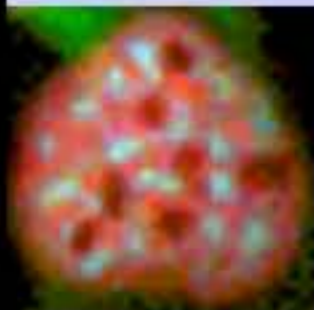
- let the computer do the work when possible
- the user take decisions when necessary
- execution of application stops
- user decides to take result into account
- user corrects automatic result
 - selection
 - mask

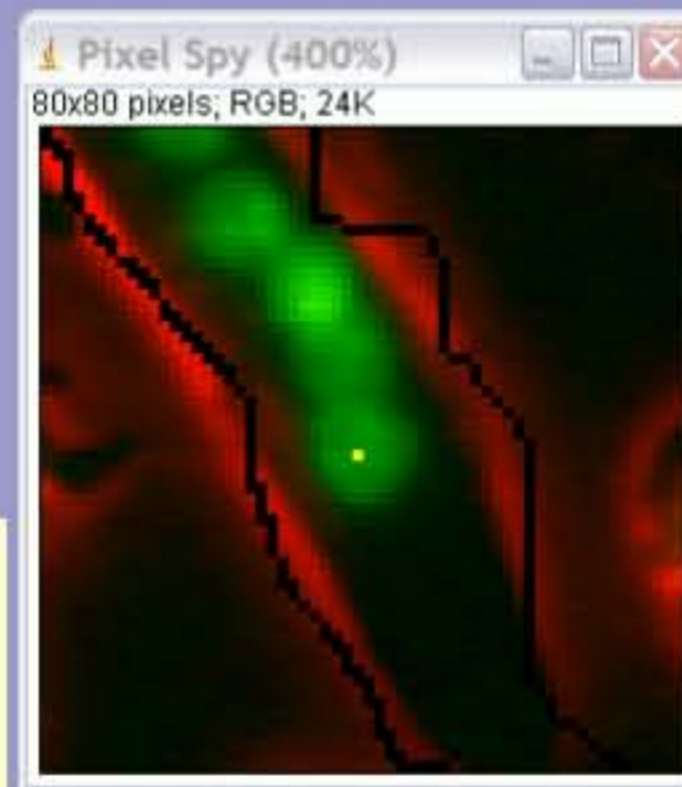
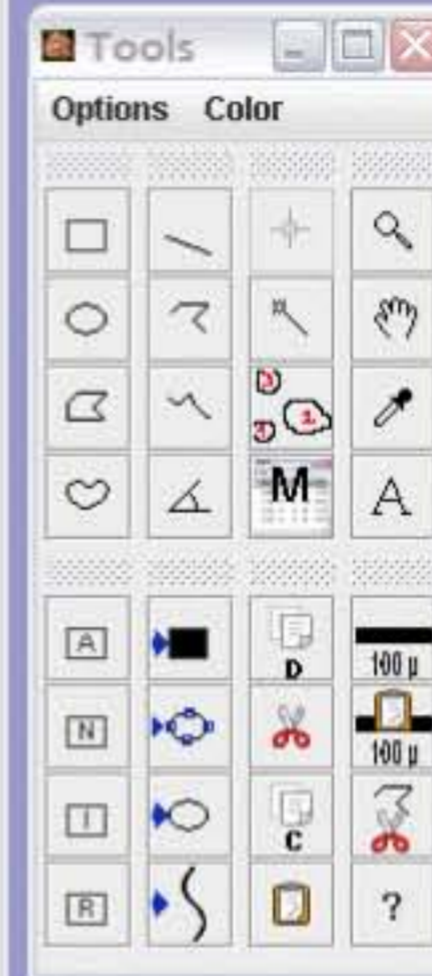
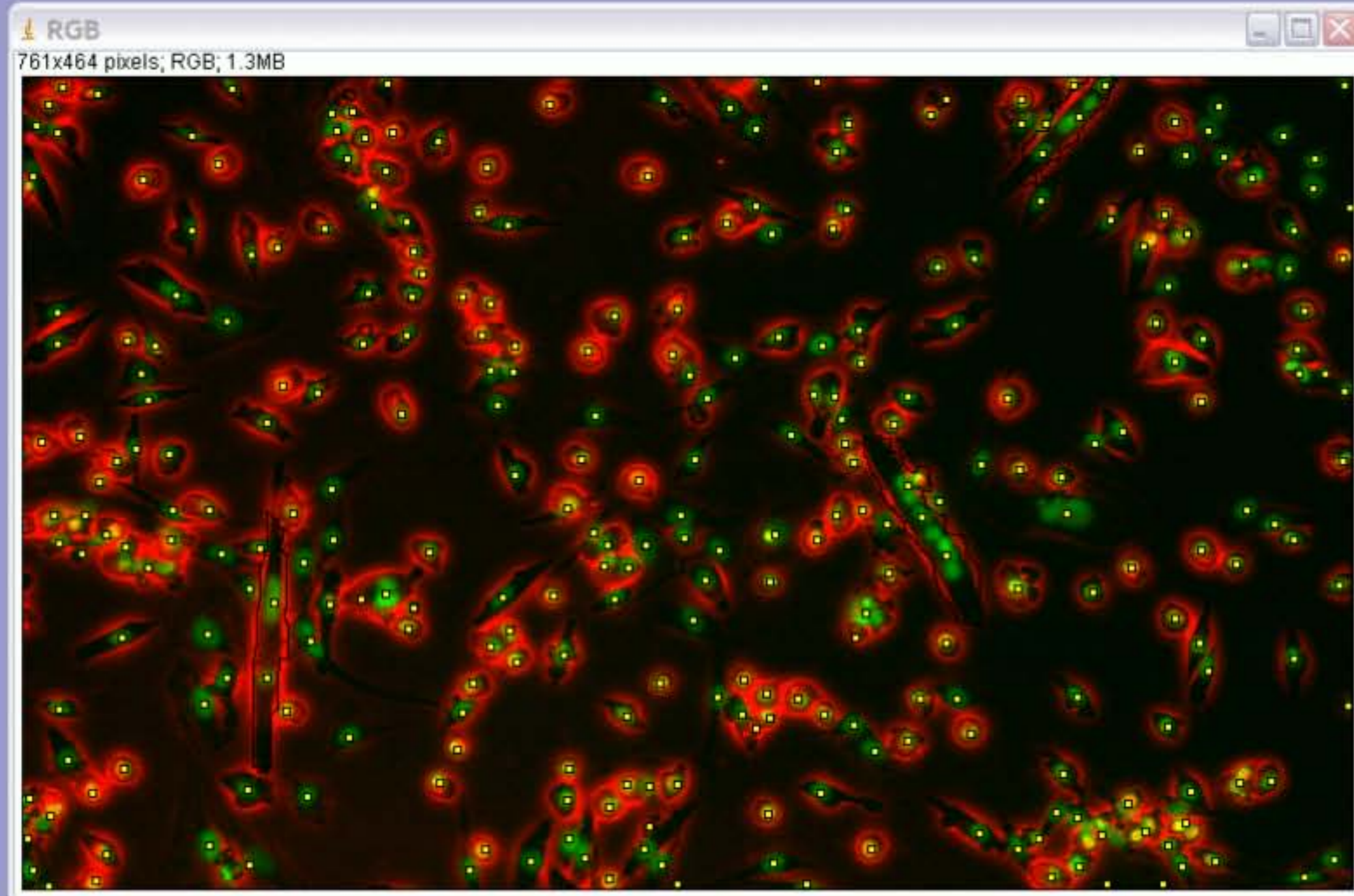
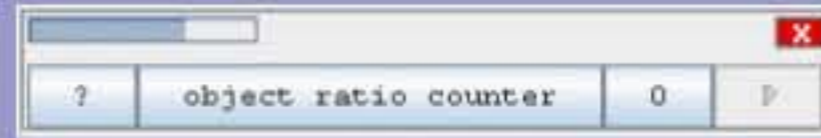
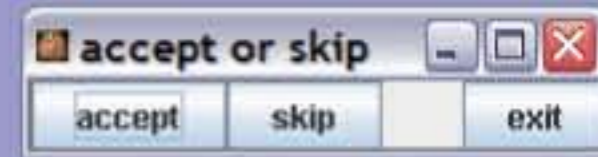


Interactive applications



- count nuclei
 - all
 - inside certain structures
- calculate the ratio

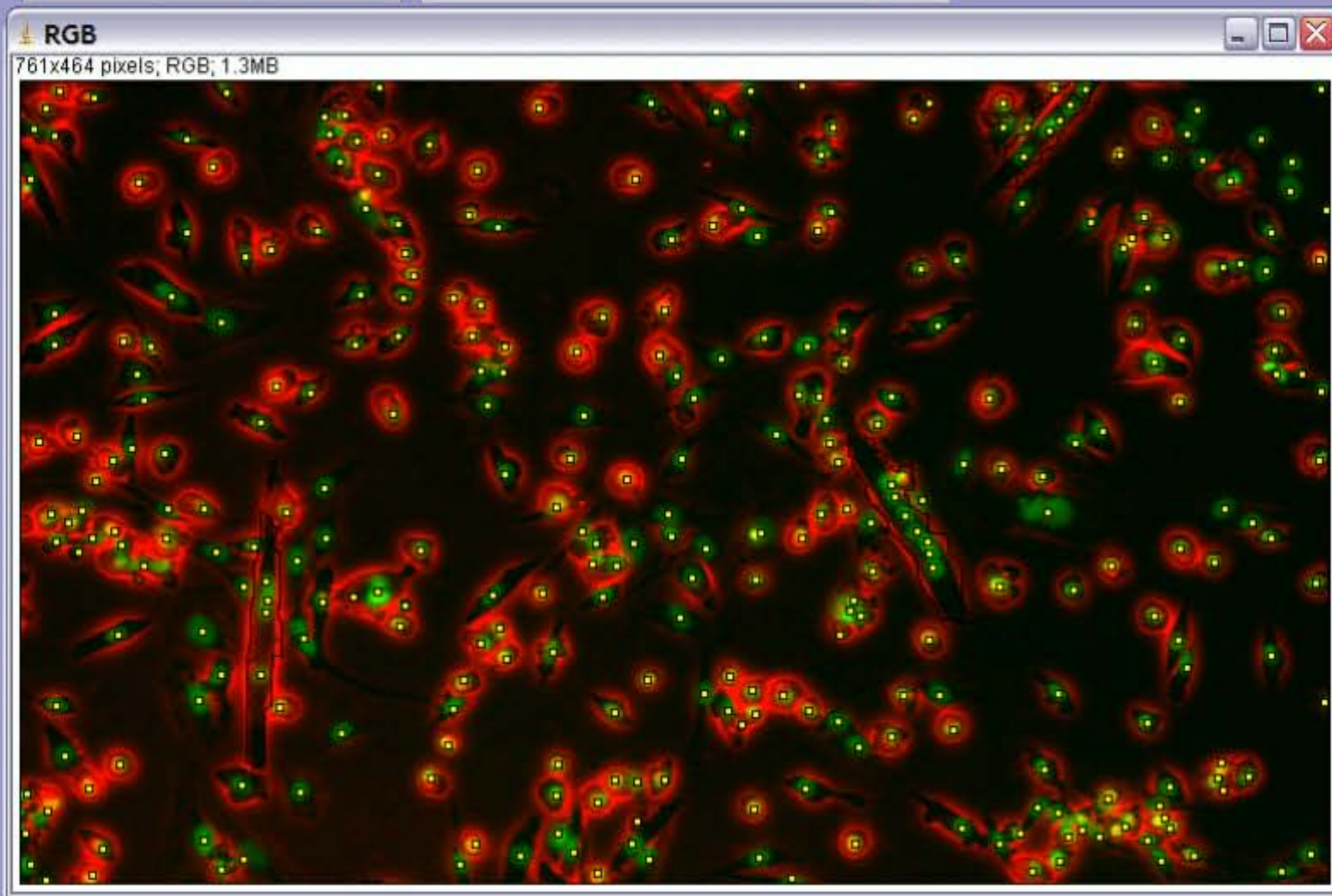




- application selects nuclei
- user corrects selection
- user presses accept to continue

accept or skip object ratio counter

accept skip exit ? object ratio counter 0 ▶



Tools

Options Color

□ / + 🔍
○ ↶ ✎ 🖱
📄 ~ 🗑 🖱
❤ ▲ M A
A ■ 📄 100 μ
N 🔍 ✂ 100 μ
T 🔍 C ✂
R 📄 ?

object ratio counter

? merge channels	0	P
? show image	0	P
? accept or skip or exit	0	P
? transfer selection	0	P
? draw	0	P
? subtract background	0	P
? find and subtract background	0	P
? local threshold	0	P
? convert image type	0	P
? mean threshold	0	P
? invert image	0	P
? find objects	0	P
? objects to point selection	0	P
? accept or skip or exit	0	P
? draw	0	P
? save image	0	P
? measure	0	P

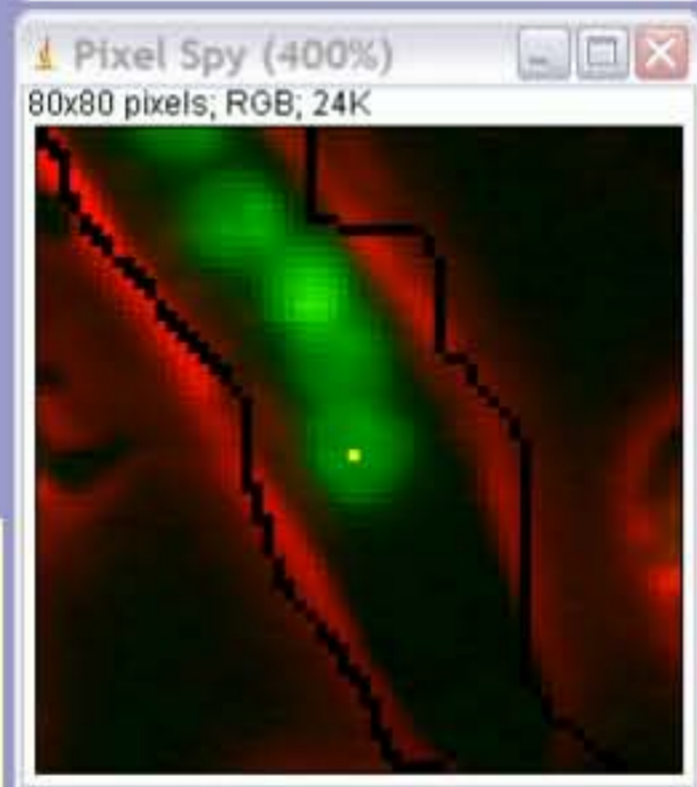
Channel Mixer

mix: min keep adjustment

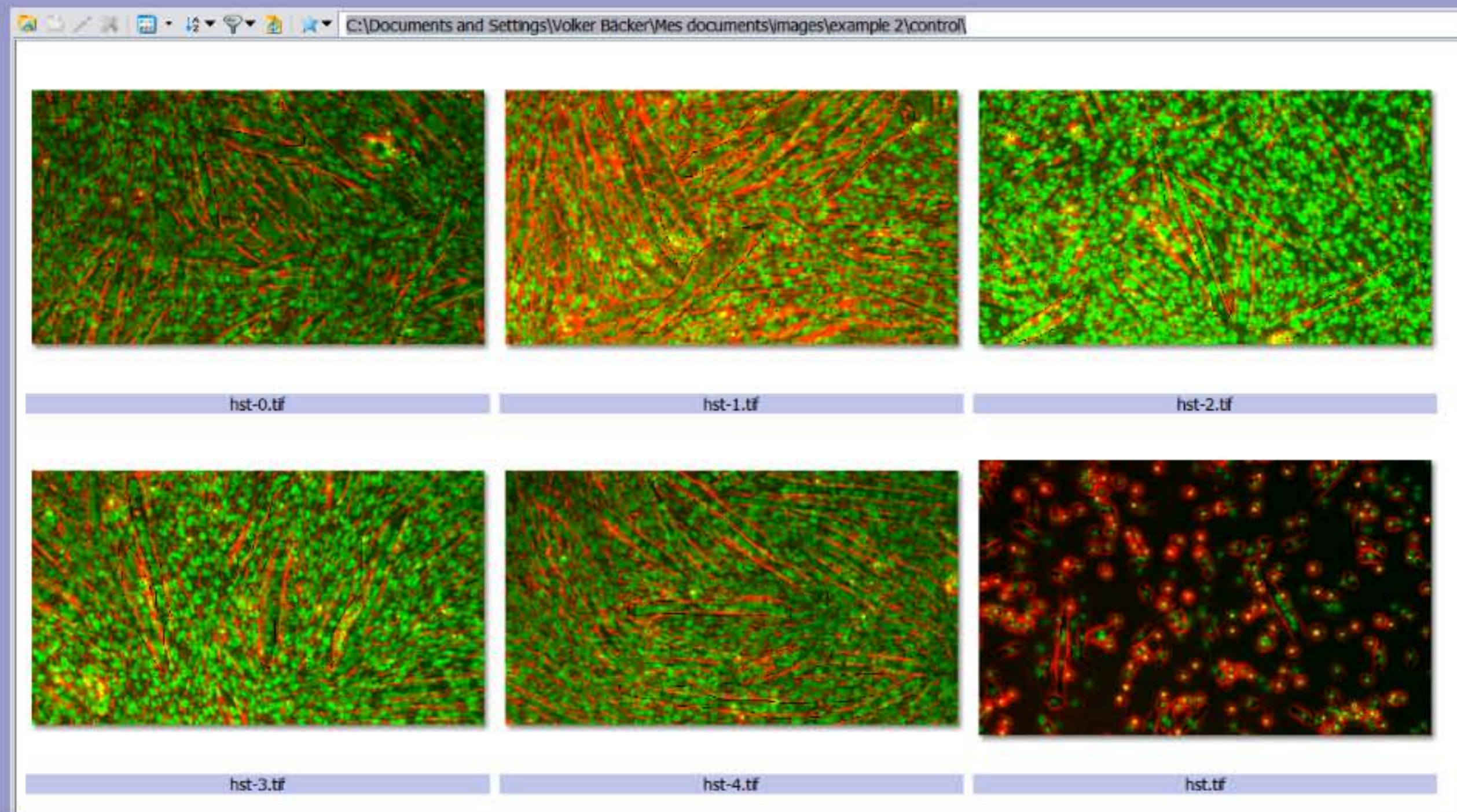
red

green

blue

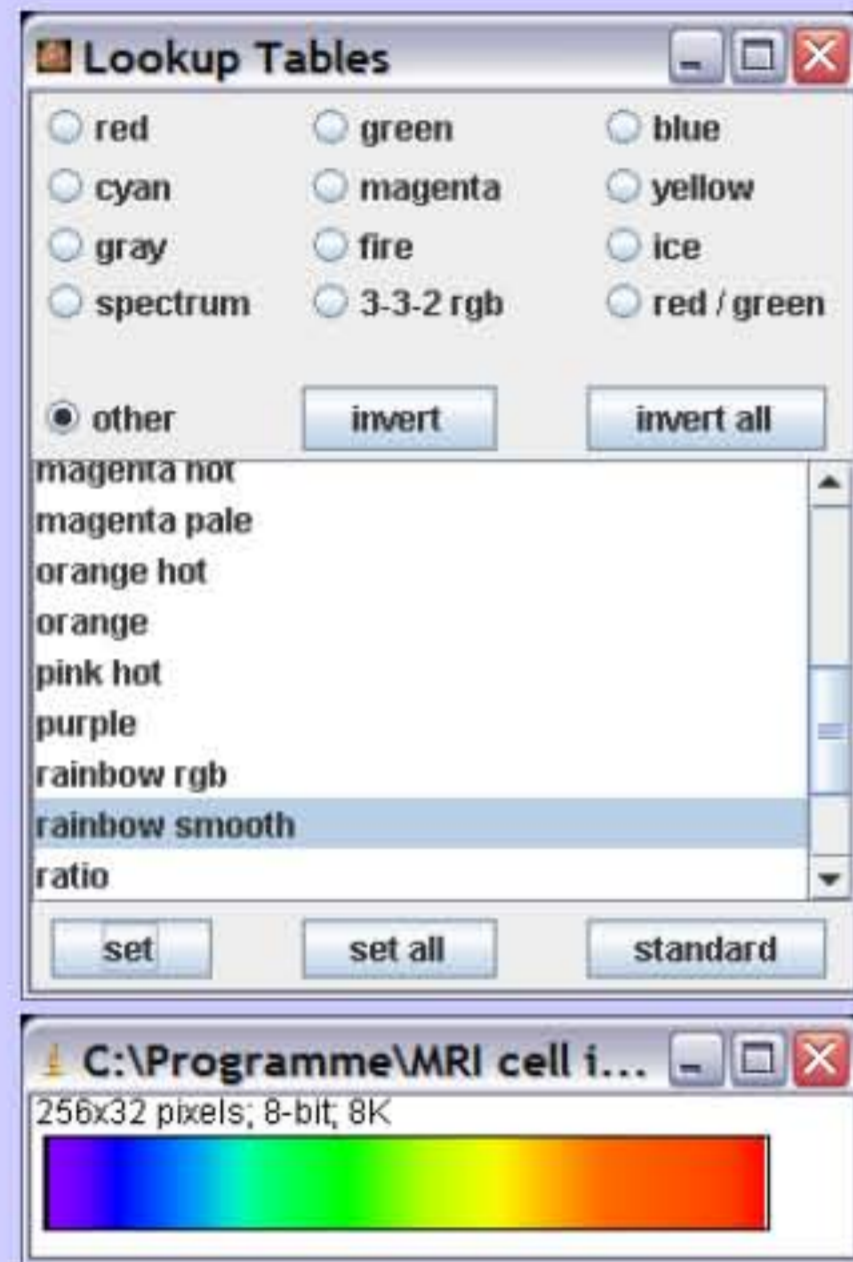


- application selects nuclei
- user corrects selection
- user presses accept to continue

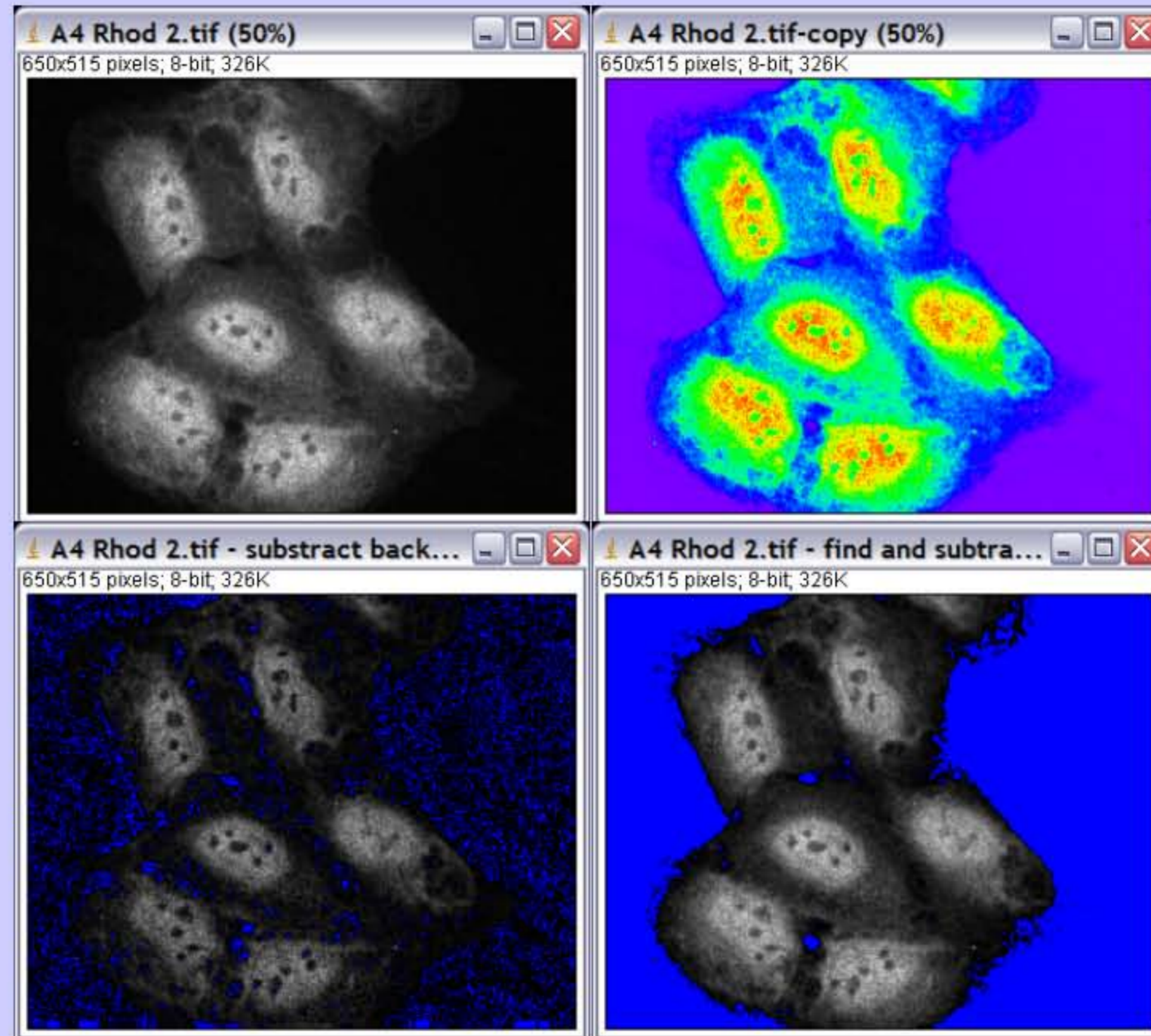


	A	B	C	D	E	F
1	Image	Areas	Objects	Objects inside areas	ratio	folder
2	hst-0.tif	3	891	52	0,06	C:\images\example 2\
3	hst-1.tif	4	918	89	0,1	C:\images\example 2\
4	hst-2.tif	3	942	61	0,06	C:\images\example 2\
5	hst-3.tif	4	899	70	0,08	C:\images\example 2\
6	hst-4.tif	9	966	116	0,12	C:\images\example 2\
7	hst.tif	3	315	17	0,05	C:\images\example 2\

Further interactive tools - LUT tool



- apply lookup table to
 - active image
 - all open images



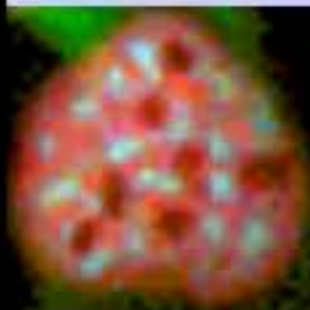
1. gray
2. rainbow smooth
3. hilo after "subtract background"
4. hilo after "find and subtract" background



Further interactive tools – slide show control



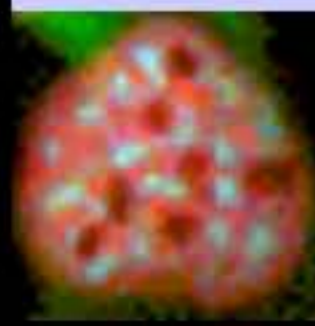
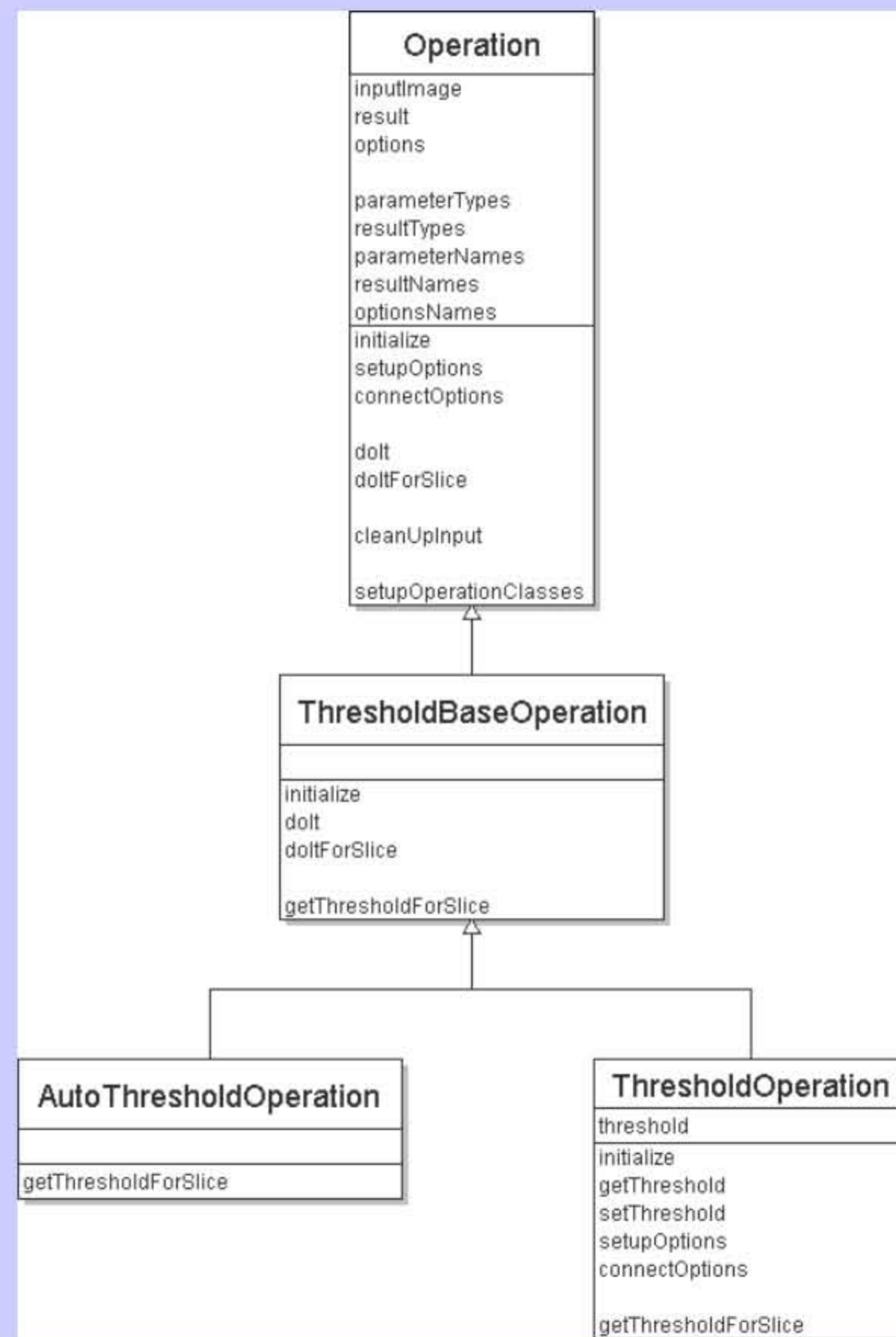
- **navigation**
 - select folder
 - goto first image
 - goto last image
 - goto next image
 - goto last image
 - reload current image
- **when opening next image**
 - close all open images
 - close last image opened with slide show control
 - keep position and zoom
 - adjust brightness/contrast (fix or auto)
 - apply LUT





Extending the framework

- to add an operation (without options)
 - declare input parameter and result in initialize
 - implement
 - dolt
 - doltForSlice



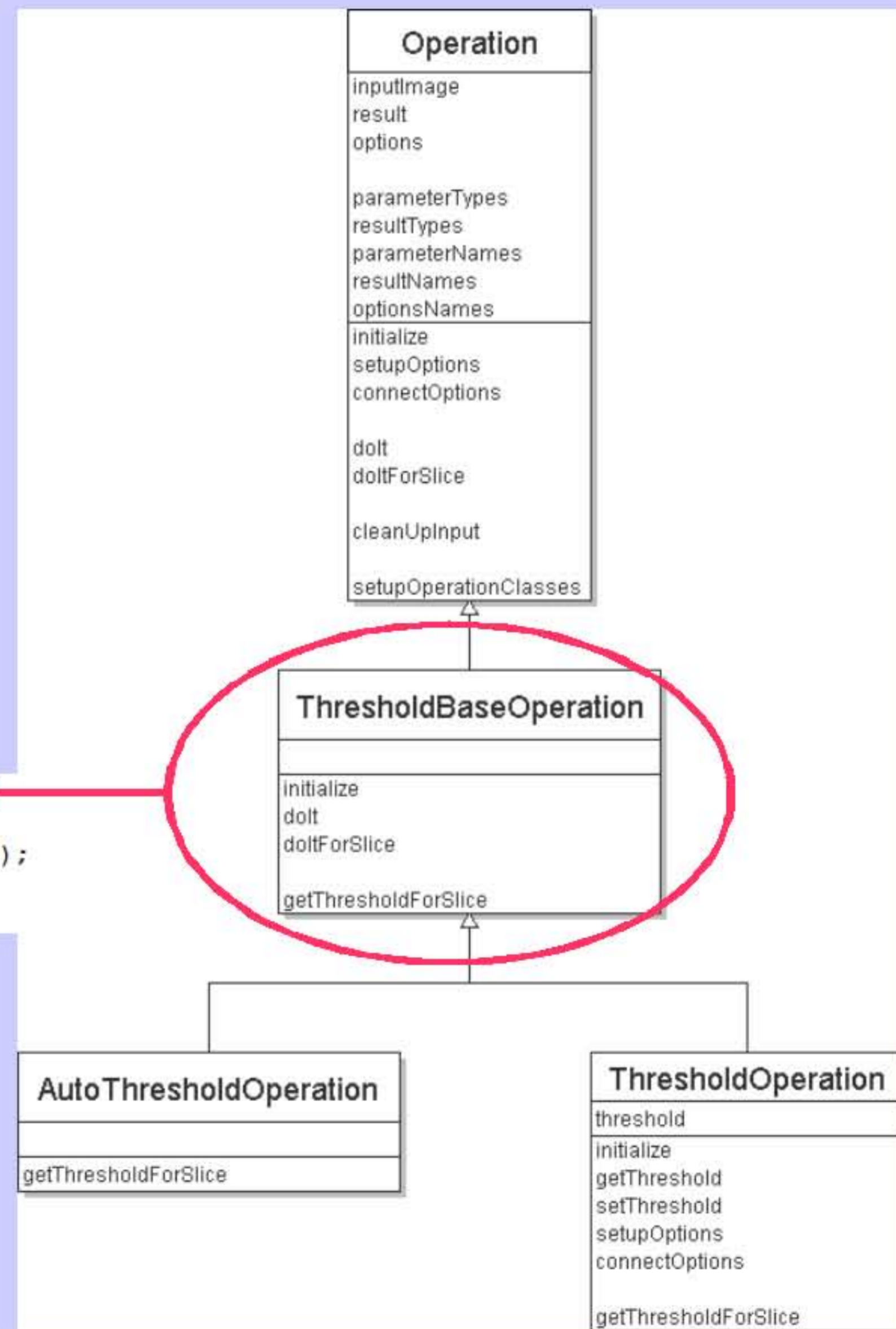


Extending the framework

```
protected void initialize() throws ClassNotFoundException {
    super.initialize();
    parameterTypes = new Class[1];
    parameterTypes[0] = Class.forName("ij.ImagePlus");
    parameterNames = new String[1];
    parameterNames[0] = "InputImage";
    resultTypes = new Class[1];
    resultTypes[0] = Class.forName("ij.ImagePlus");
    resultNames = new String[1];
    resultNames[0] = "Result";
}
```

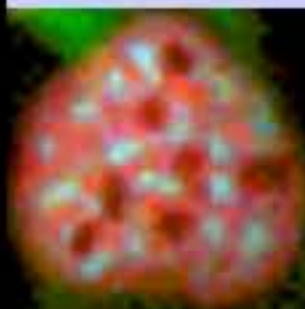
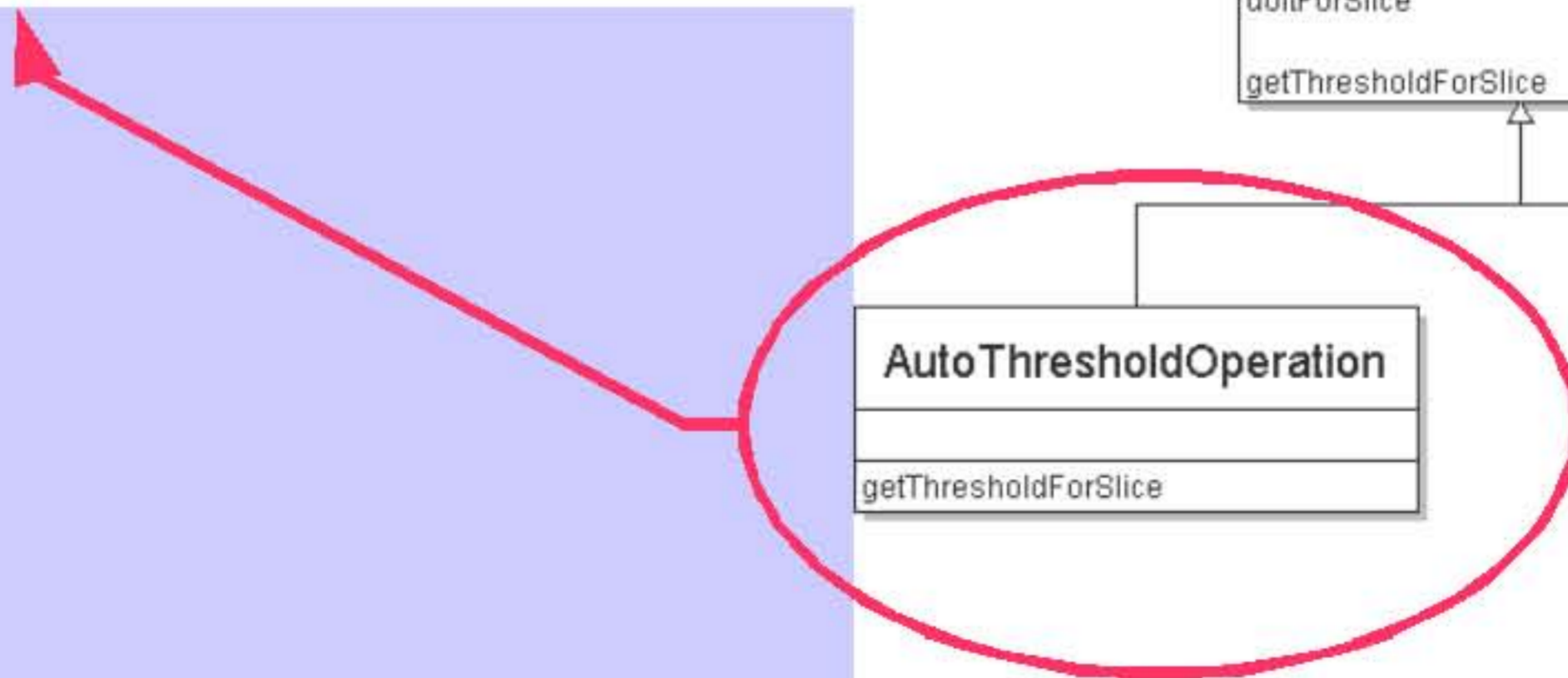
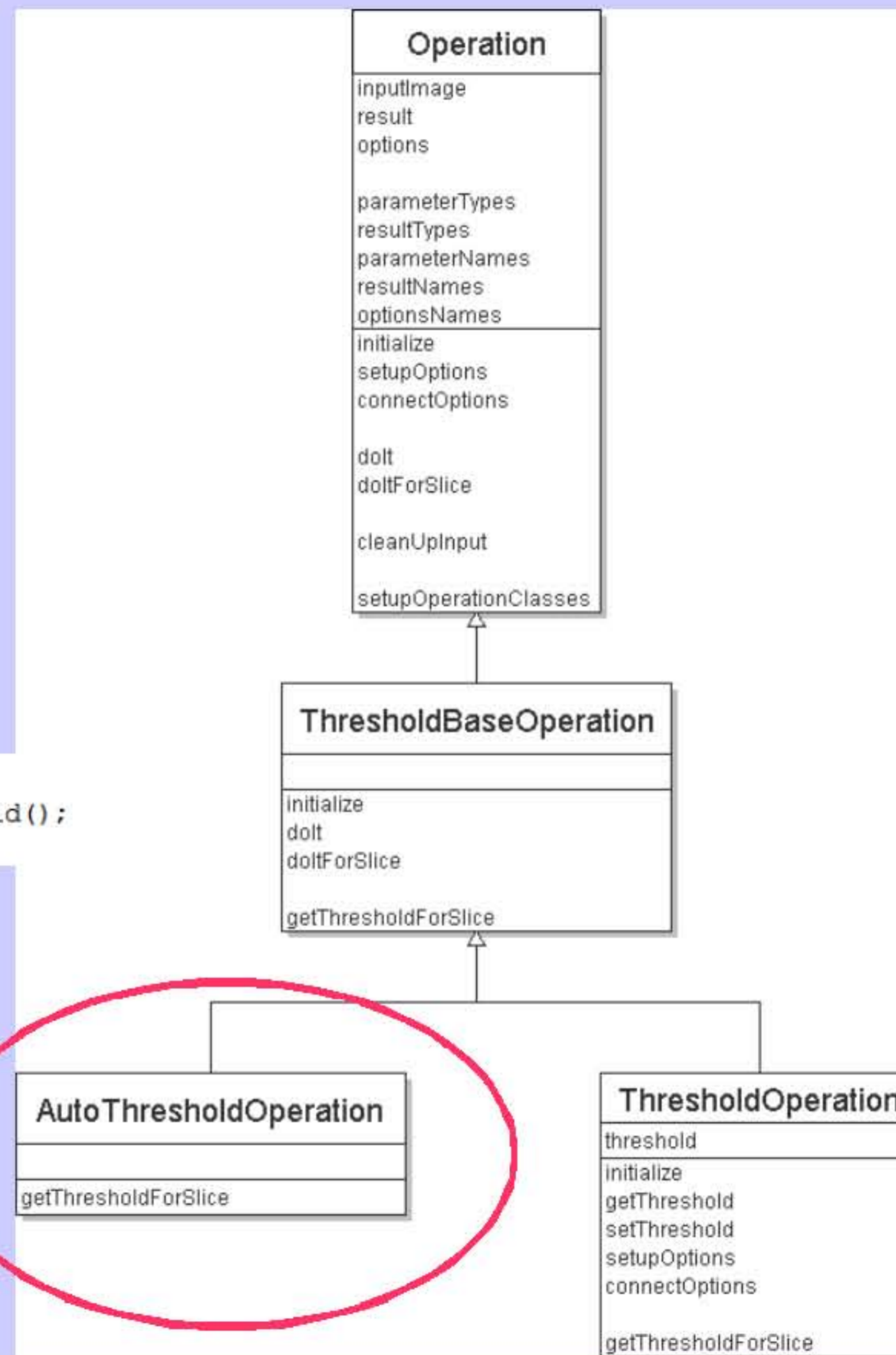
```
public void doIt() {
    ImagePlus inputImage = this.getInputImage();
    result = this.getCopyOfOrReferenceTo(inputImage, inputImage.getTitle());
    this.processSlices();
}
```

```
protected void doItForSlice(int sliceNumber, ImageStack stack) {
    ImageProcessor ip = stack.getProcessor(sliceNumber);
    ip.threshold(this.getThresholdForSlice(sliceNumber));
}
```



Extending the framework

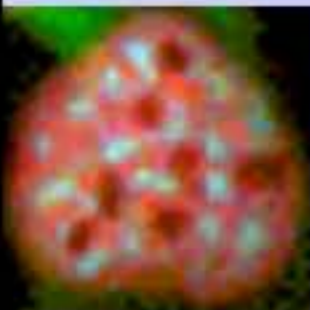
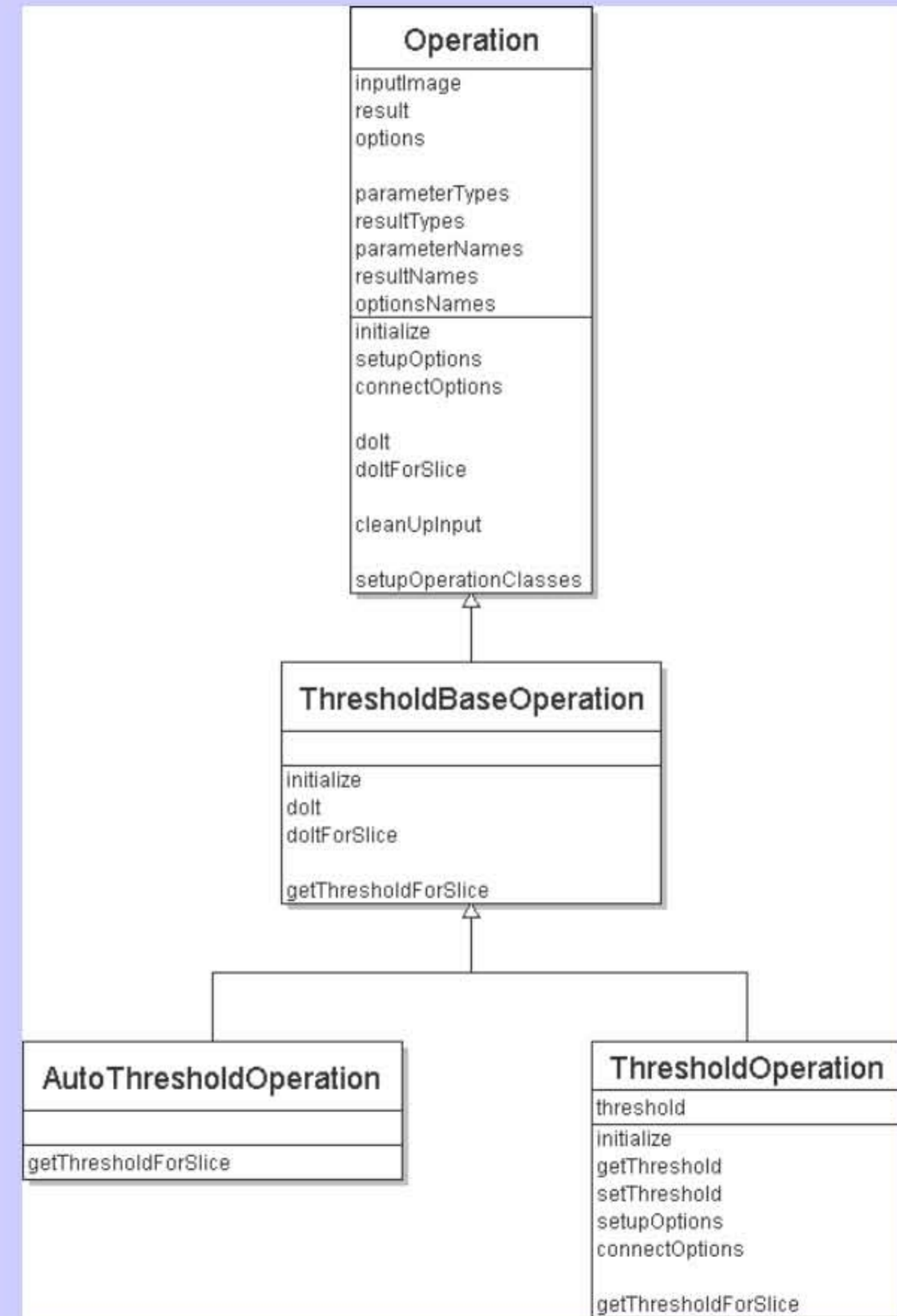
```
public int getThresholdForSlice(int i) {
    return this.getInputImage().getStack().getProcessor(i).getAutoThreshold();
}
```





Extending the framework

- to add an operation with options
 - declare options in initialize
 - setup options
 - set name, default value, min, max and short help text
 - to facilitate access
 - declare instance variable
 - connectOptions
 - getter / setter



Extending the framework

```
protected void initialize() throws ClassNotFoundException {
    super.initialize();
    optionsNames = new String[1];
    optionsNames[0] = "threshold";
}
```

```
protected void setupOptions() {
    super.setupOptions();
    this.setThreshold(128);
    threshold.setMin(0);
    threshold.setShortHelpText("all intensities above threshold" +
        " will be mapped to 255 and all below to 0");
}
```

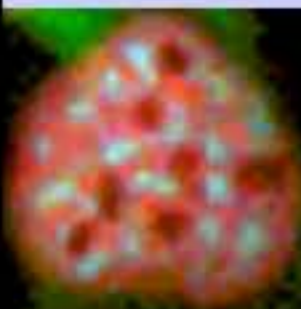
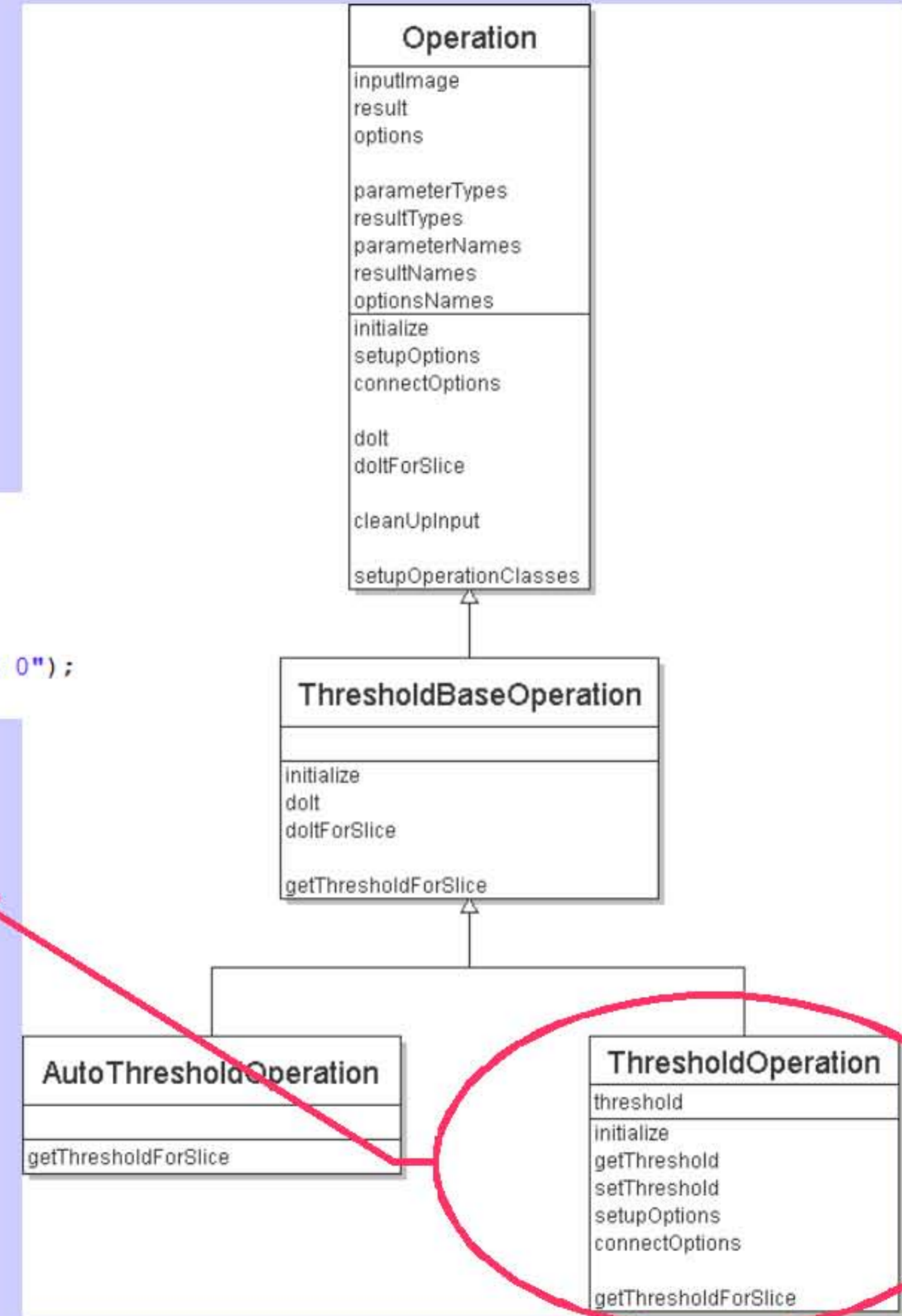
```
public int getThreshold() {
    return threshold.getIntegerValue();
}

public void setThreshold(int threshold) {
    this.threshold.setValue(new Integer(threshold).toString());
}
```

• getter / setter

```
public void connectOptions() {
    this.threshold = (Option) this.options.getOptions().elementAt(0);
}
```

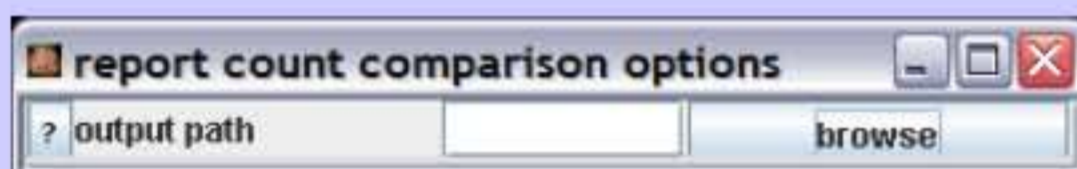
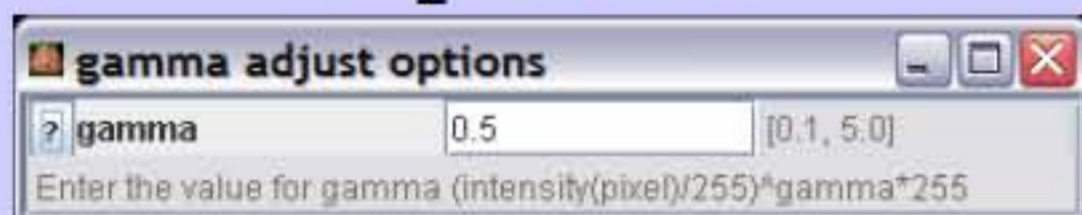
```
public int getThresholdForSlice(int i) {
    return this.getThreshold();
}
```



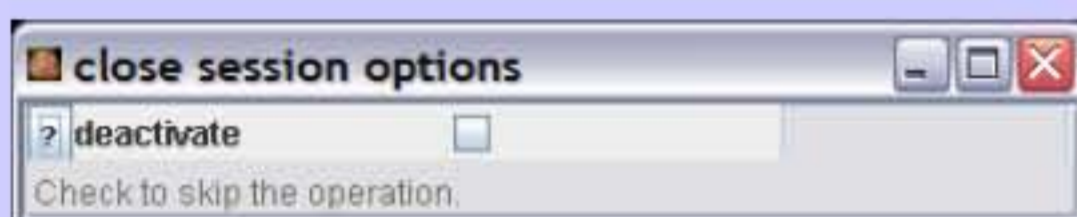
Options

ChoiceOption

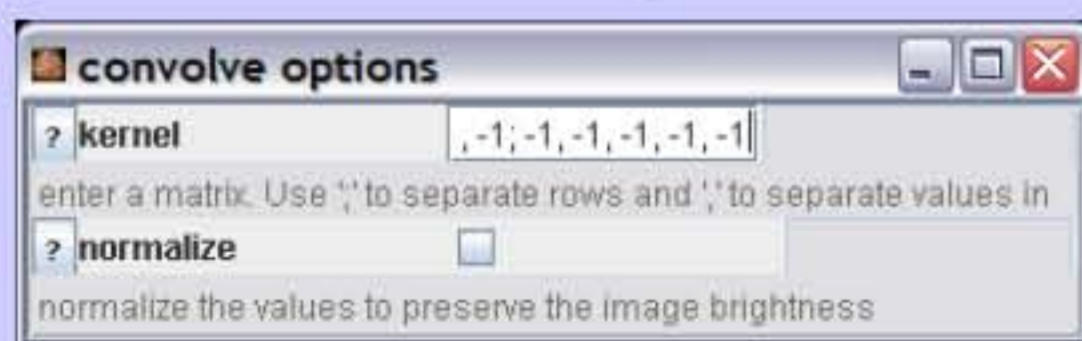
Option



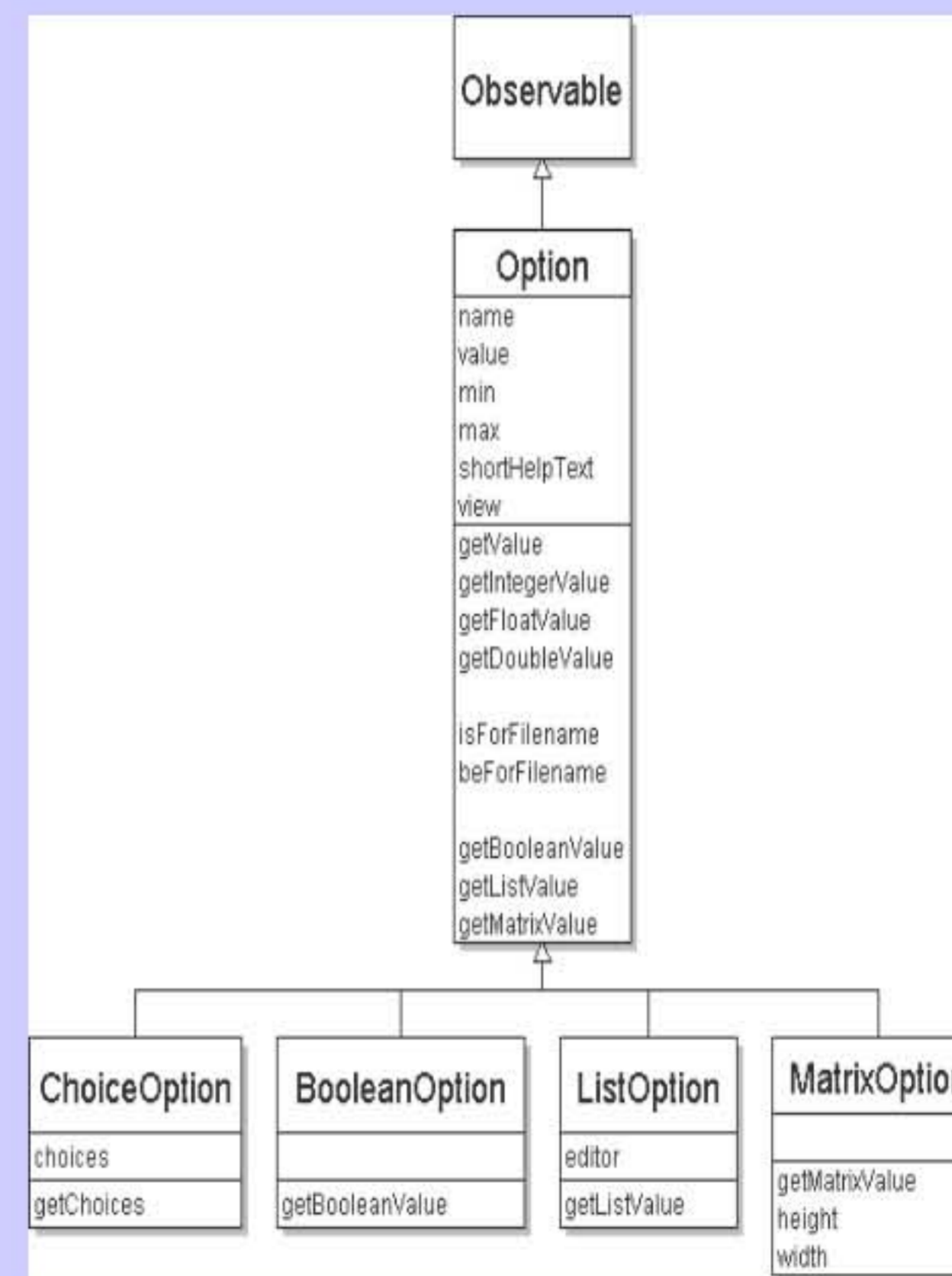
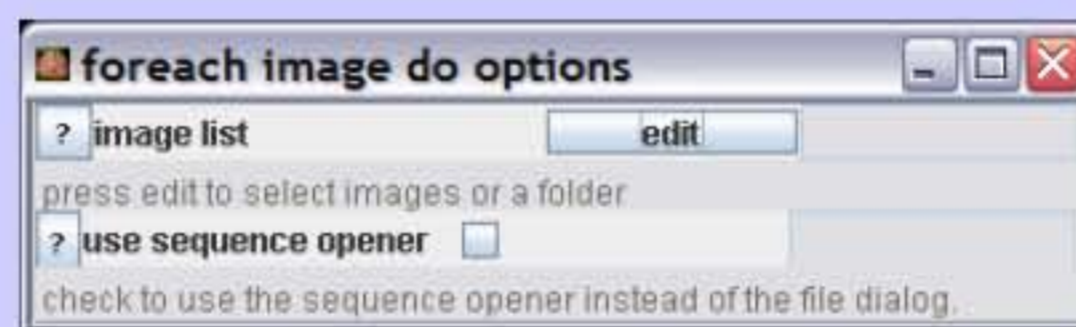
BooleanOption



MatrixOption



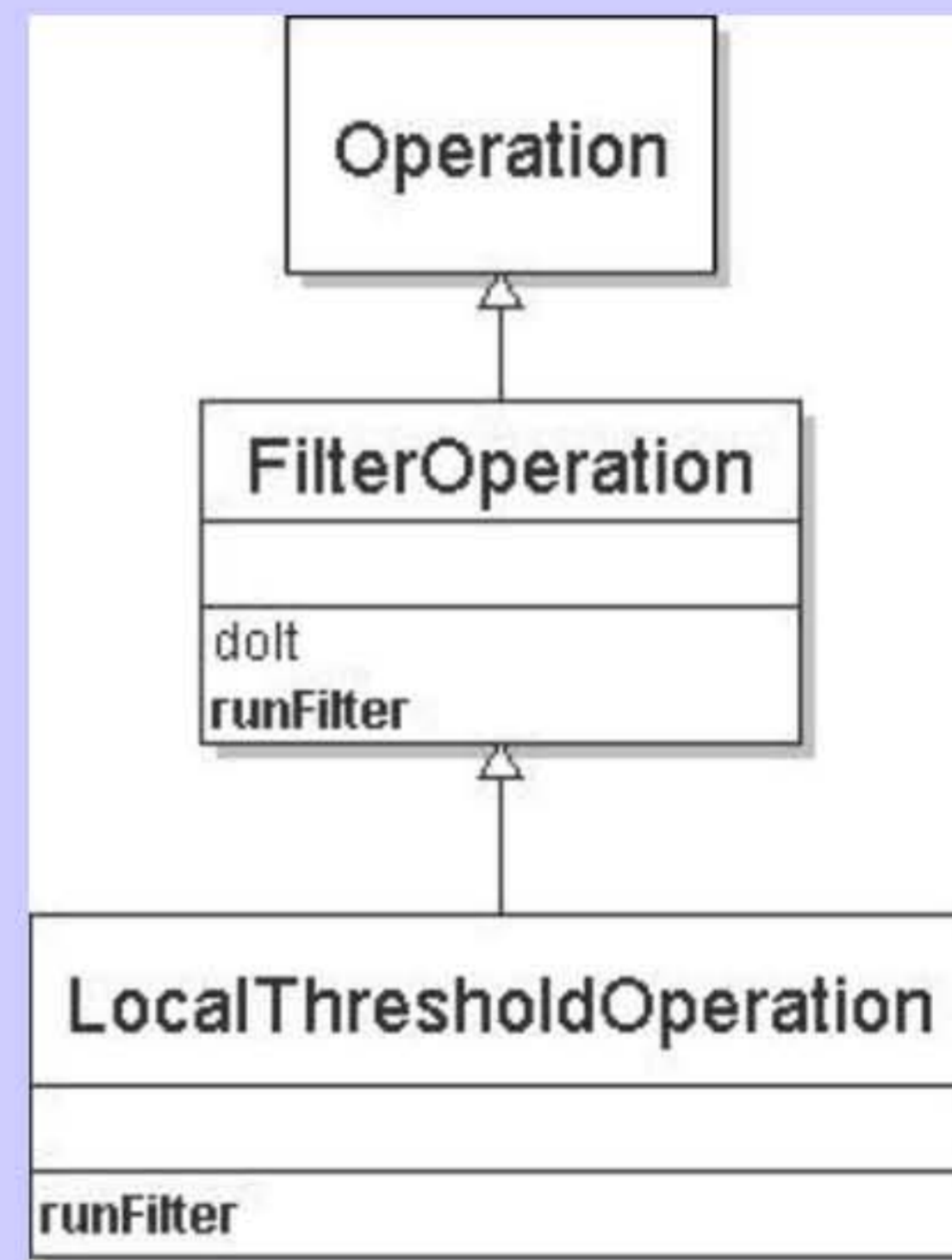
ListOption



Accessing ImageJ methods

- simplest method:
- use the ImageJ macros interface
 - IJ.run(...)

```
public void runFilter() {  
    IJ.run("Duplicate...", "title=Original");  
    IJ.run("32-bit");  
    IJ.run("Duplicate...", "title=Filtered");  
    IJ.selectWindow("Filtered");  
    IJ.run(this.getFilter(), "radius=" + this.getFilterRadius());  
    IJ.run("Image Calculator...",  
          "image1=Original operation=Subtract image2=Filtered create");  
    IJ.run("Rename...", "title=Result");  
    IJ.run("8-bit");  
}
```



Local threshold macro by R.Couture 11-14-03 Dept. of Radiology Washington University School of Medicine, couture@wuerl.wustl.edu

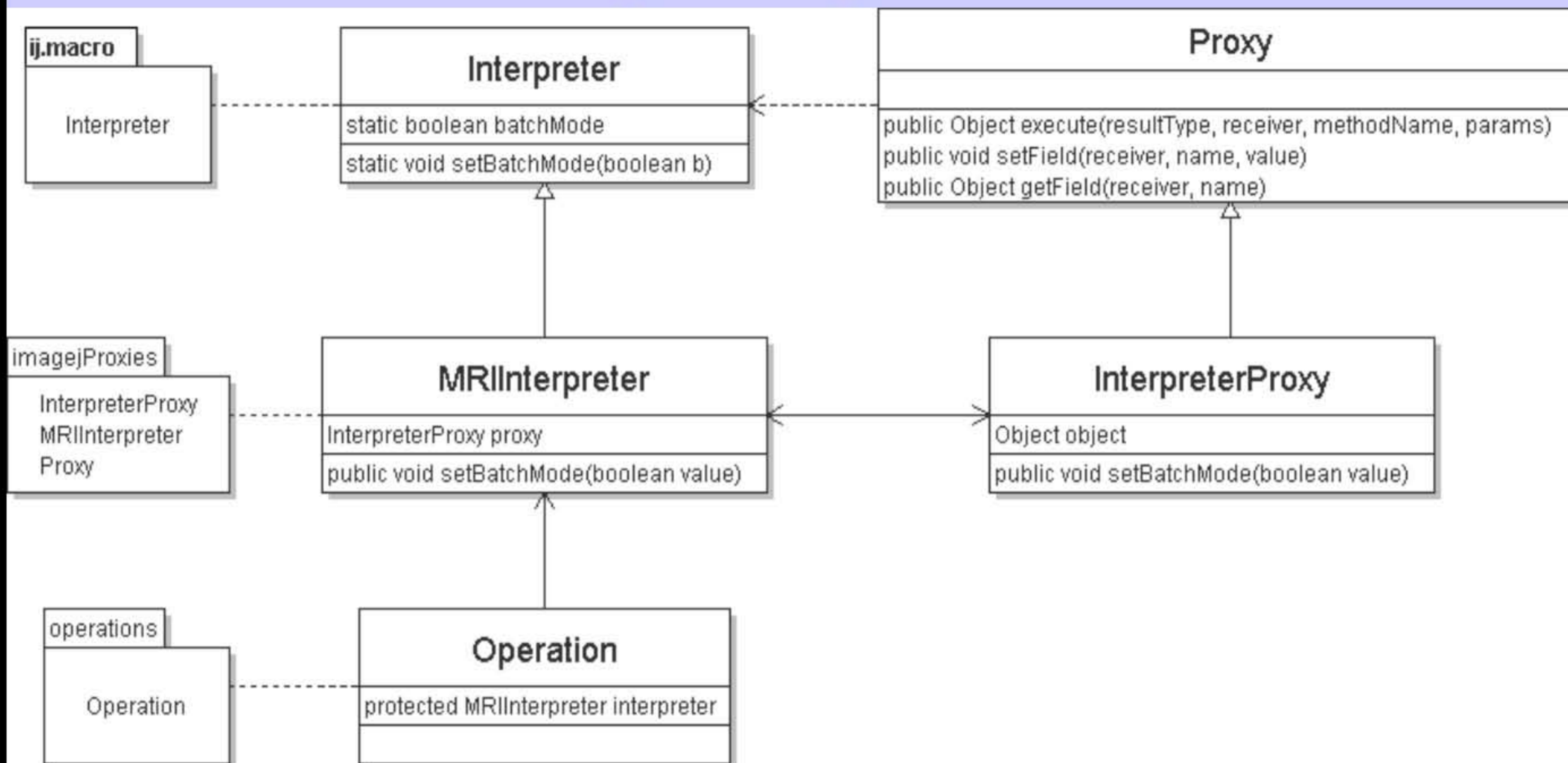


Accessing ImageJ methods

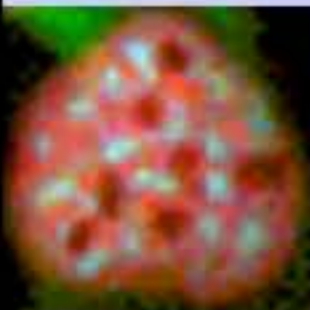
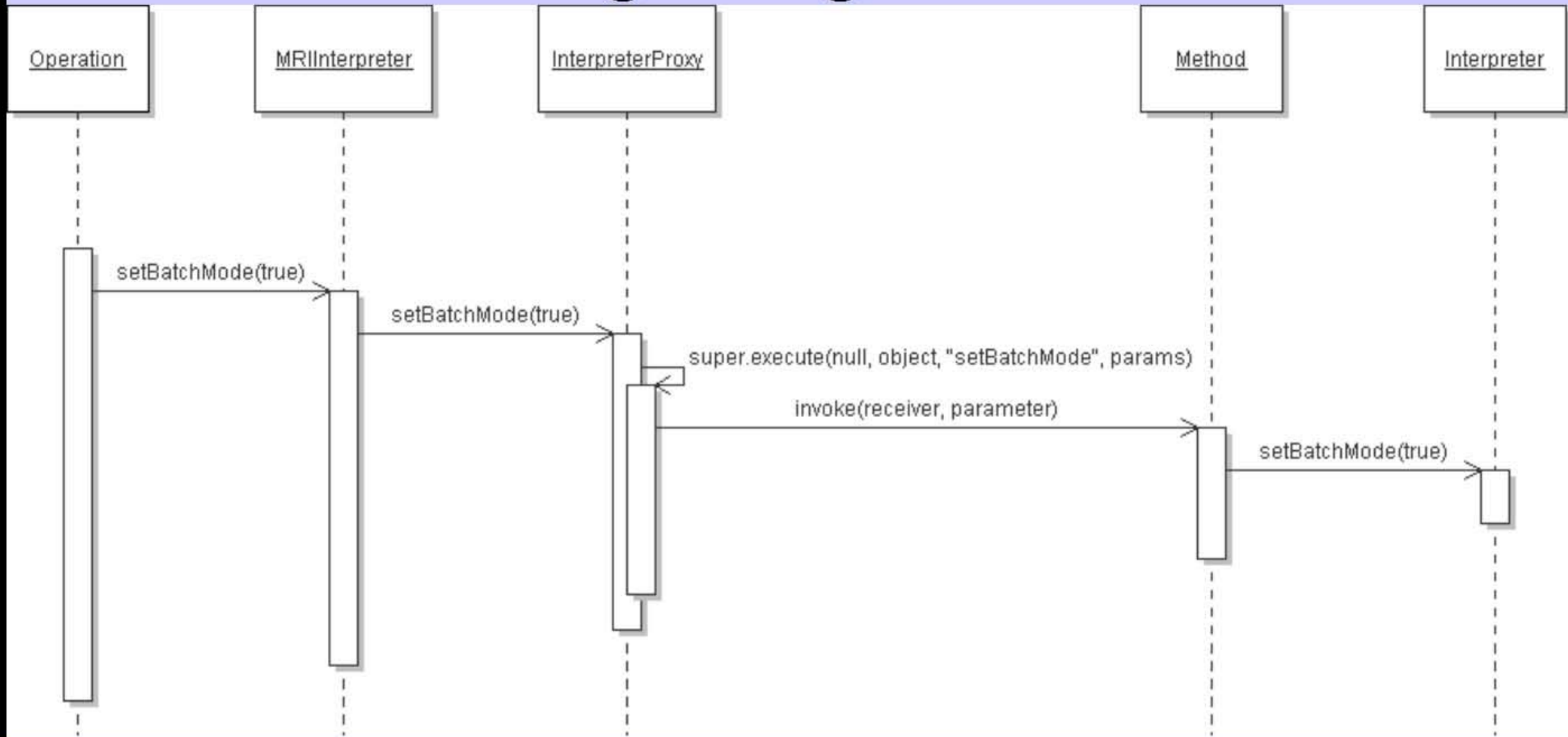
- what if method or data member not accessible
 - access to protected:
 - subclass in own package
 - access to private:
 - use java reflection api
- class Proxy facilitates reflection api usage
 - Object execute(...)
 - void setField(...)
 - Object getField(...)



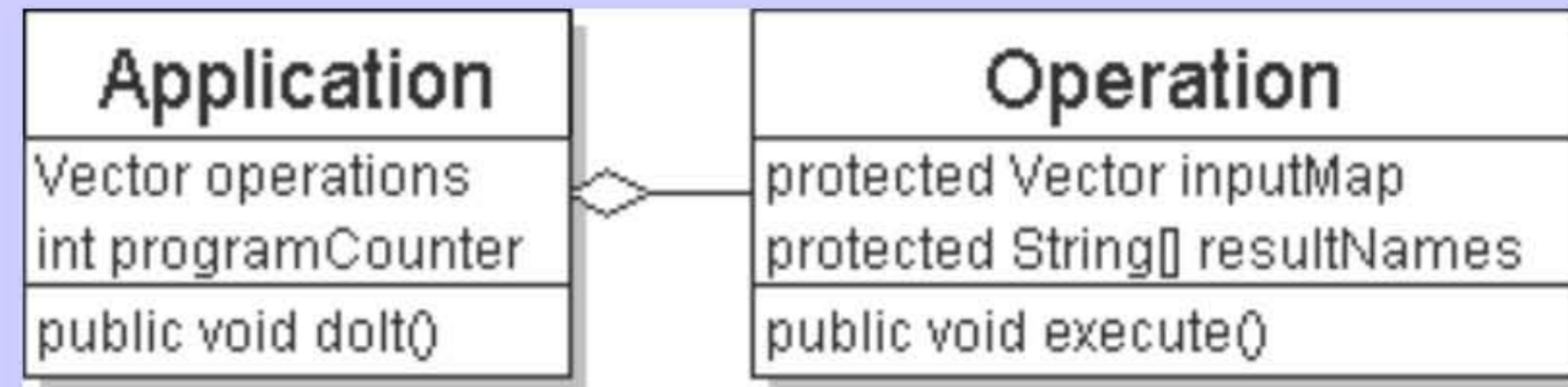
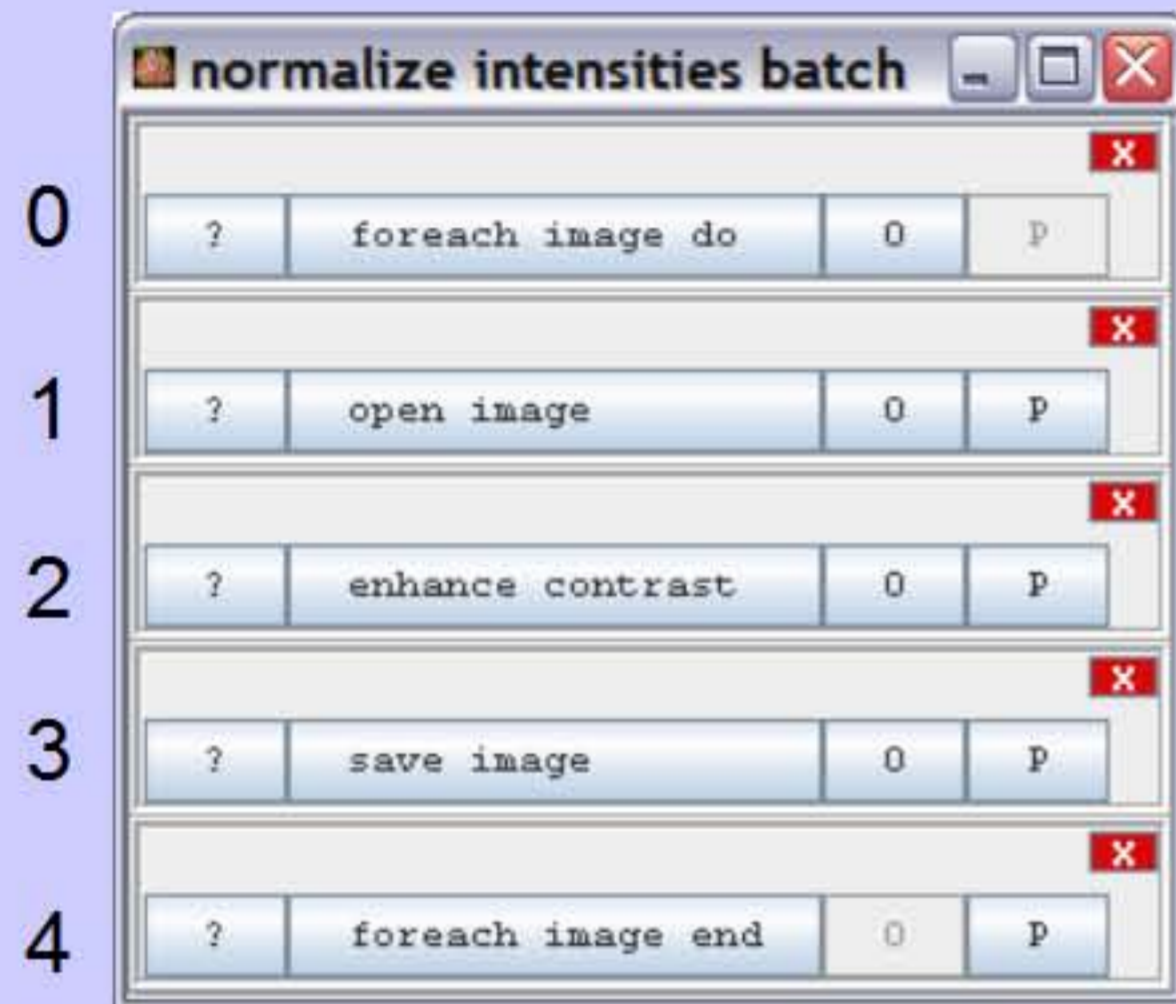
Accessing ImageJ methods



Accessing ImageJ methods



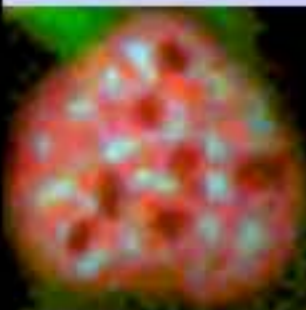
The implementation



- When application is loaded from text file
 - objects are created from names, using reflection
 - parameters of operation are connected to results of preceding operations

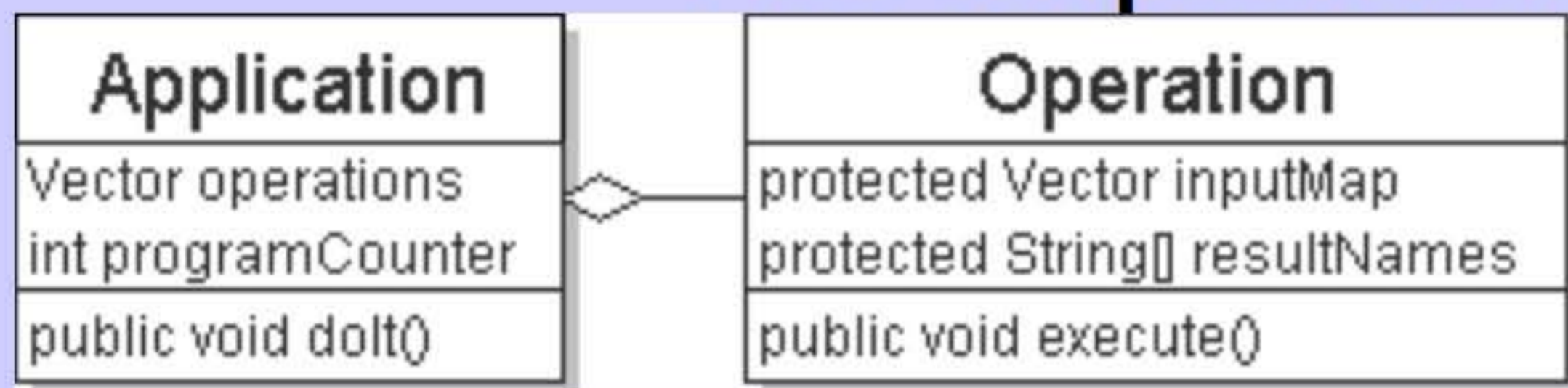
```

input maps:
  operation      result nr.
0:      -          -
1:      0          0
2:      1          0
3:      2          0
         0          0
4:      0          2
  
```





The implementation



Application>>doIt

FOR programCounter = 0 TO numberOfOperations - 1

if (stopped) return

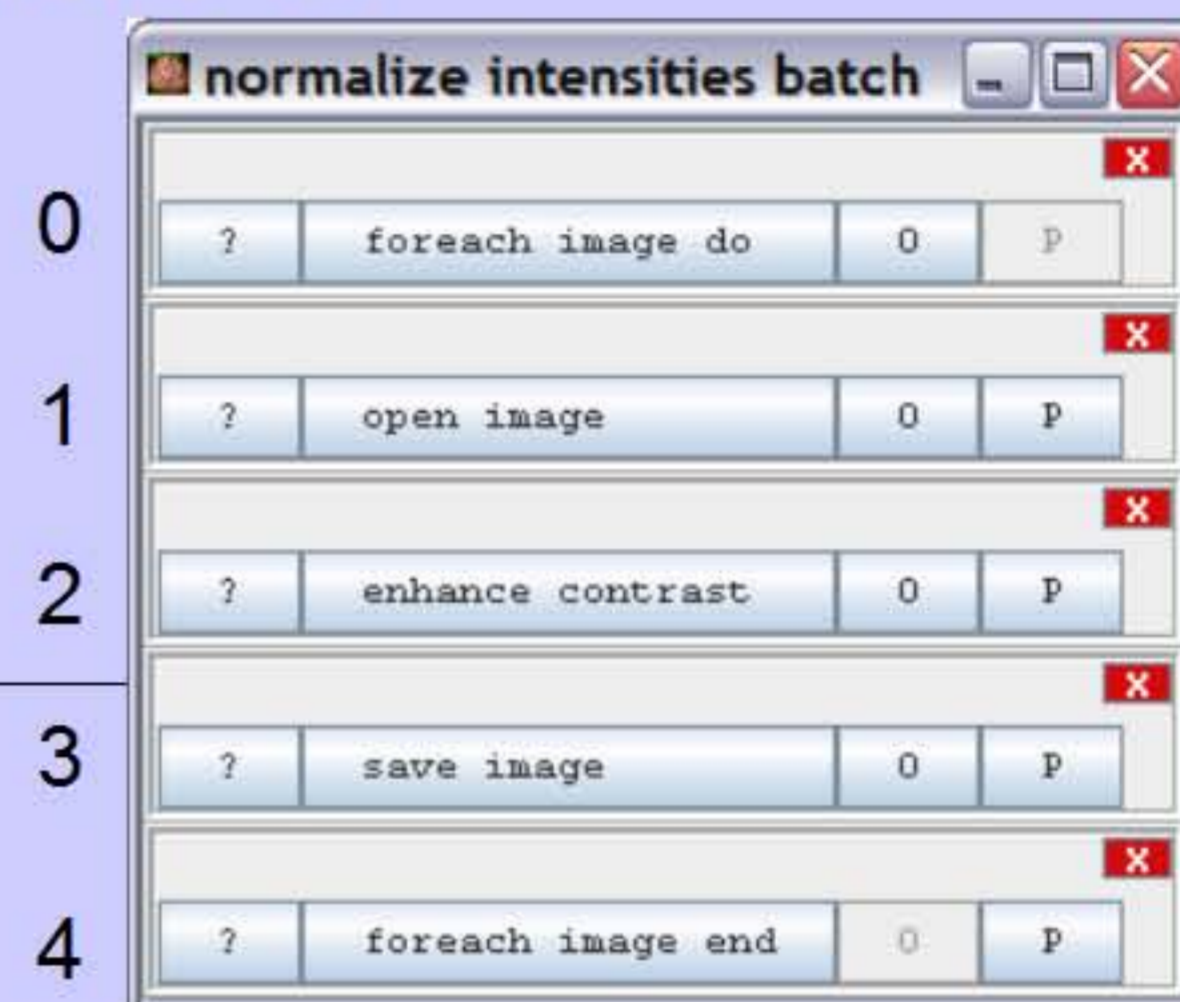
execute operation

cleanup preceding results not referenced afterwards

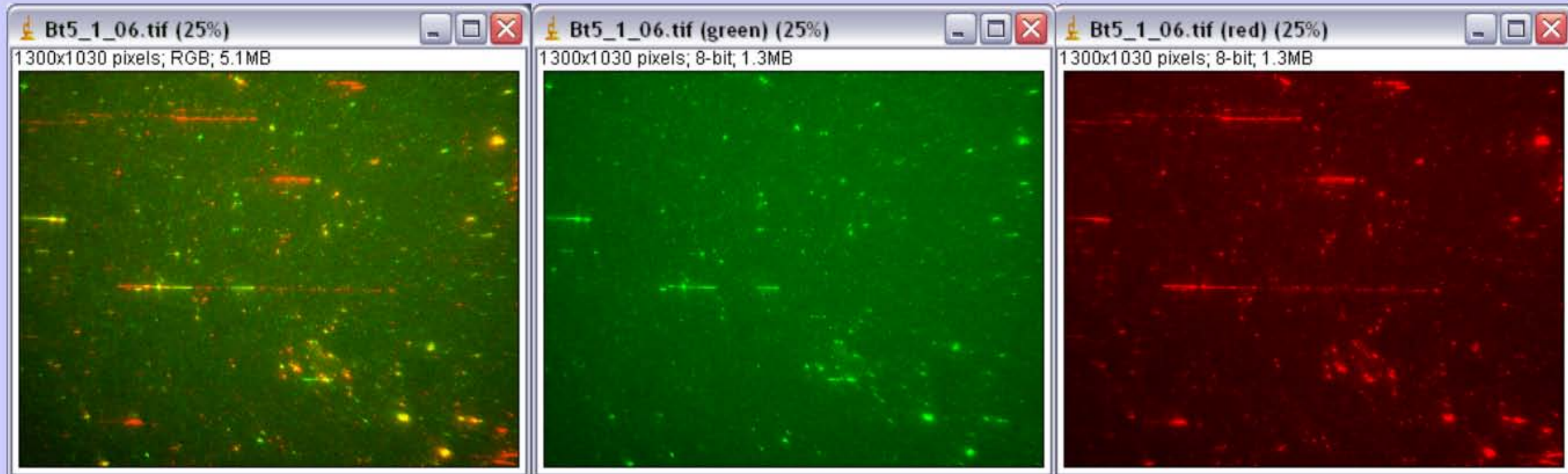
get next operation

set parameters of next operation from results of preceding operations, using reflection

jump and loop operations modify the programCounter



DNA combing



the images:

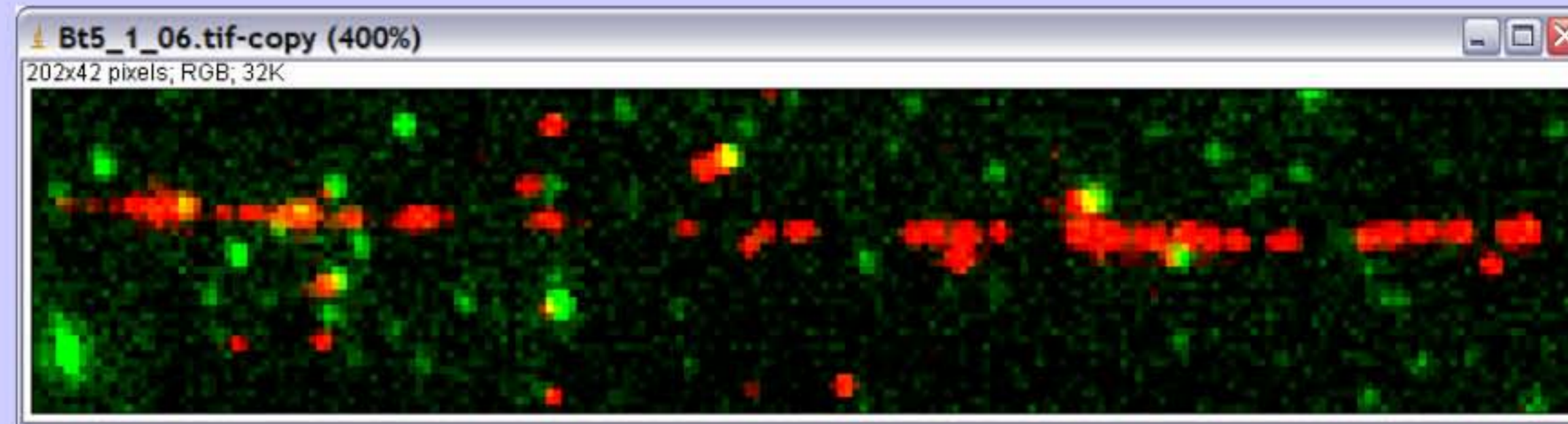
red: combed DNA
green: sites where replication takes place

the task, measure:

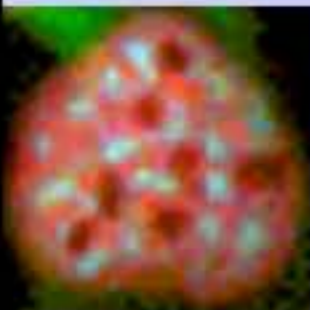
- the lengths of the DNA molecules
- the lengths of the replication sites within each DNA molecule
- the distances between replication sites for each DNA molecule



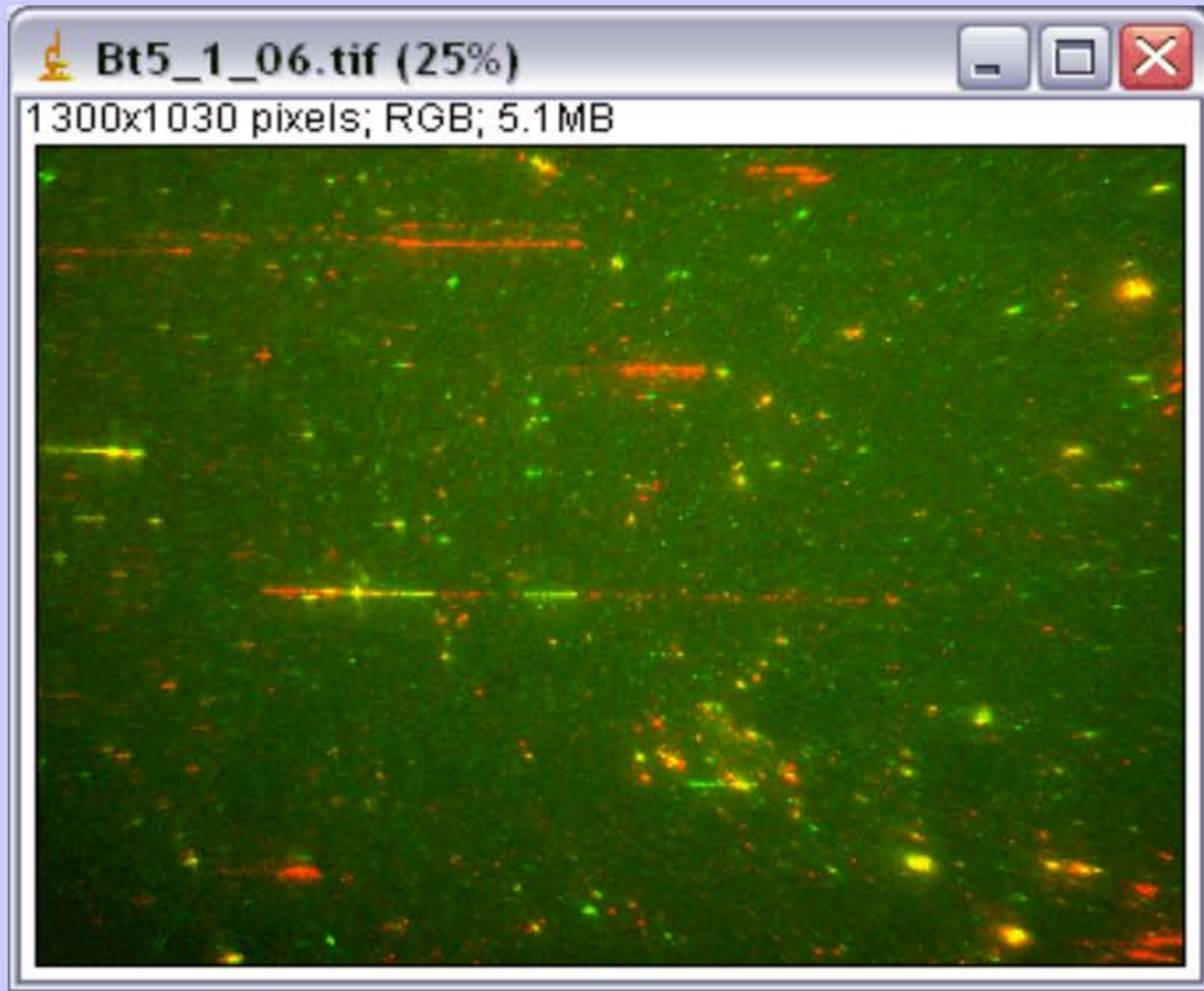
DNA combing



- difficulties
 - high level of (biochemical) noise
 - stretched out molecules are not straight
 - signal has large gaps



DNA combing



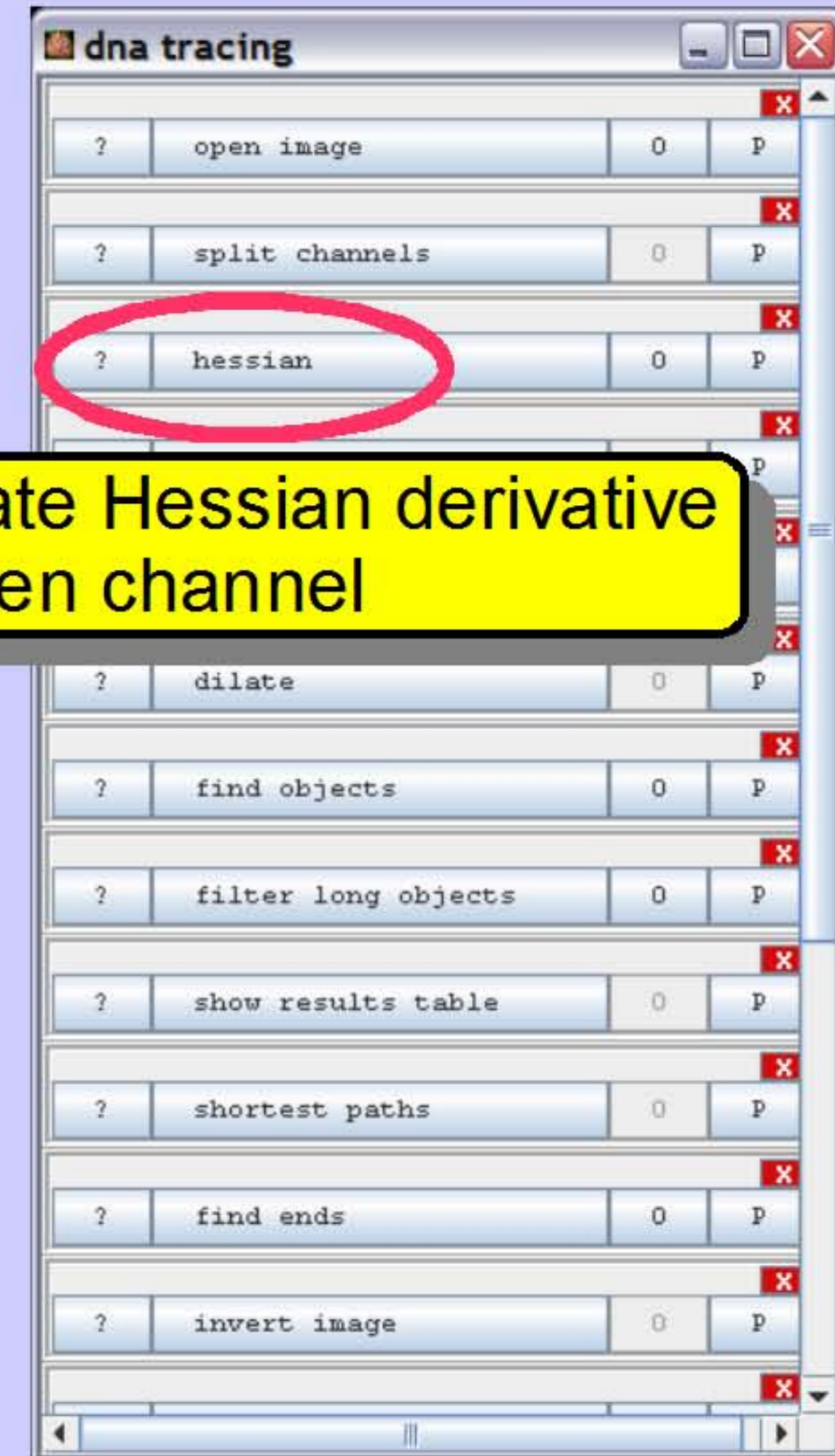
Screenshot of the ImageJ macro editor window titled 'dna tracing batch'. The macro contains the following steps:

? foreach image do	0	P
? open image	0	P
? split channels	0	P
? hessian	0	P
? get image from hessian	0	P
? auto threshold	0	P
? dilate	0	P
? find objects	0	P
? filter long objects	0	P
? shortest paths	0	P
? find ends	0	P
? invert image	0	P

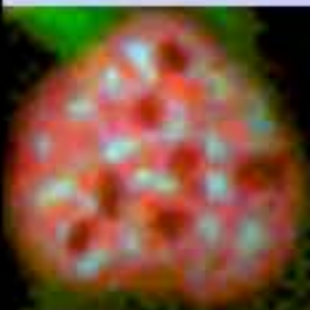




DNA combing

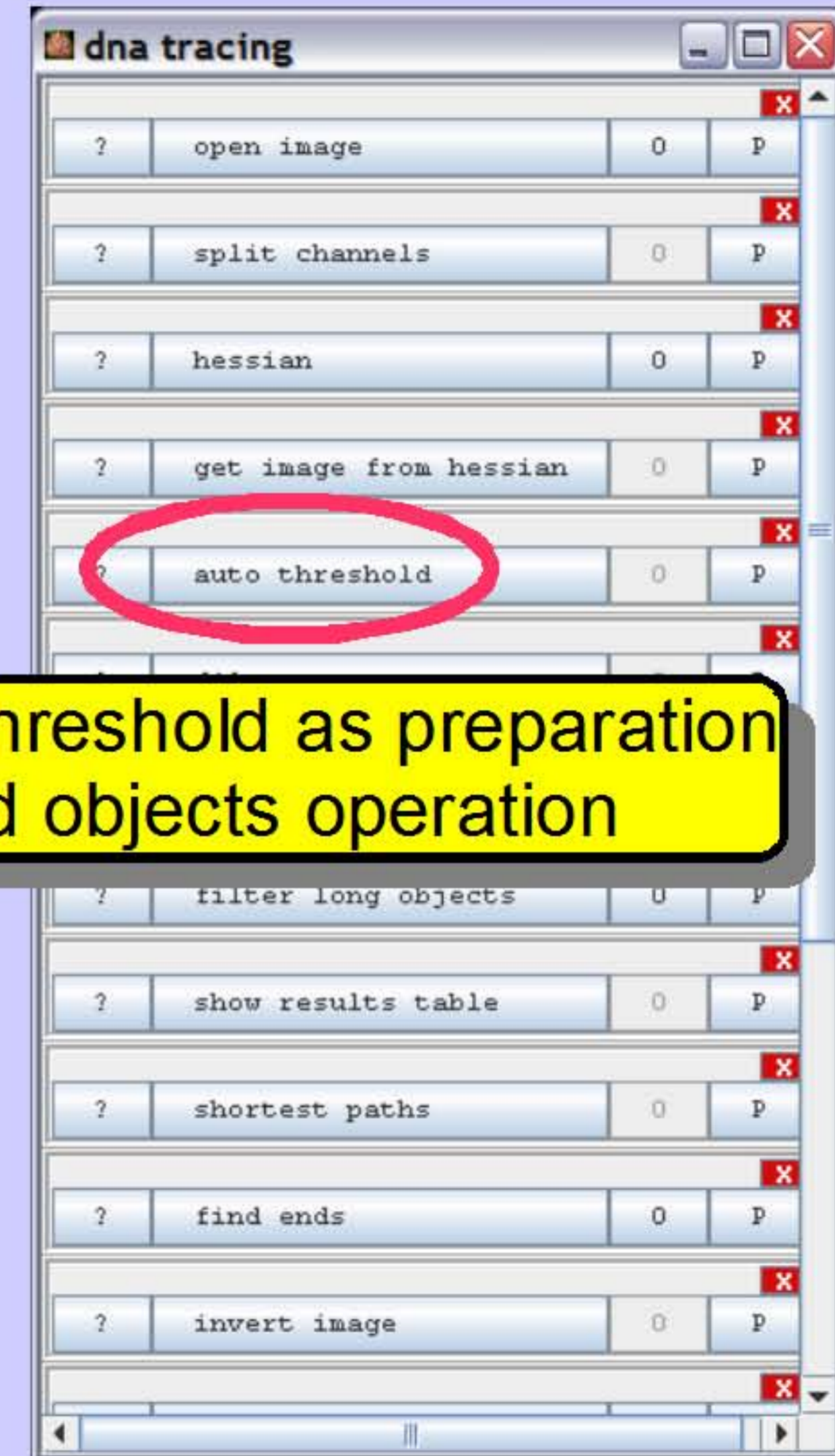
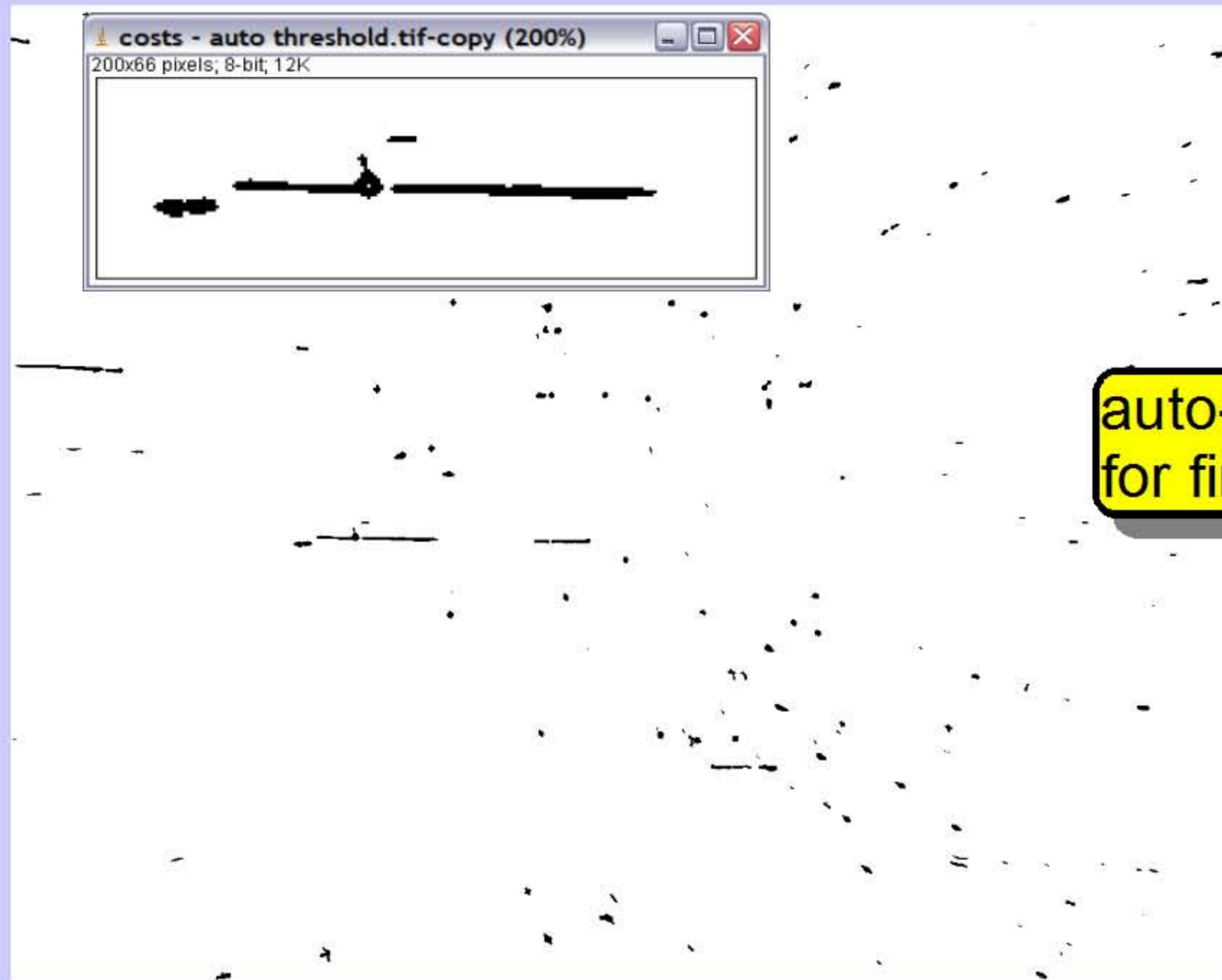


calculate Hessian derivative
on green channel

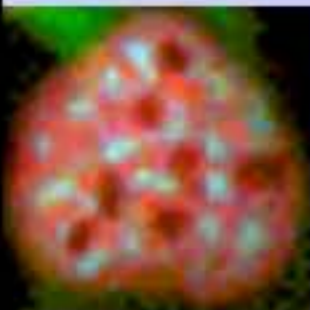




DNA combing

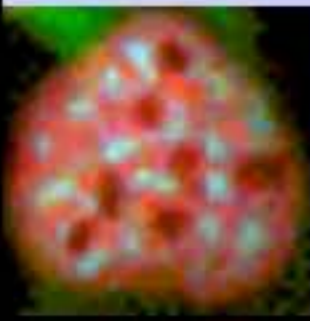
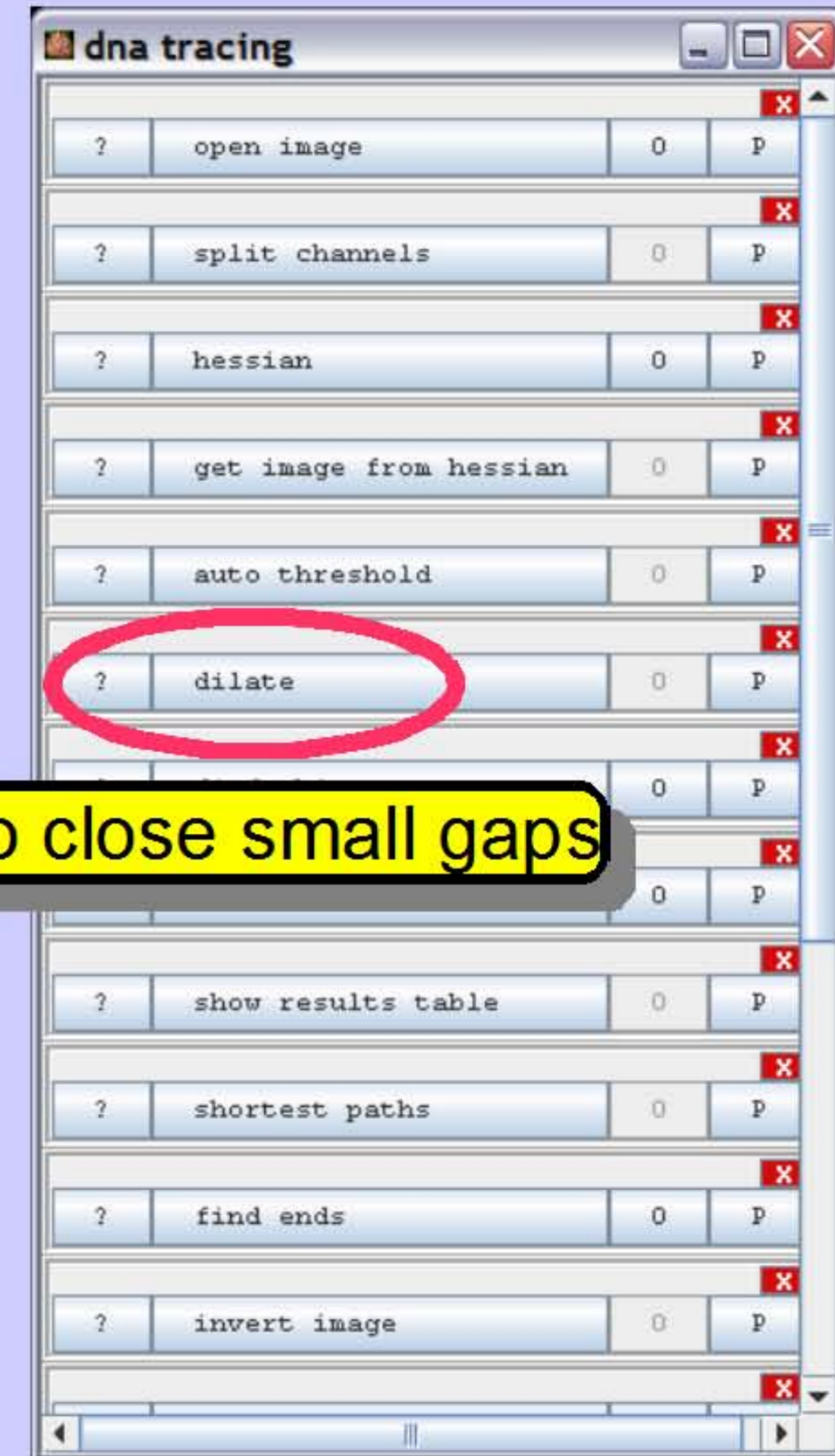
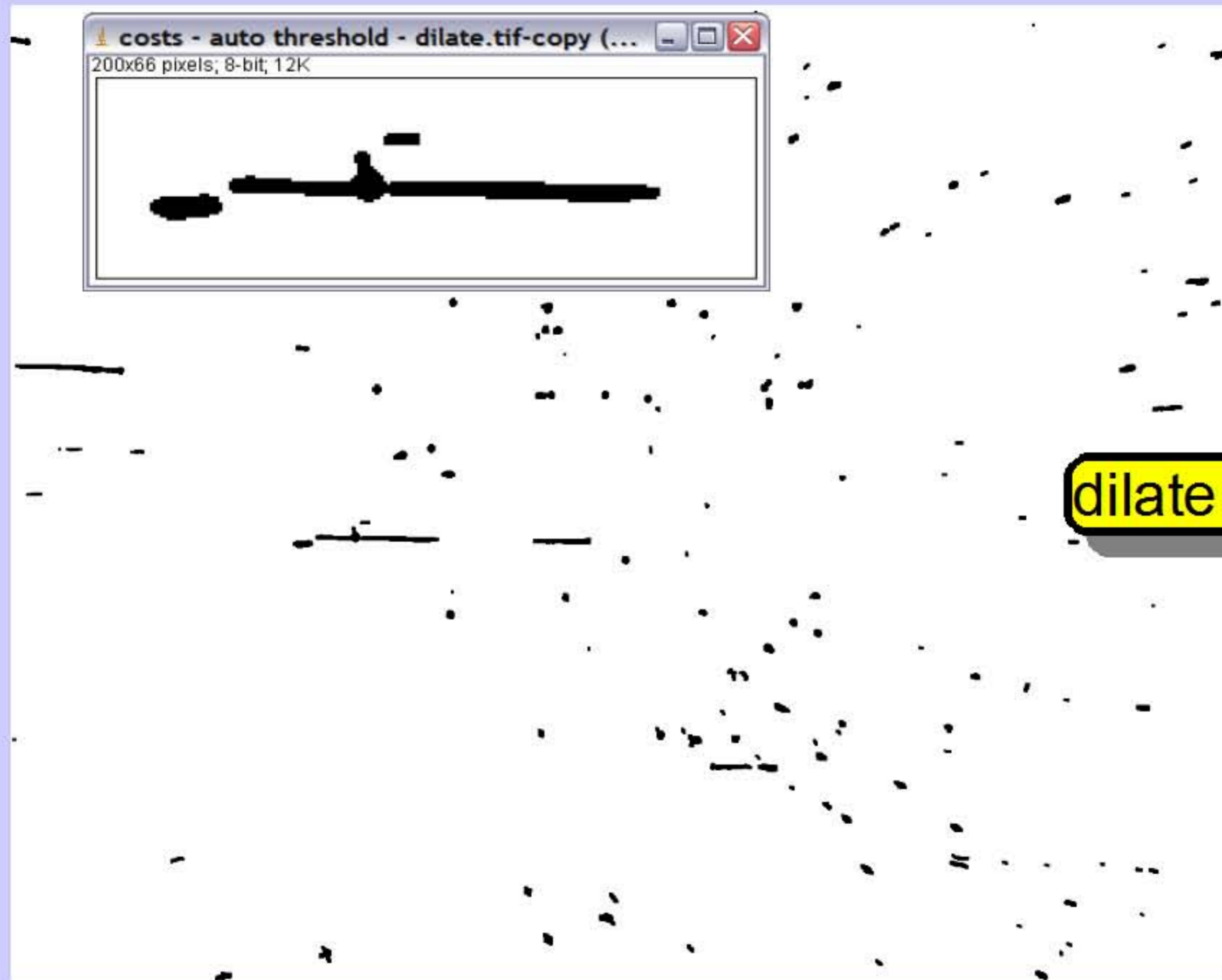


auto-threshold as preparation
for find objects operation



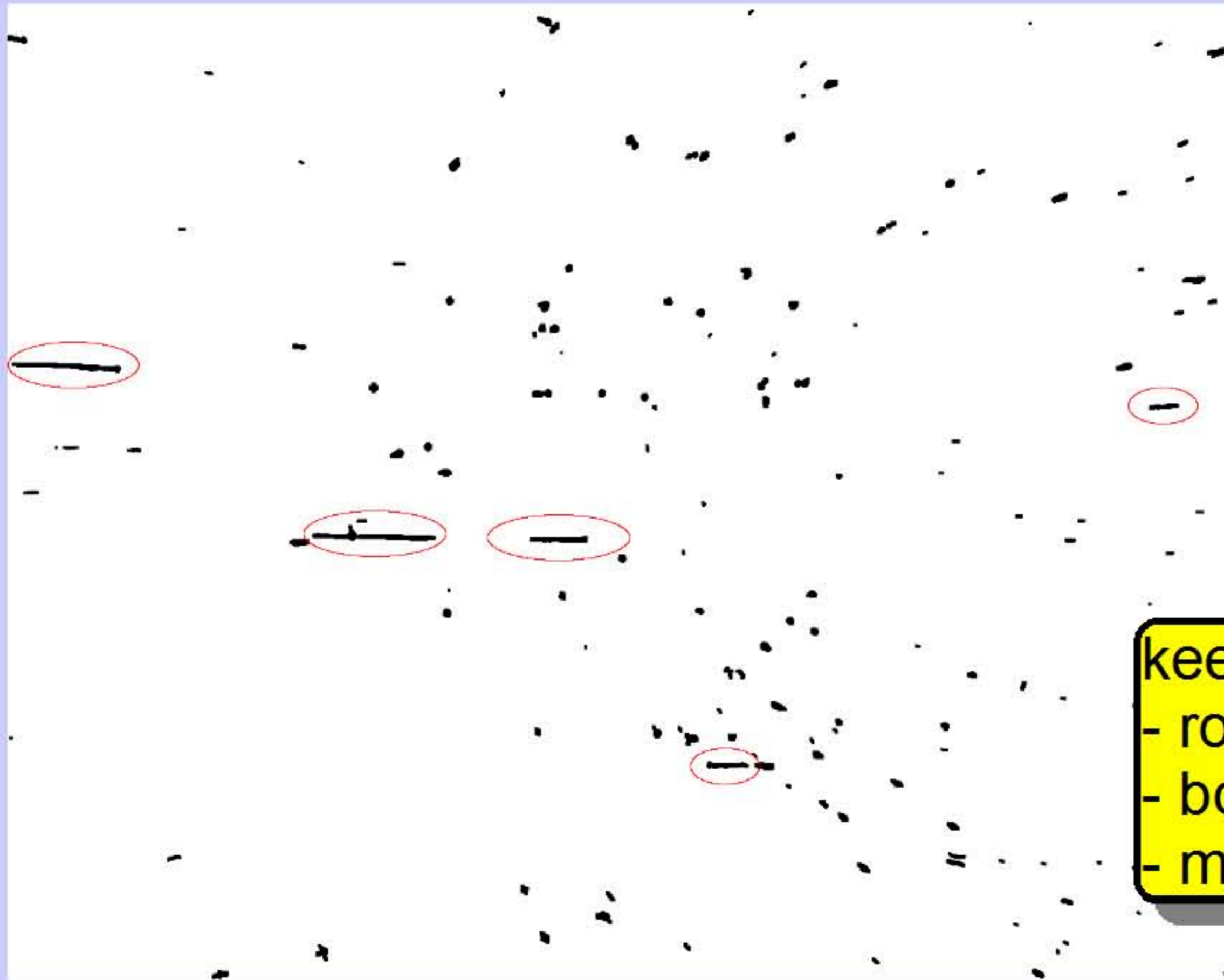


DNA combing



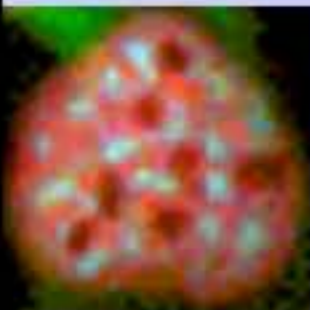


DNA combing



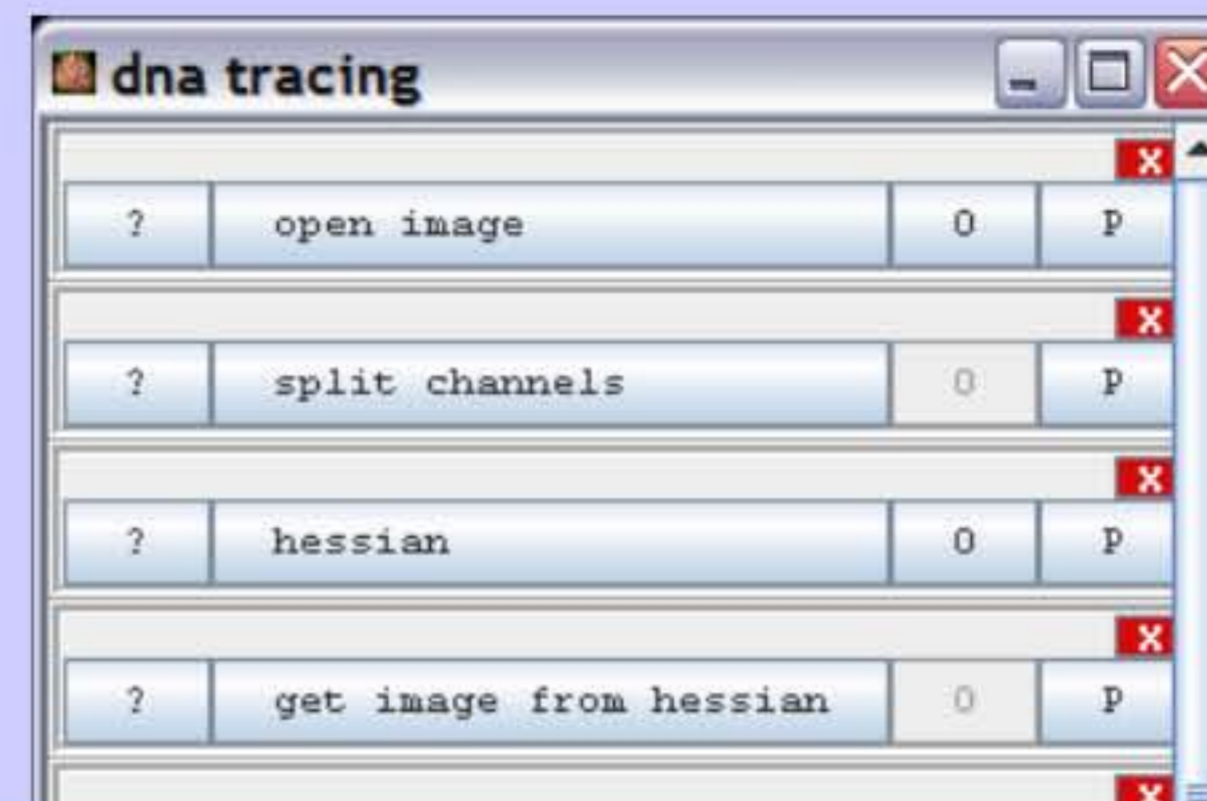
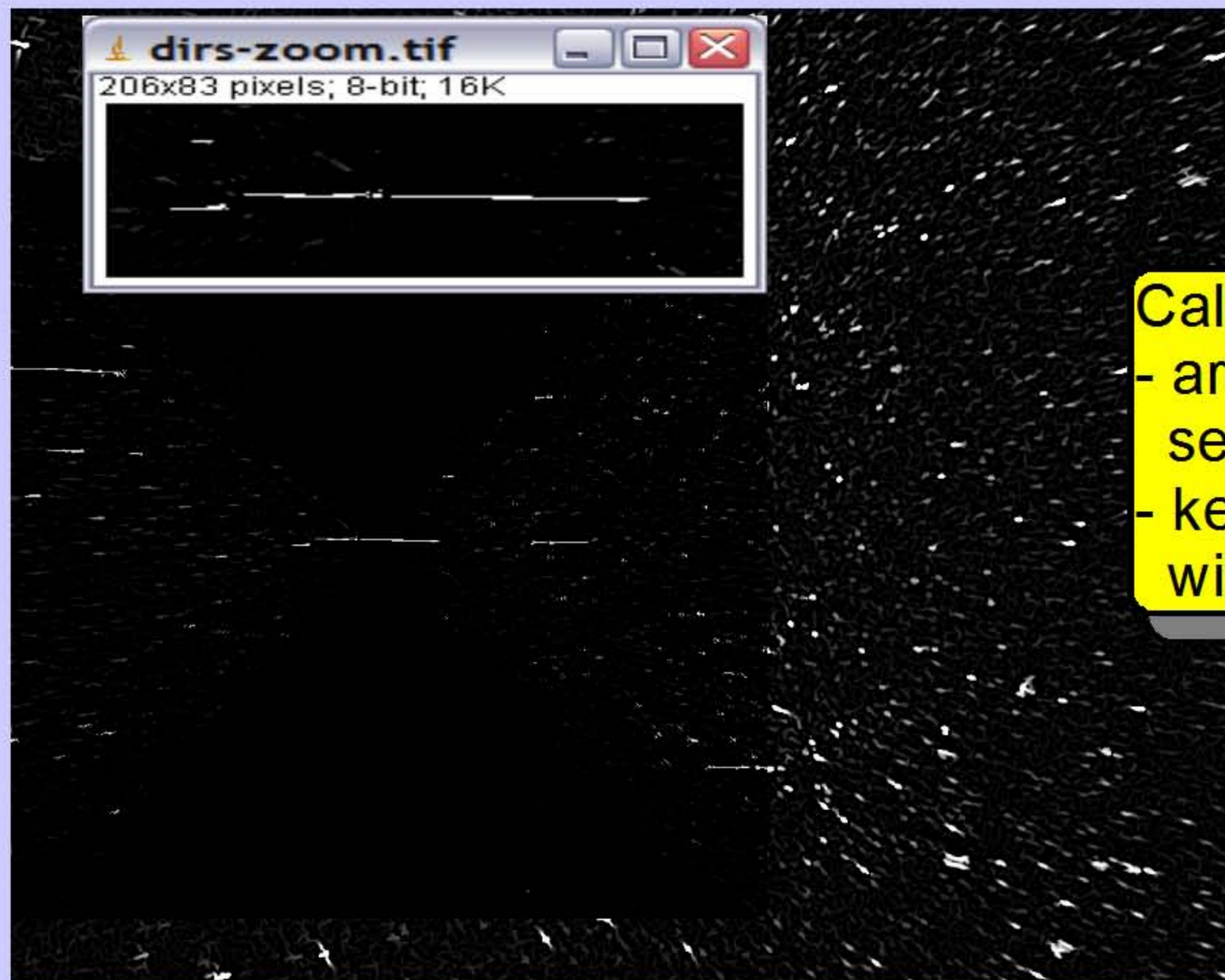
keep only "long" objects

- roundness
- bounding box length/height
- min size

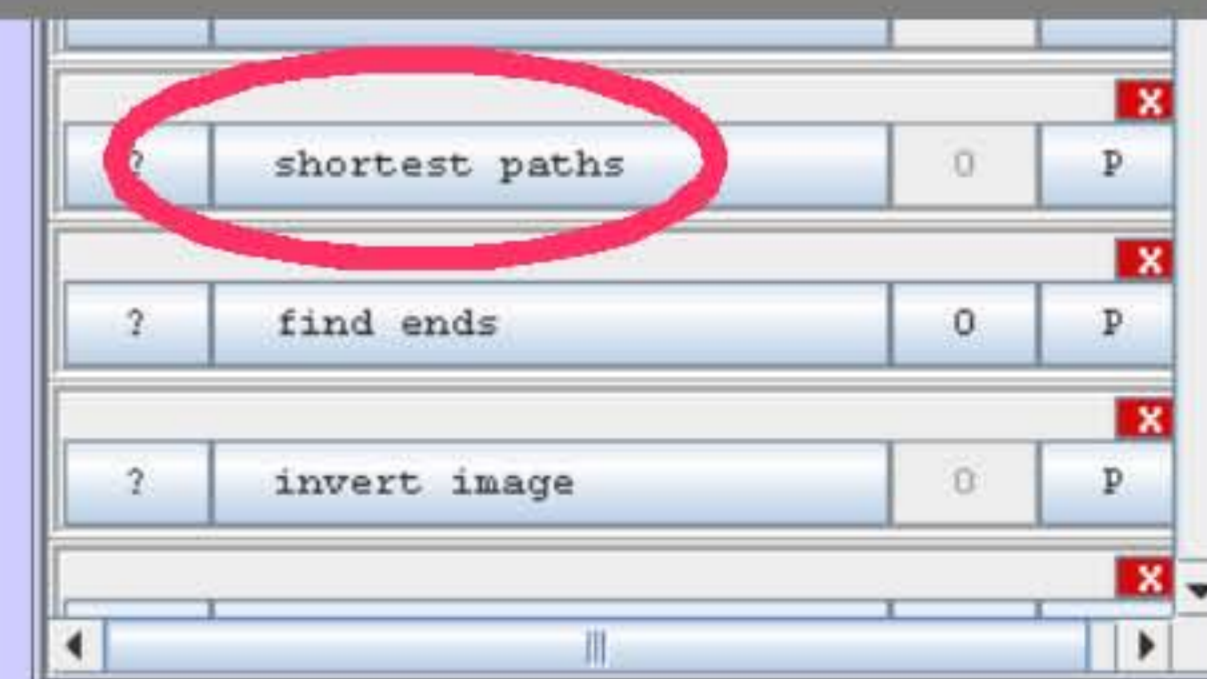




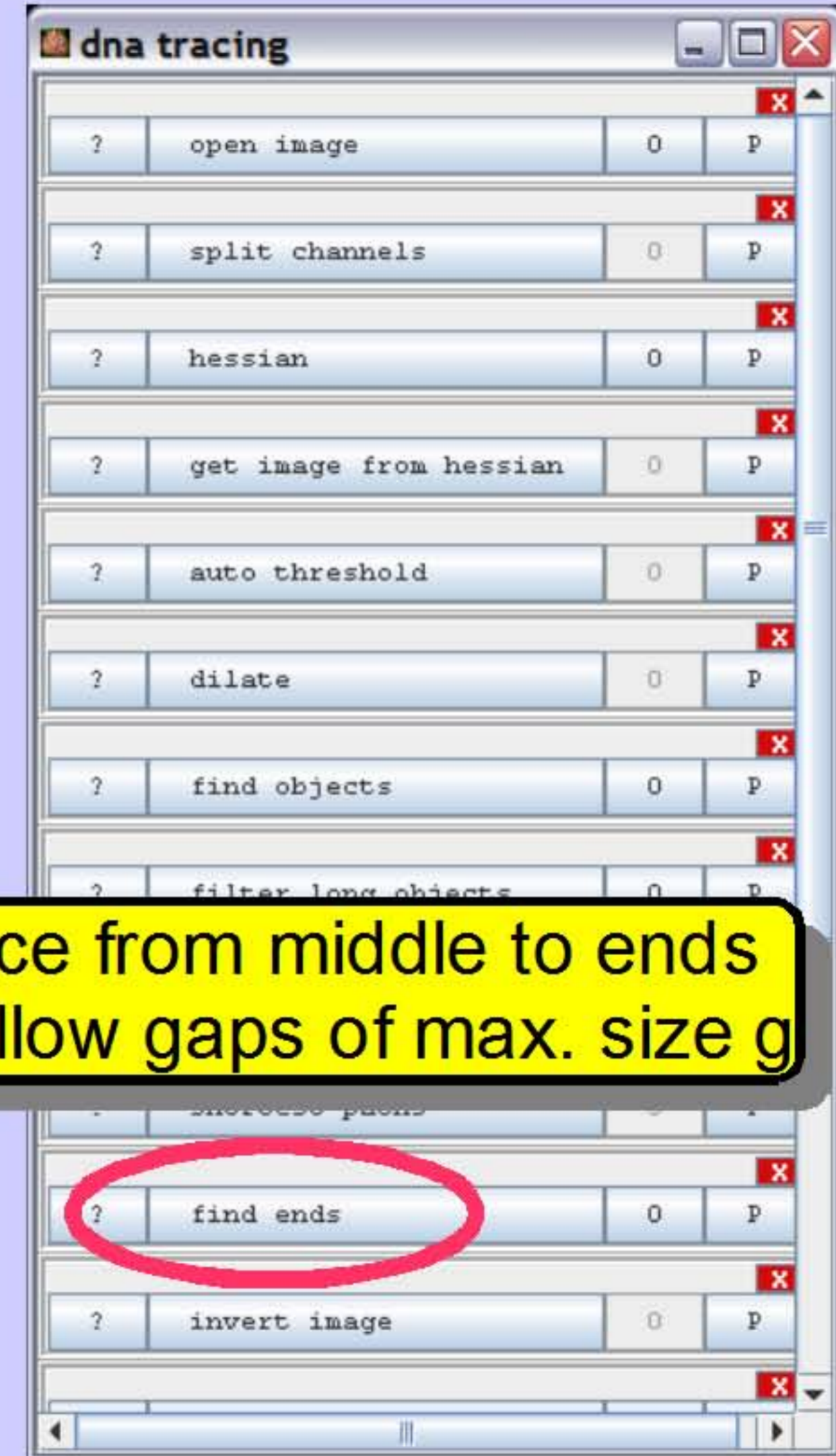
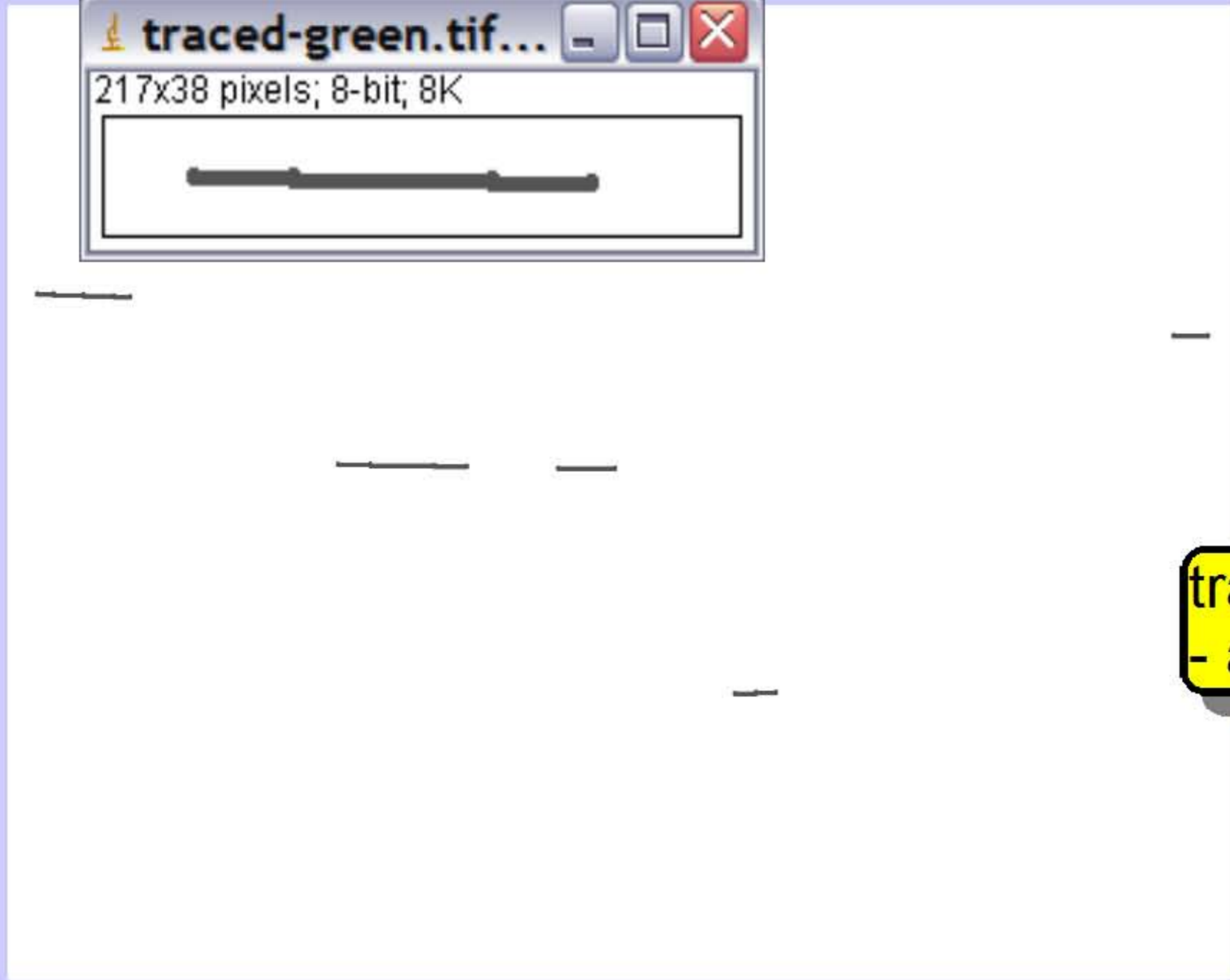
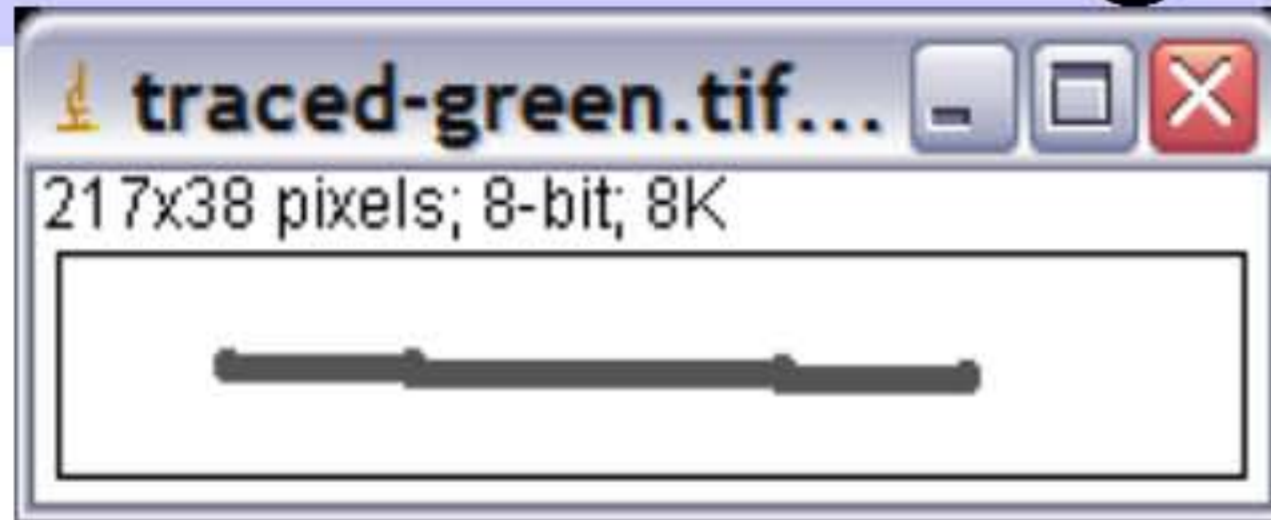
DNA combing



Calculate shortest pathes
- around remaining
segment centers
- keep horizontal steps
with intensity from derivative



DNA combing

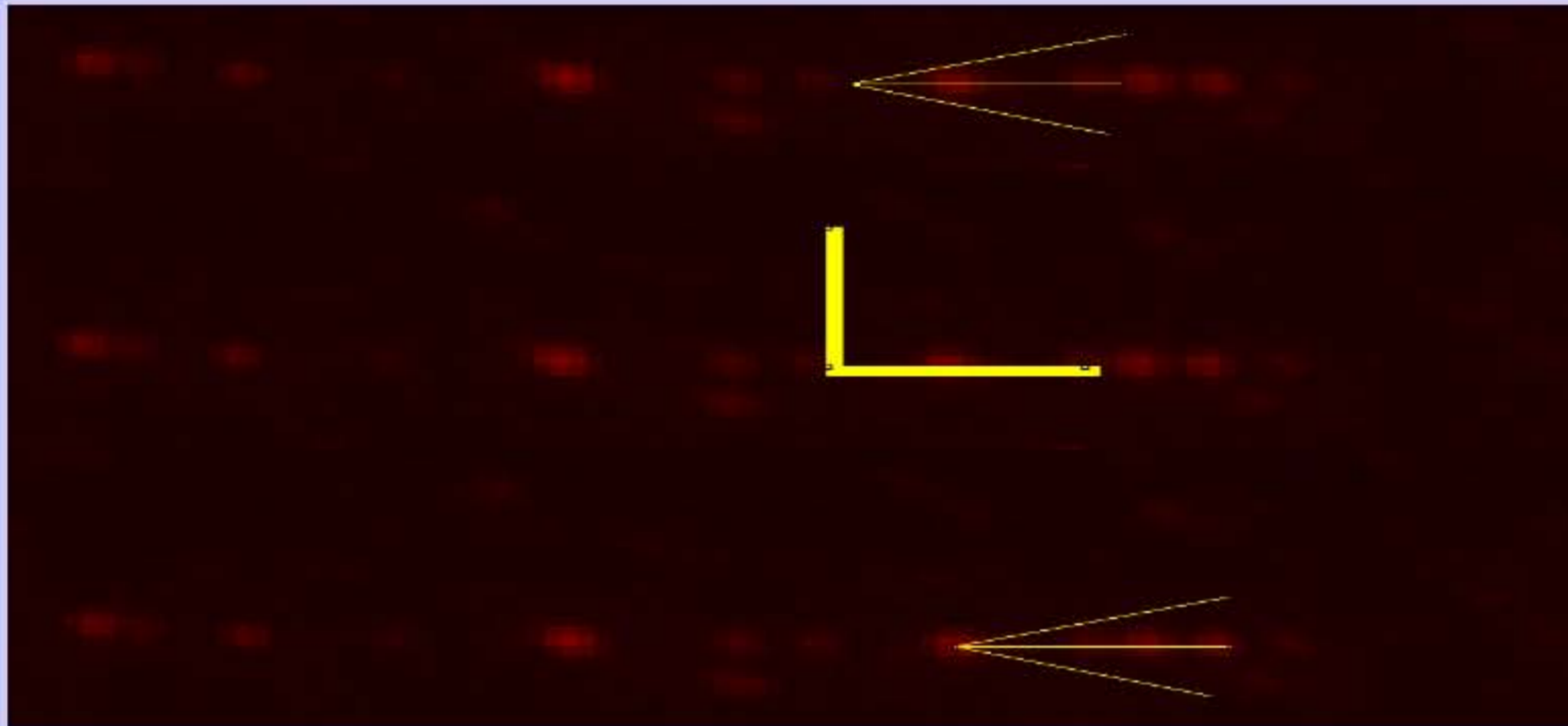


trace from middle to ends
- allow gaps of max. size g

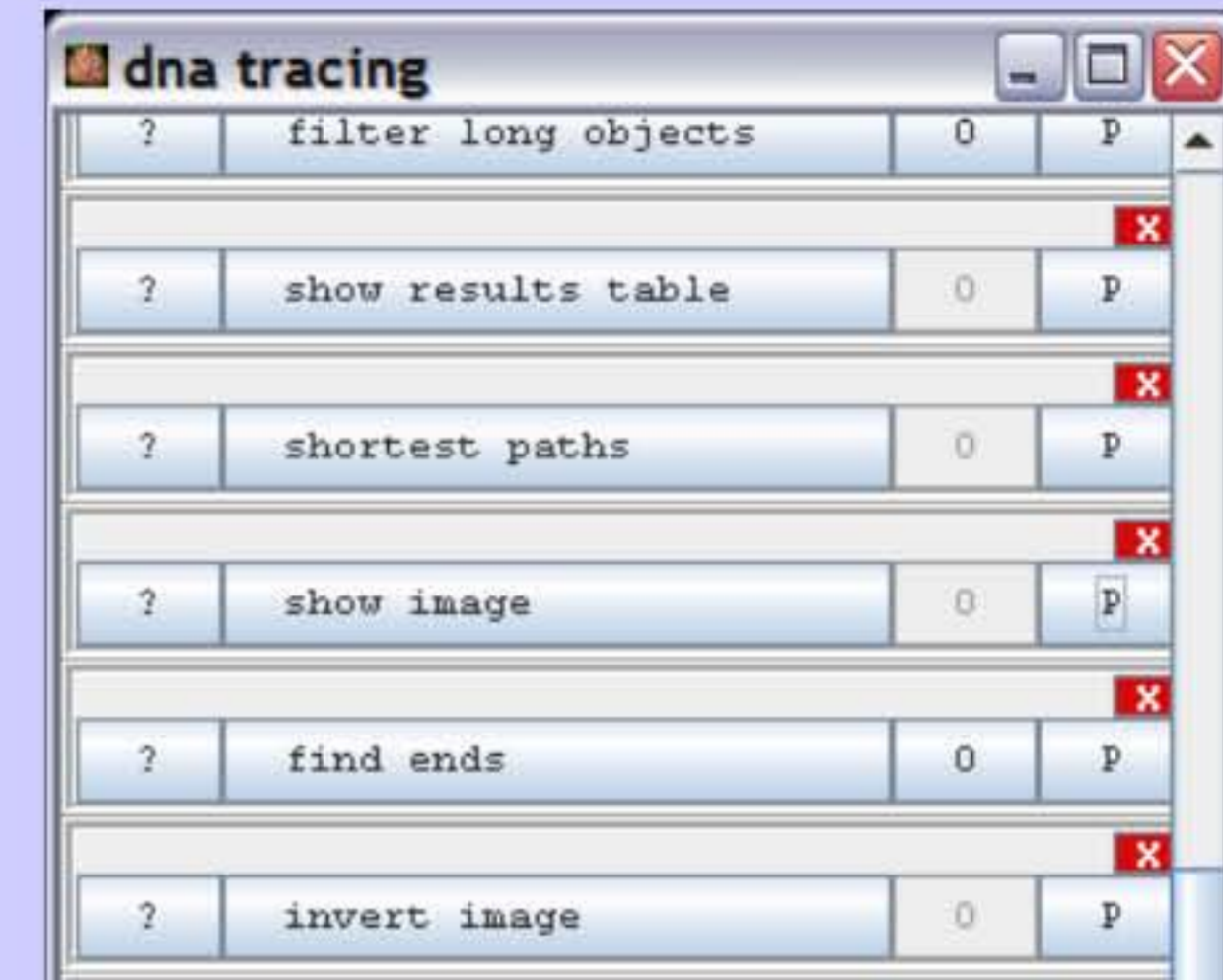
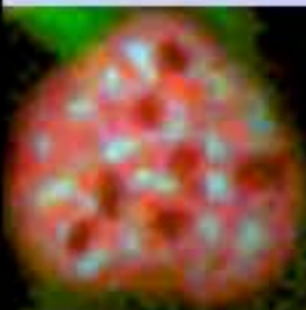




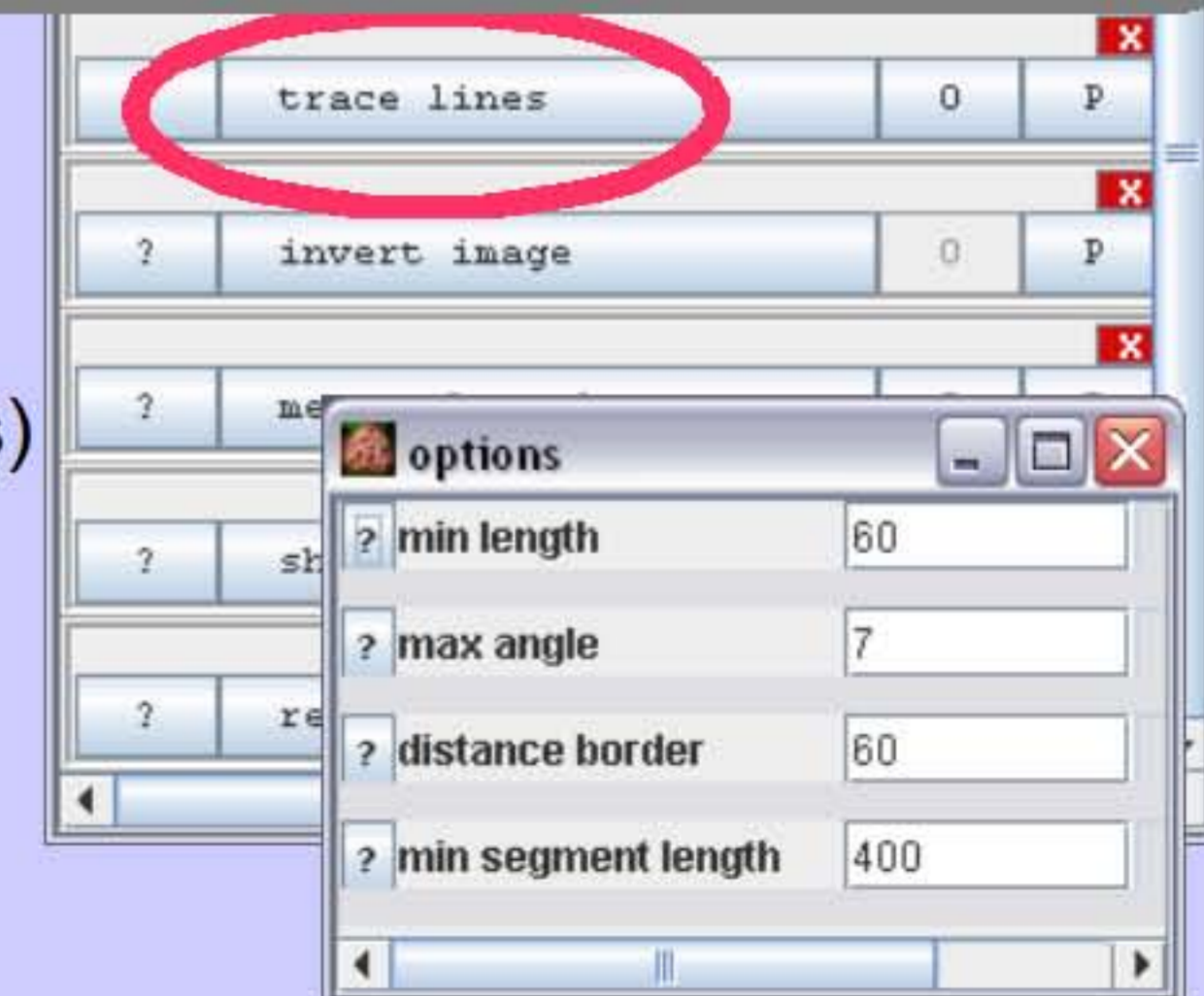
DNA combing



- to find start and end of the molecule (red)
- start in the middle of a green segment
 - find the best direction to go
(highest average intensity for a line segment of size s)
 - move one pixel in that direction, if intensity in a line segment in that direction is higher than in the perpendicular direction
 - else stop



trace lines in red signal
- from centers of green lines



DNA combing

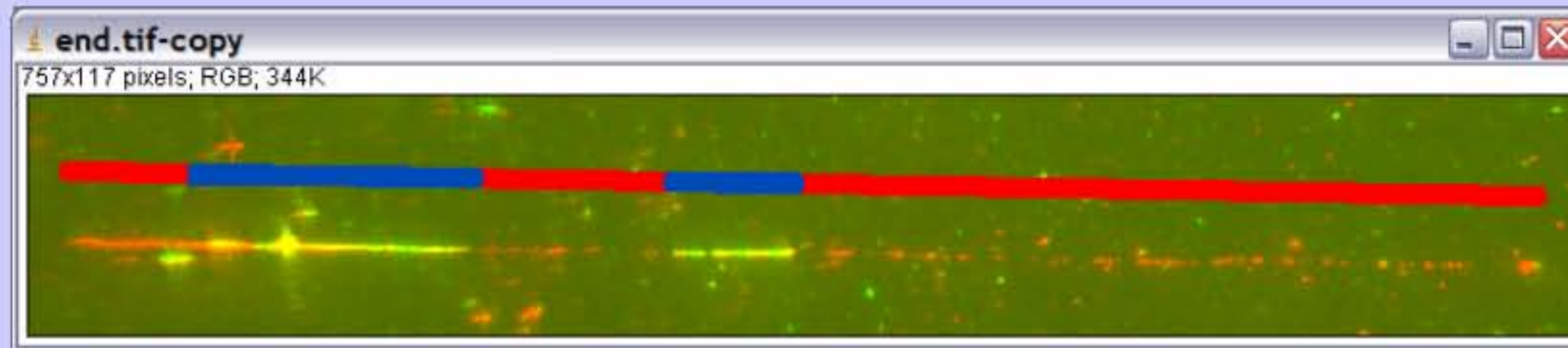


image	na brin	start x	start y	end x	end y	total length	folder
Bt5_1_06.tif	1	254,86	559,48	973,72	570,55	718,95	Z:\baecker\combing\
		1		2			
		318.0, 560.0, 453.0, 562.0		551.0, 564.0, 610.0, 564.0			
	red	green	red	green	red		
	63,14	135,01	98,02	59	363,78		
	1-2	2-3	3-4	4-5			
	195,02						

Report:

- total length
- length of each segment
- distances between centers of green segments





DNA combing, manual

use **slide show control** to select image

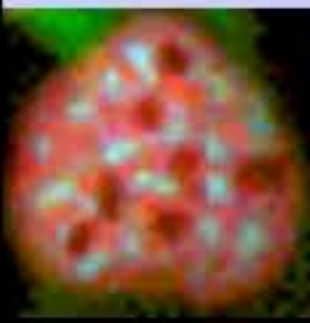
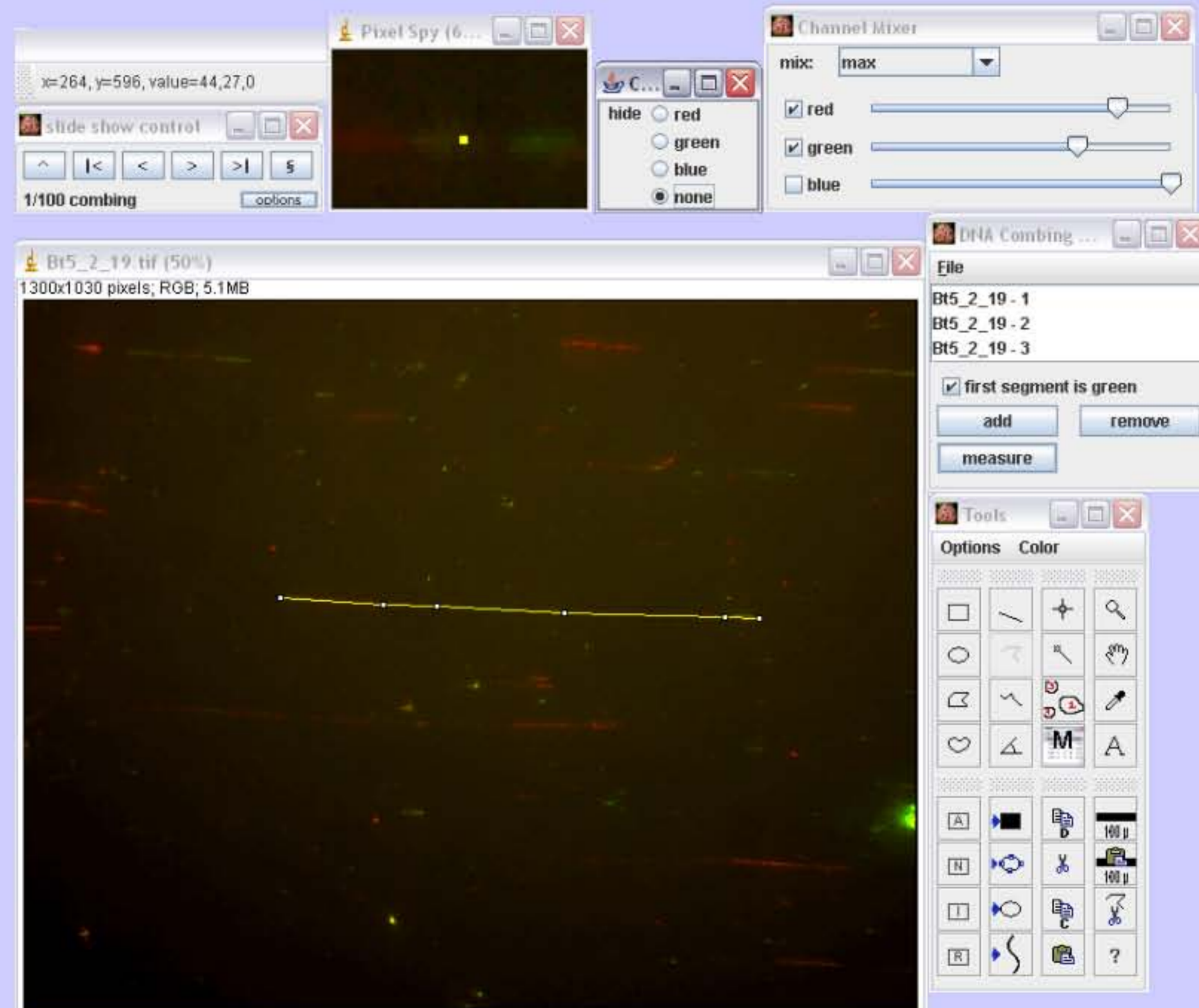
use **channel mixer** to adjust view

use **pixel spy** to see exactly where you set marks

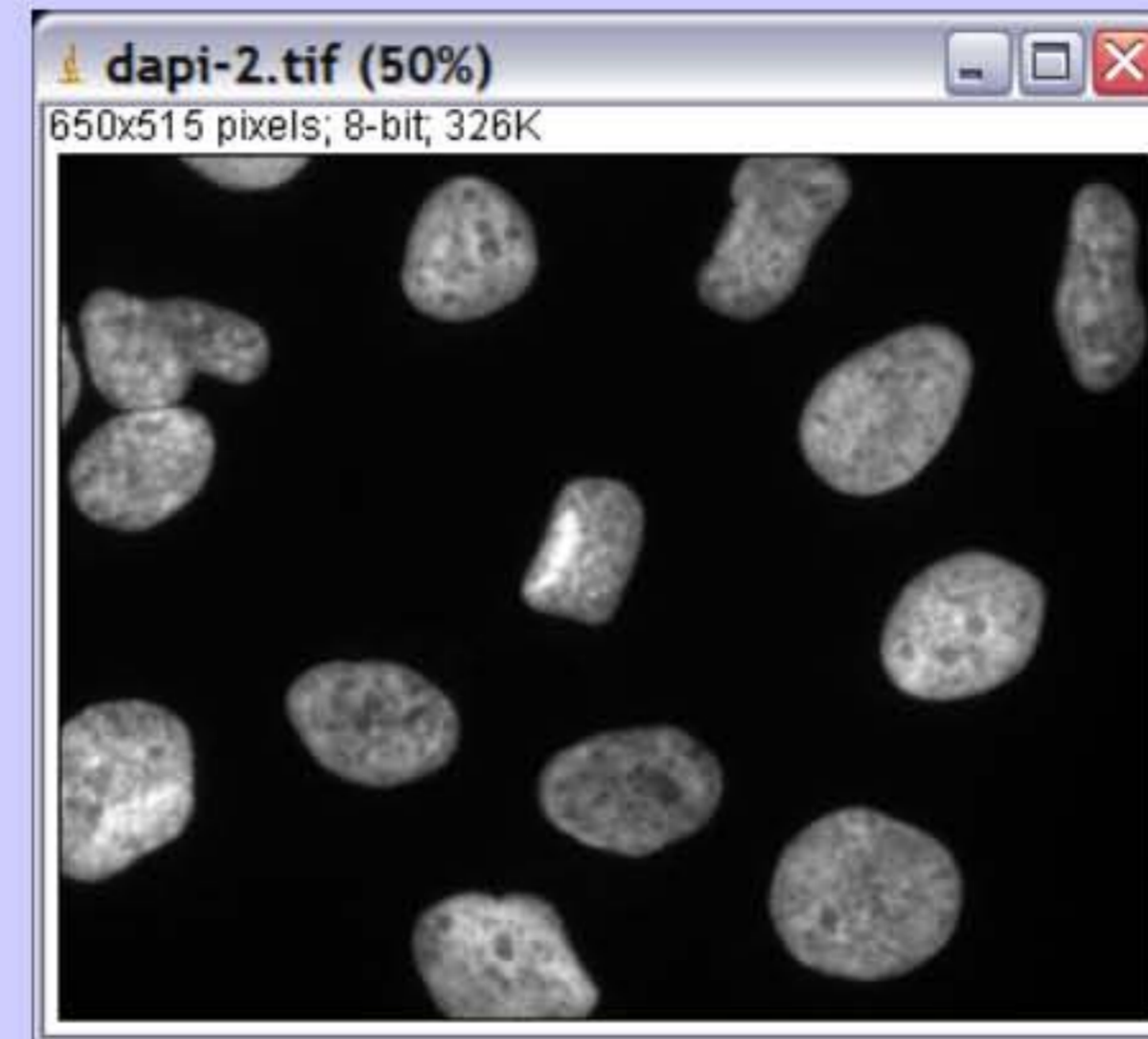
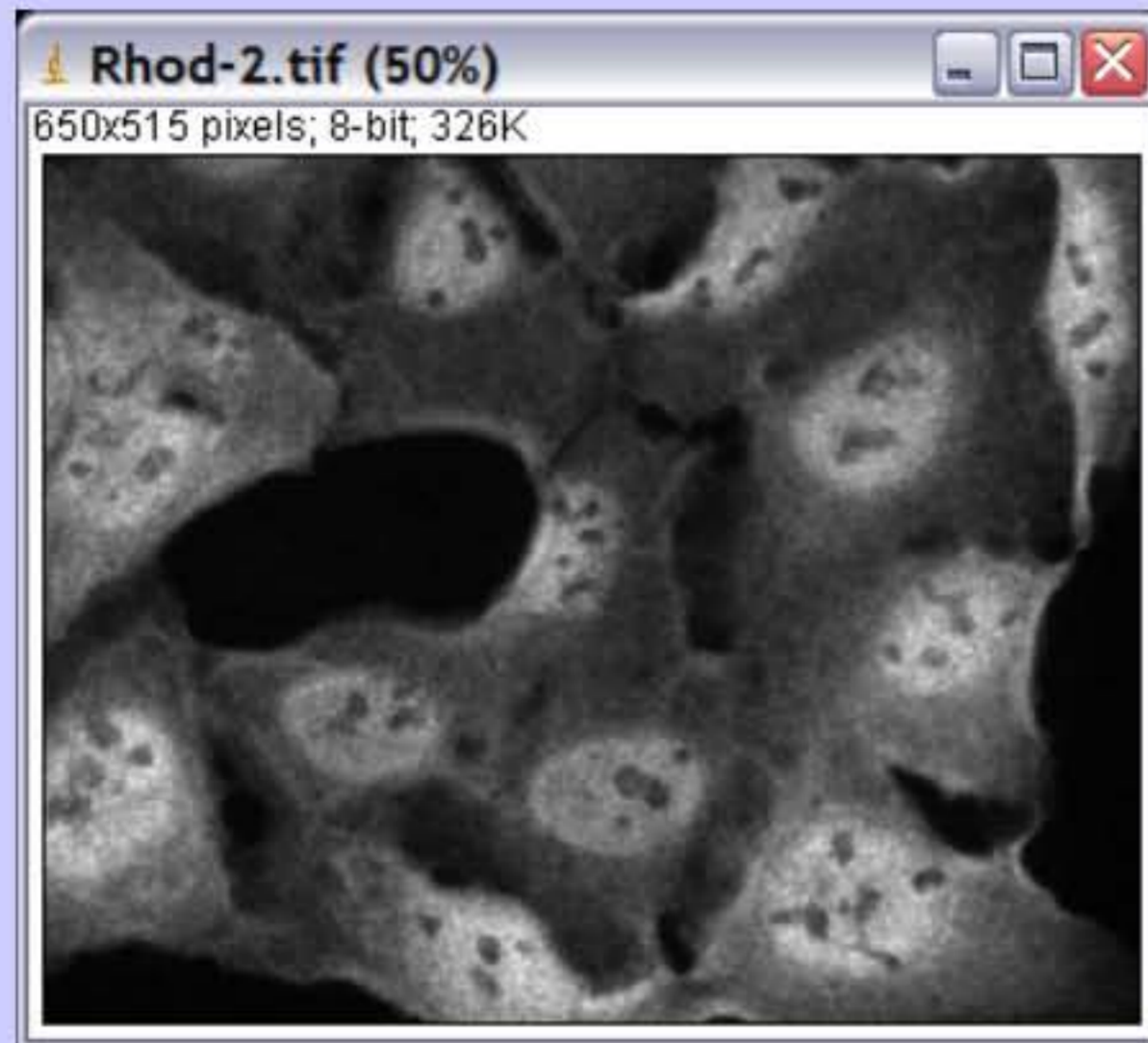
use **polygon selection tool** to mark red and green segments

use **DNA combing tool** to save / load selections and to create reports

reports have the same format as in the automatic application

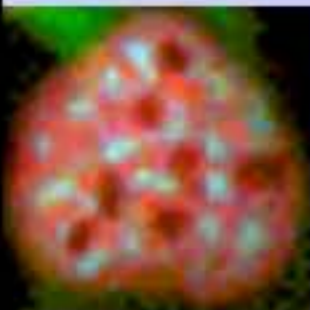


Measuring intensity ratios



what is the proportion of fluorescence between nuclei and cytoplasm in the first image?

the second image is used to identify the nuclei





Measuring intensity ratios

The screenshot displays two windows from the ImageJ software. The 'Ouvrir' (Open) window is on the left, showing a search for 'testdata' in the 'coux' folder. The 'list editor' window is on the right, showing a list of image files with a 'rhod' filter and a 'select' button.

Ouvrir

Rechercher dans :

testdata

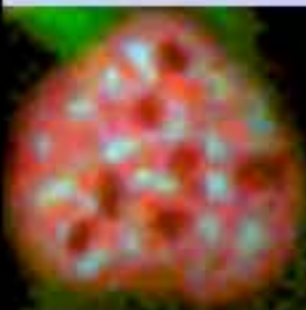
Nom de fichier :

Fichiers du type :

list editor

Z:\baecker\coux\testdata\U2OS A4 05-05-05 copie\A4 dapi 2.tif
Z:\baecker\coux\testdata\U2OS A4 05-05-05 copie\A4 dapi 3.tif
Z:\baecker\coux\testdata\U2OS A4 05-05-05 copie\A4 dapi 4.tif
Z:\baecker\coux\testdata\U2OS A6 05-05-05 copie\dapi-2.tif
Z:\baecker\coux\testdata\U2OS A6 05-05-05 copie\dapi-3.tif
Z:\baecker\coux\testdata\U2OS A6 05-05-05 copie\dapi-4.tif
Z:\baecker\coux\testdata\U2OS A6 05-05-05 copie\dapi-1.tif
Z:\baecker\coux\testdata\U2OS A7 05-05-05 copie\dapi 1.tif
Z:\baecker\coux\testdata\U2OS A7 05-05-05 copie\dapi 2.tif
Z:\baecker\coux\testdata\U2OS A7 05-05-05 copie\dapi 3.tif
Z:\baecker\coux\testdata\U2OS A7 05-05-05 copie\dapi 4.tif
Z:\baecker\coux\testdata\U2OS A7 05-05-05 copie\dapi 5.tif
Z:\baecker\coux\testdata\U2OS A7 05-05-05 copie\rhod 1.tif
Z:\baecker\coux\testdata\U2OS A7 05-05-05 copie\rhod 2.tif

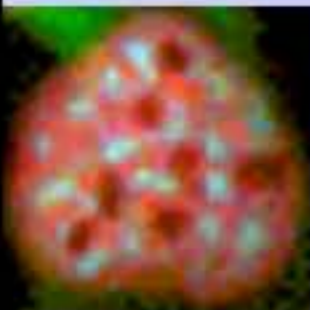
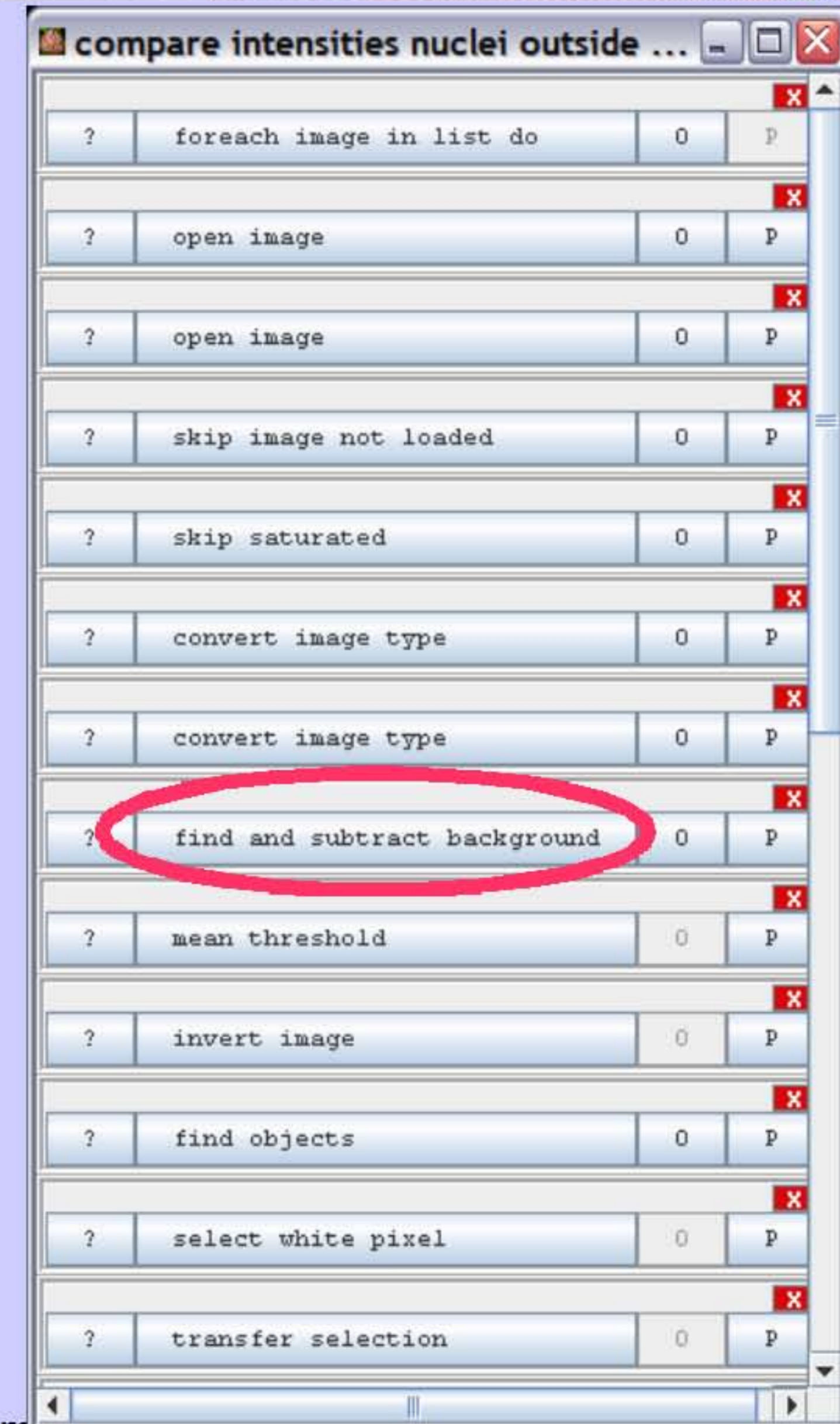
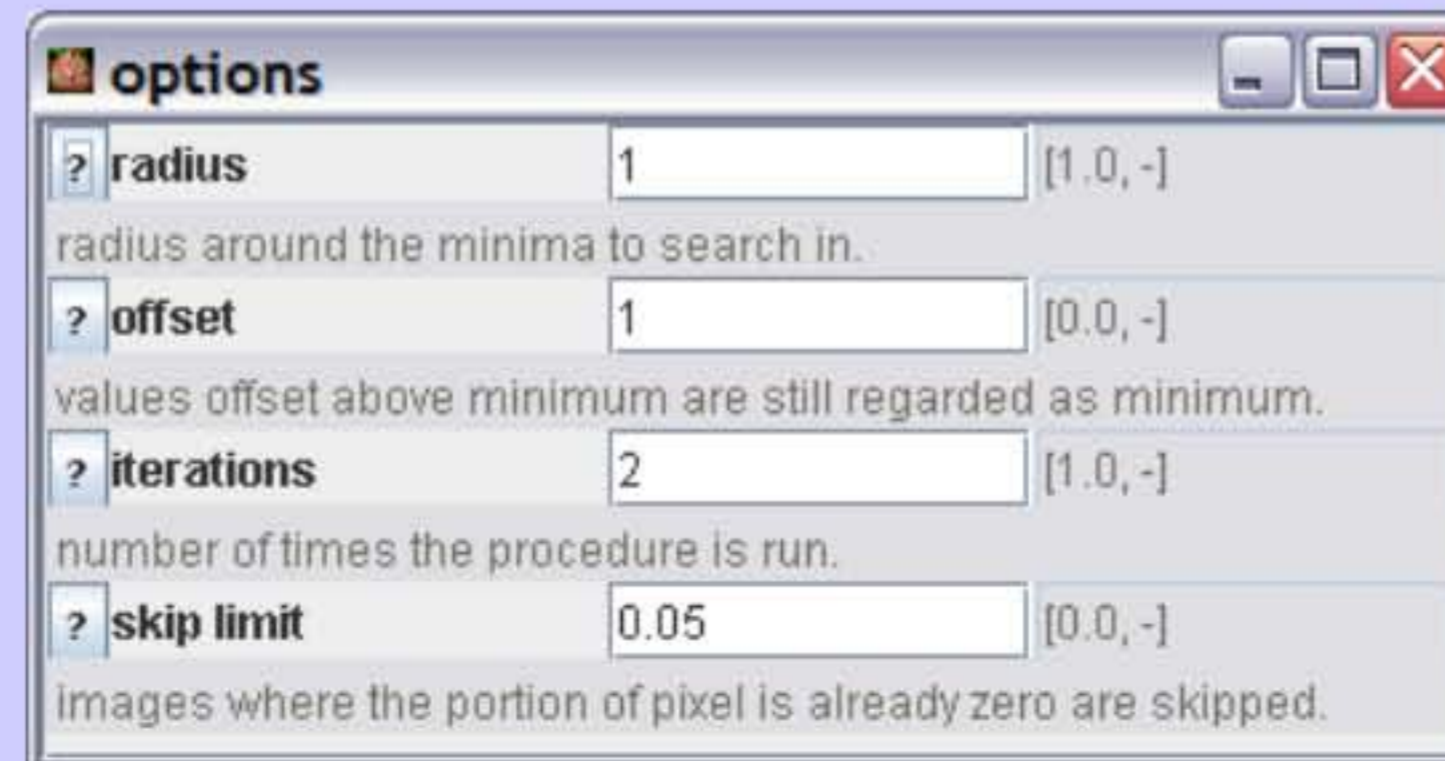
- click « add » on the list editor and select a folder
 - all images in all sub-folders are added to list
 - use list editor to remove unwanted images



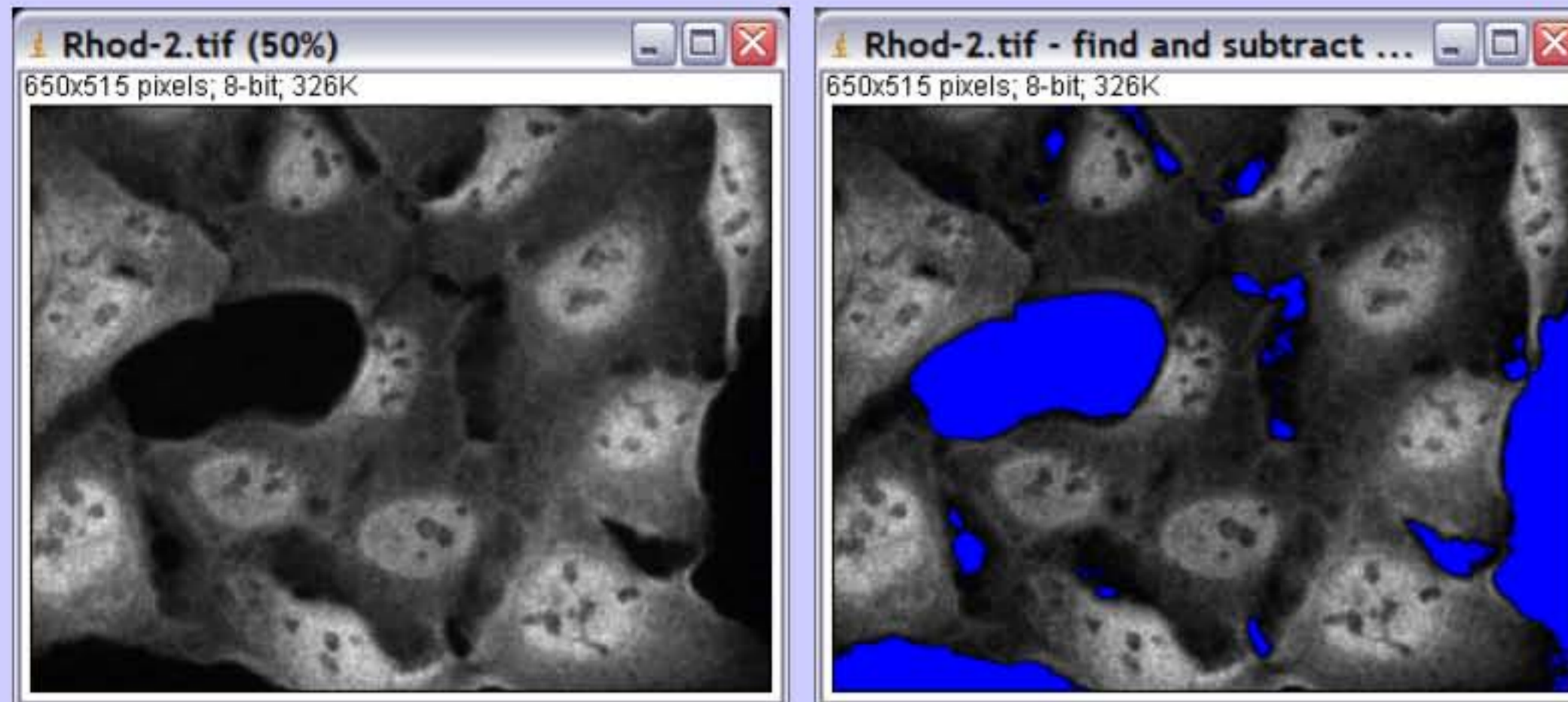


Measuring intensity ratios

- find and subtract background
 - find pixels P with intensity in [min, offset]
 - Find maximum intensity in area of size radius around pixels in P
 - subtract the max value from the image

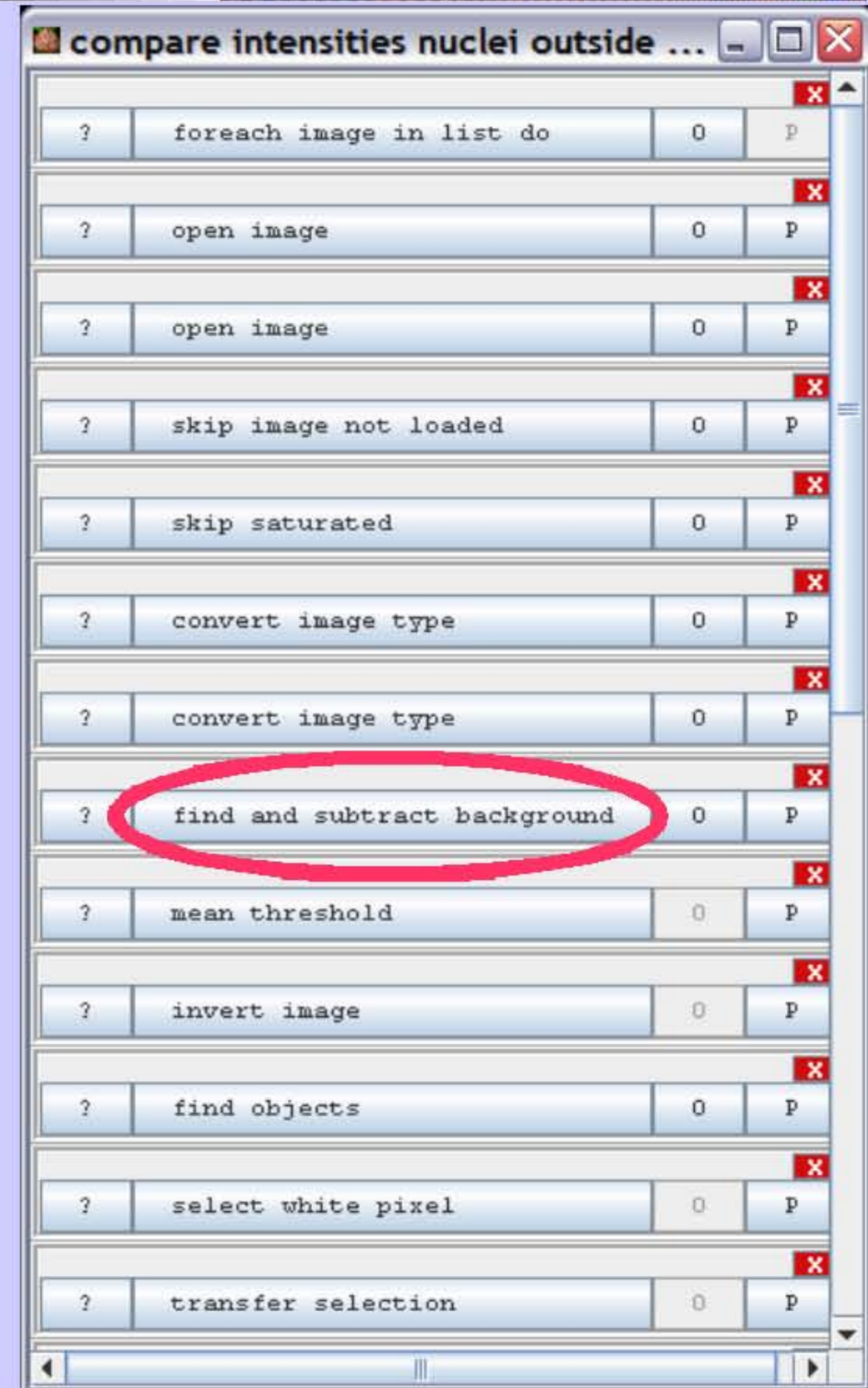


Measuring intensity ratios



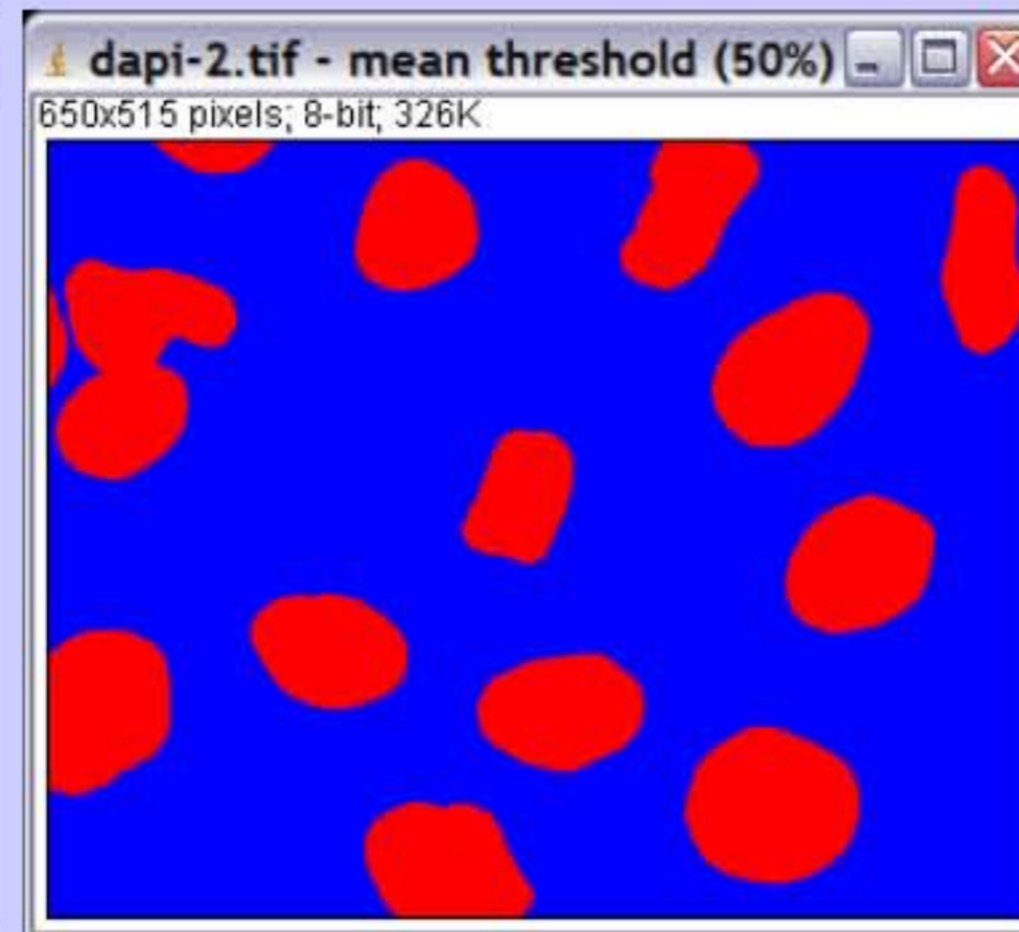
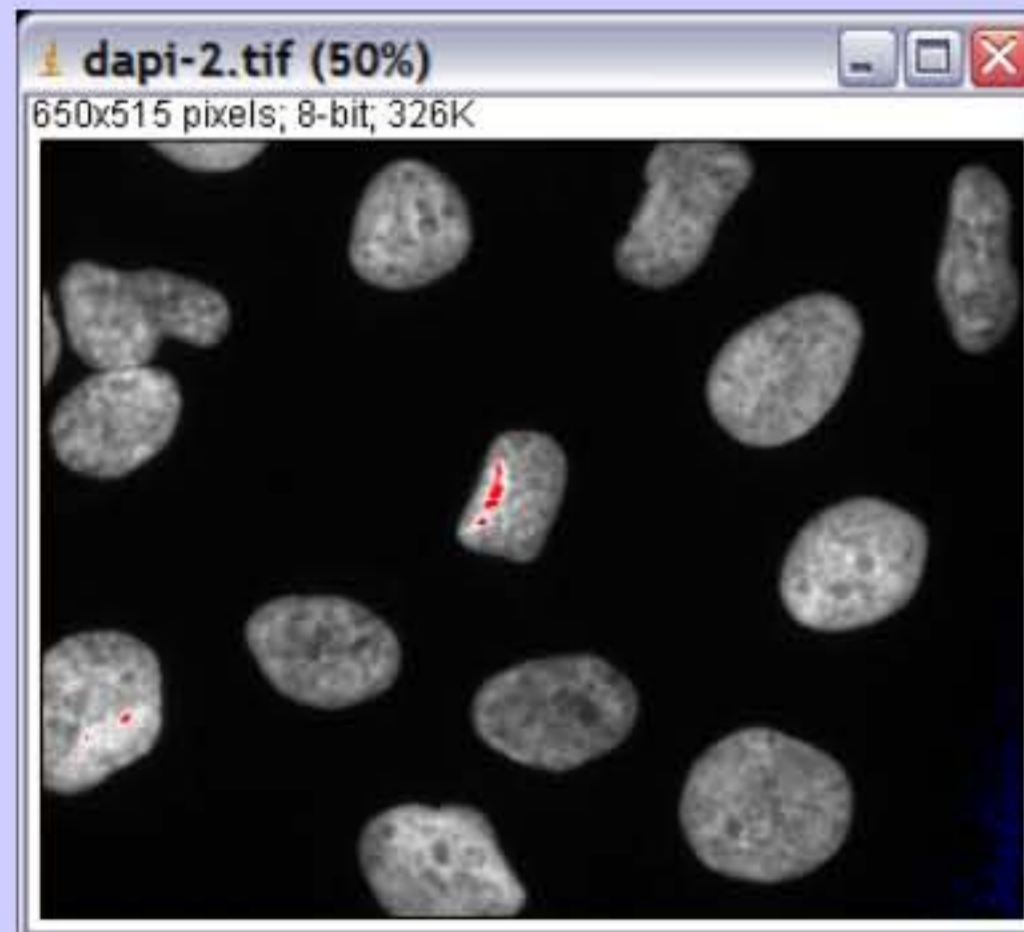
both images with hilo LUT
after background subtraction

blue = intensity is zero

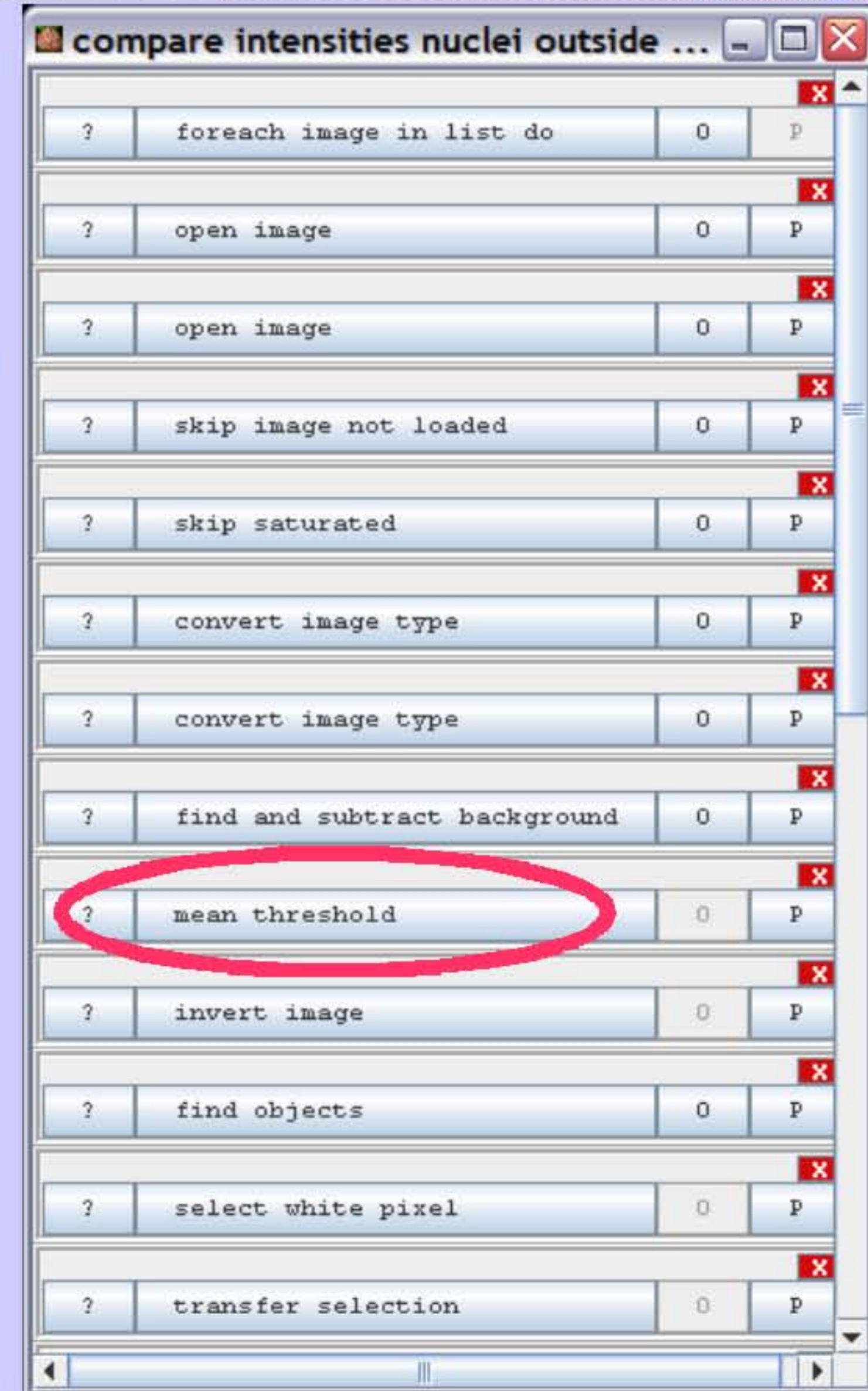




Measuring intensity ratios

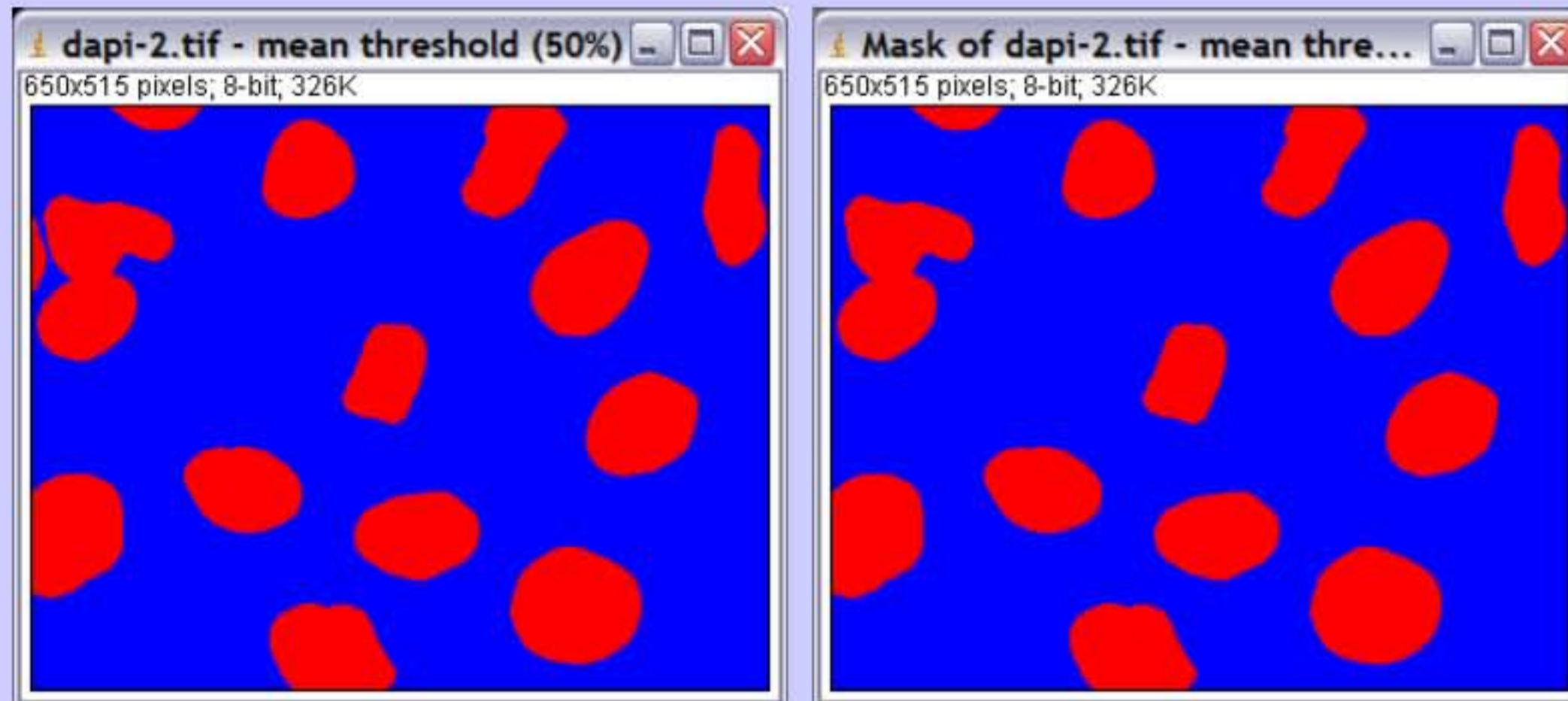


- to find nuclei
- threshold with mean intensity value





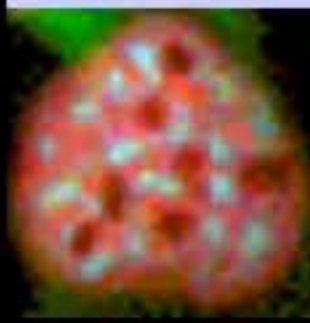
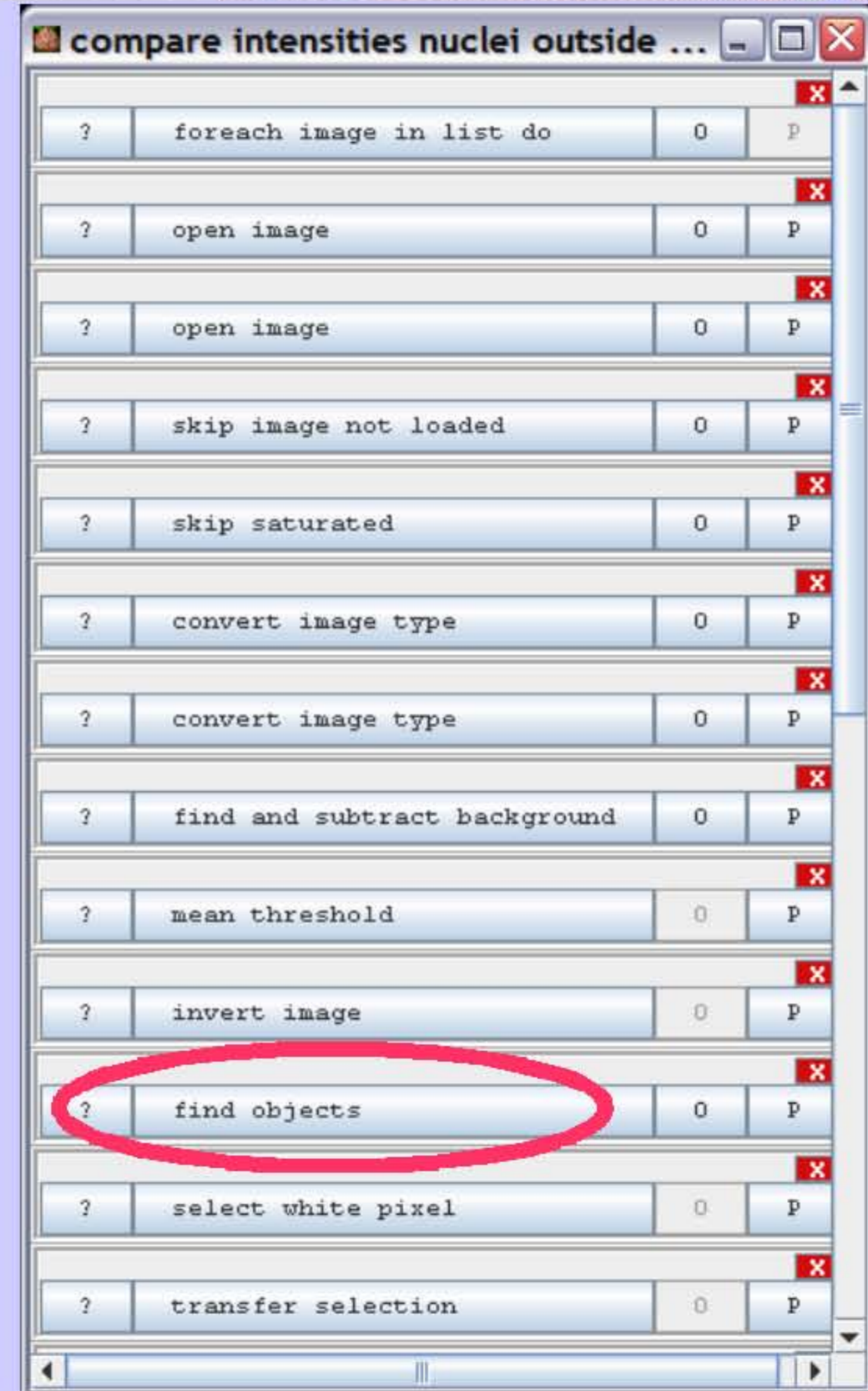
Measuring intensity ratios



find objects (analyze particles)

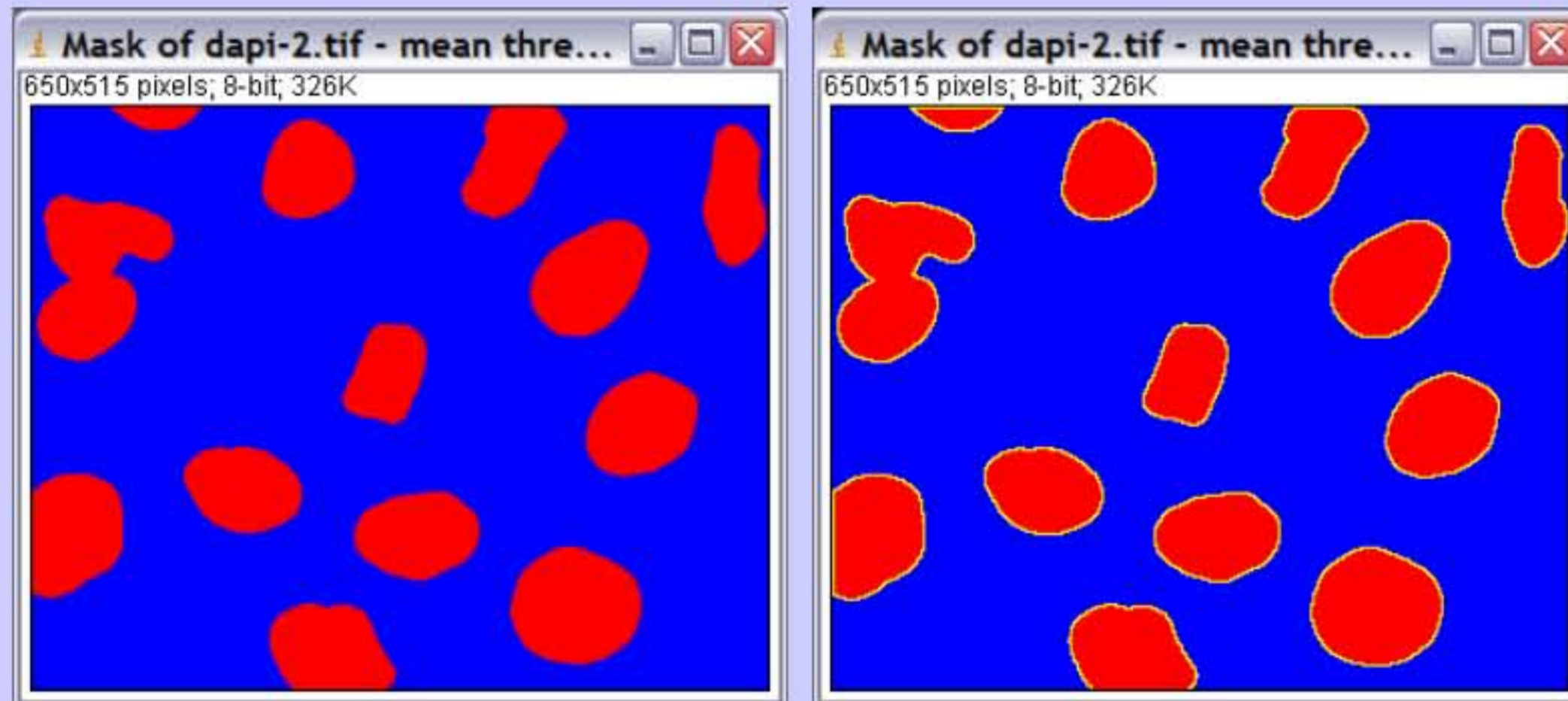
- exclude objects smaller min size

gets rid of objects that are not nuclei

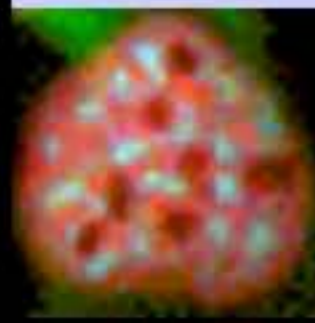
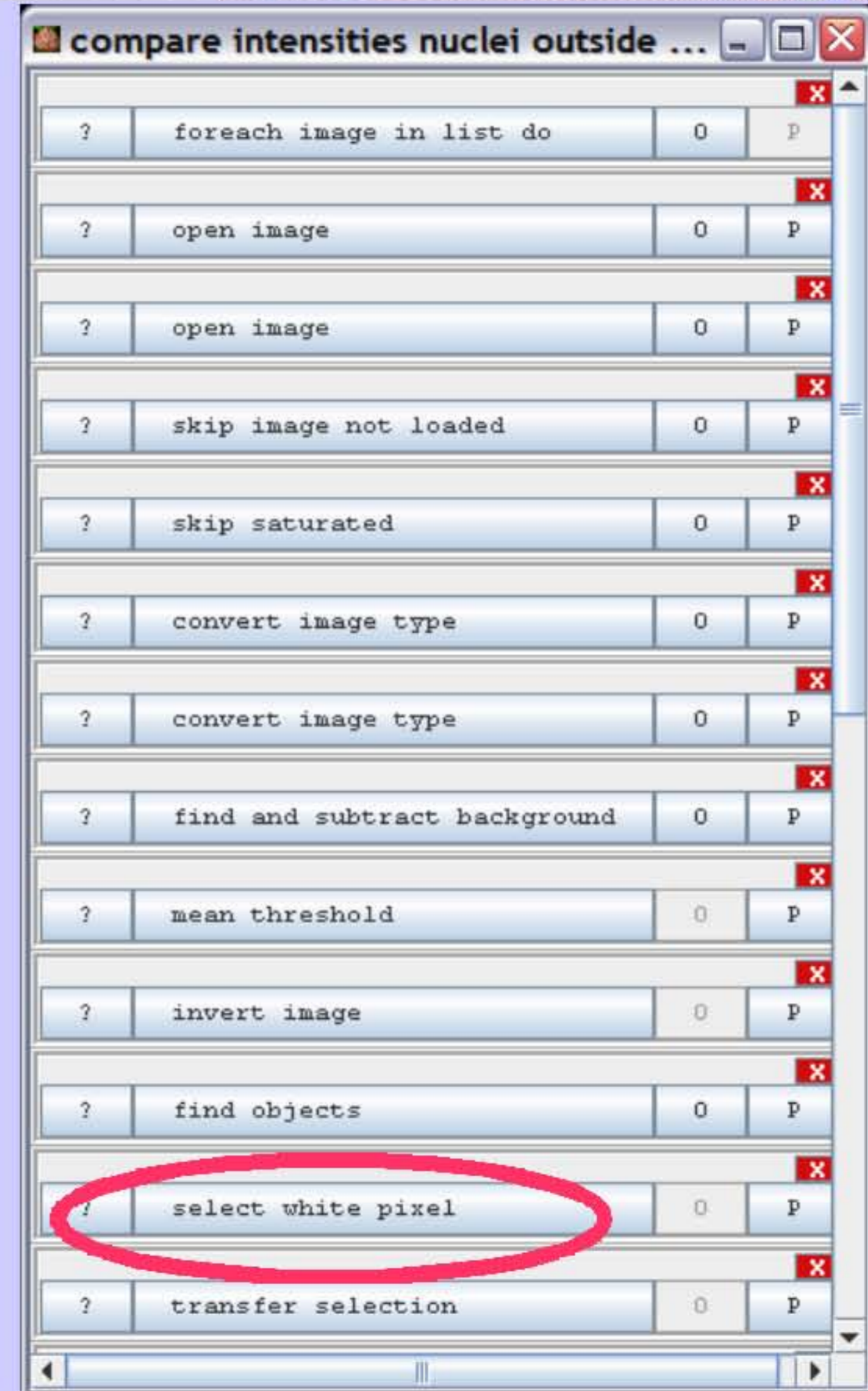




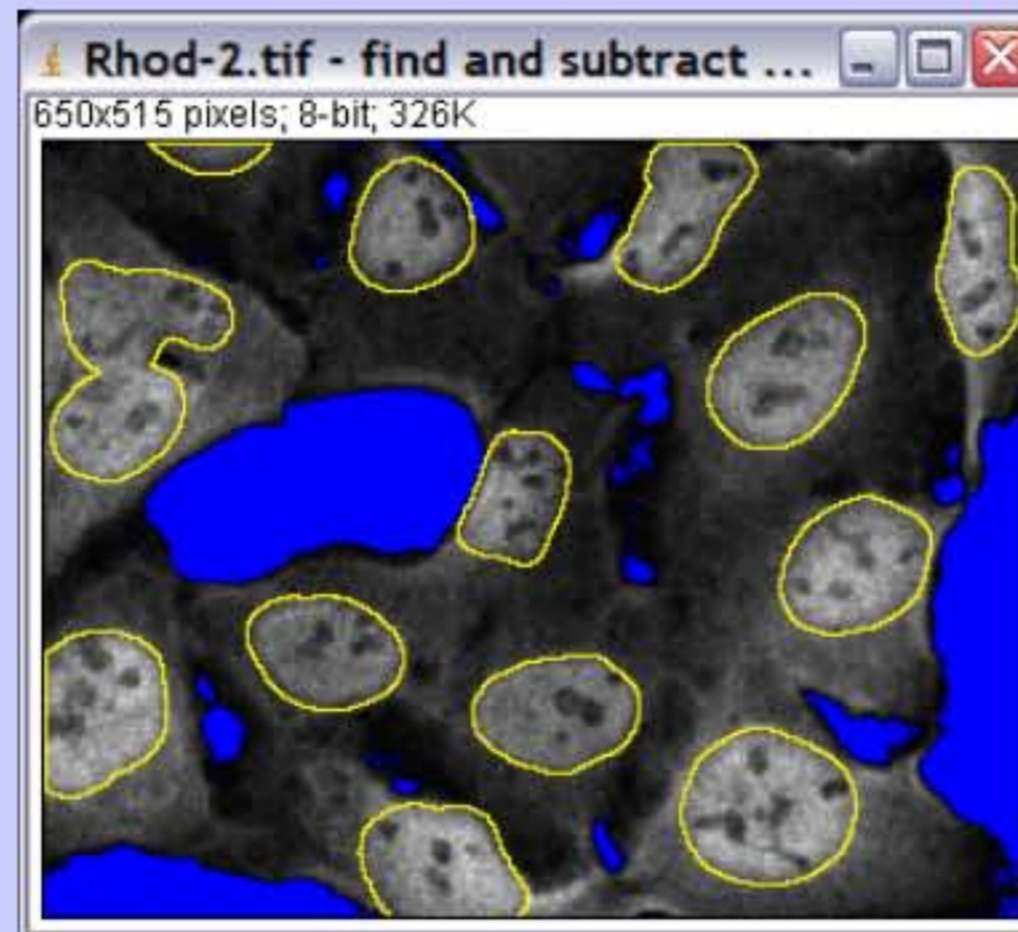
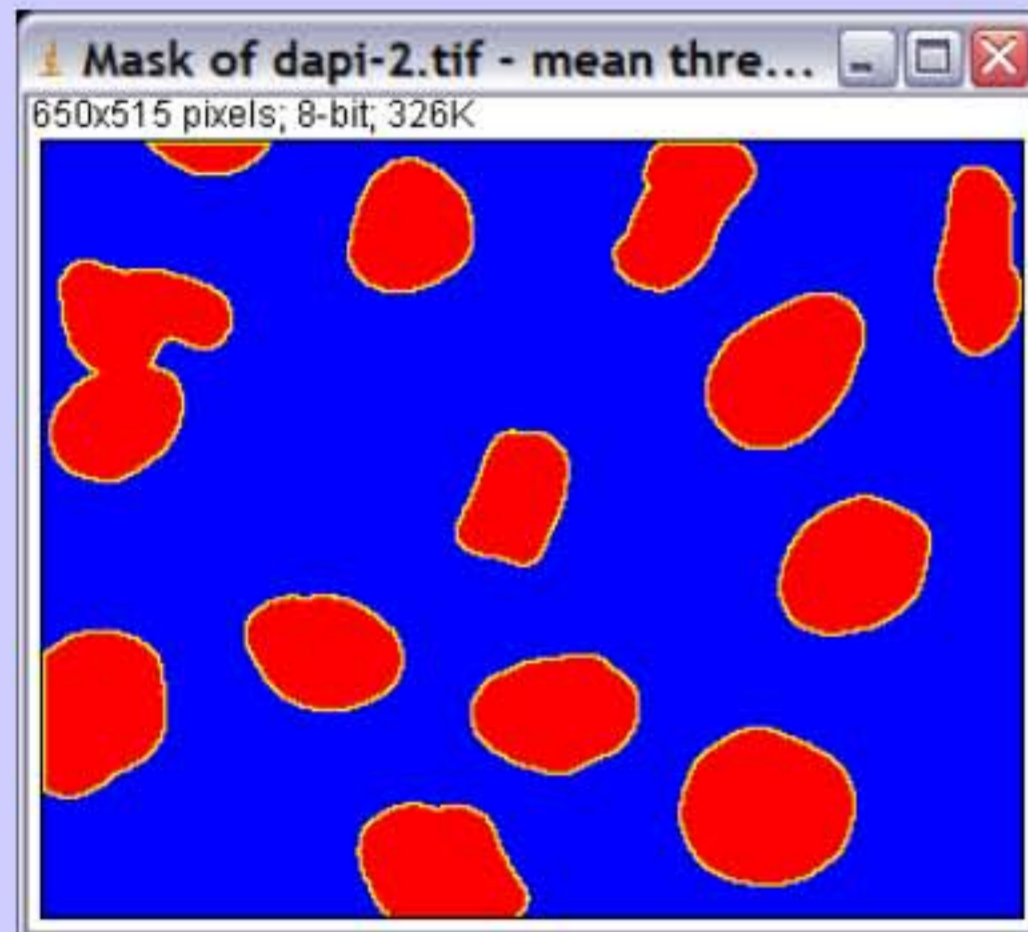
Measuring intensity ratios



create selection (roi) from mask



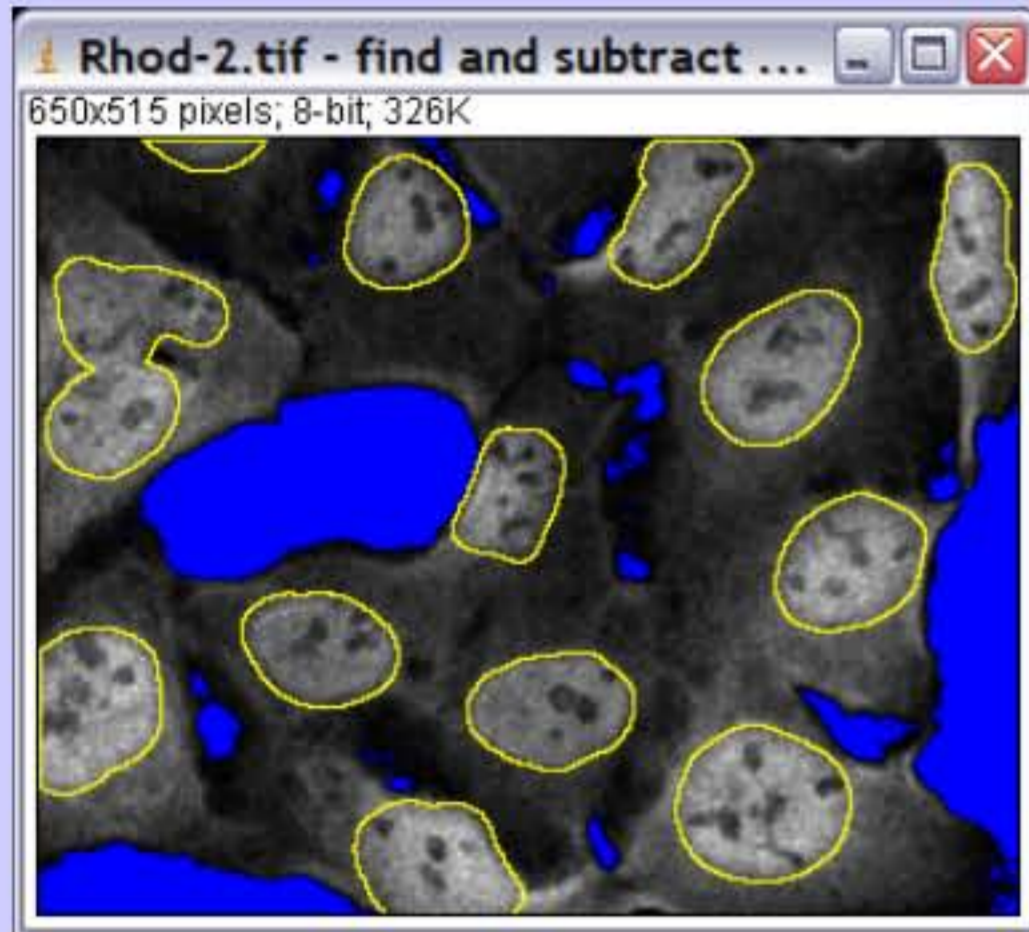
Measuring intensity ratios



transfer selection to background
corrected input image



Measuring intensity ratios



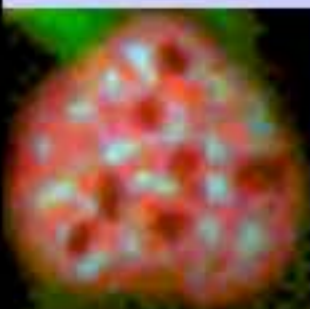
Results

	Area	IntDen
1	84203	9347903



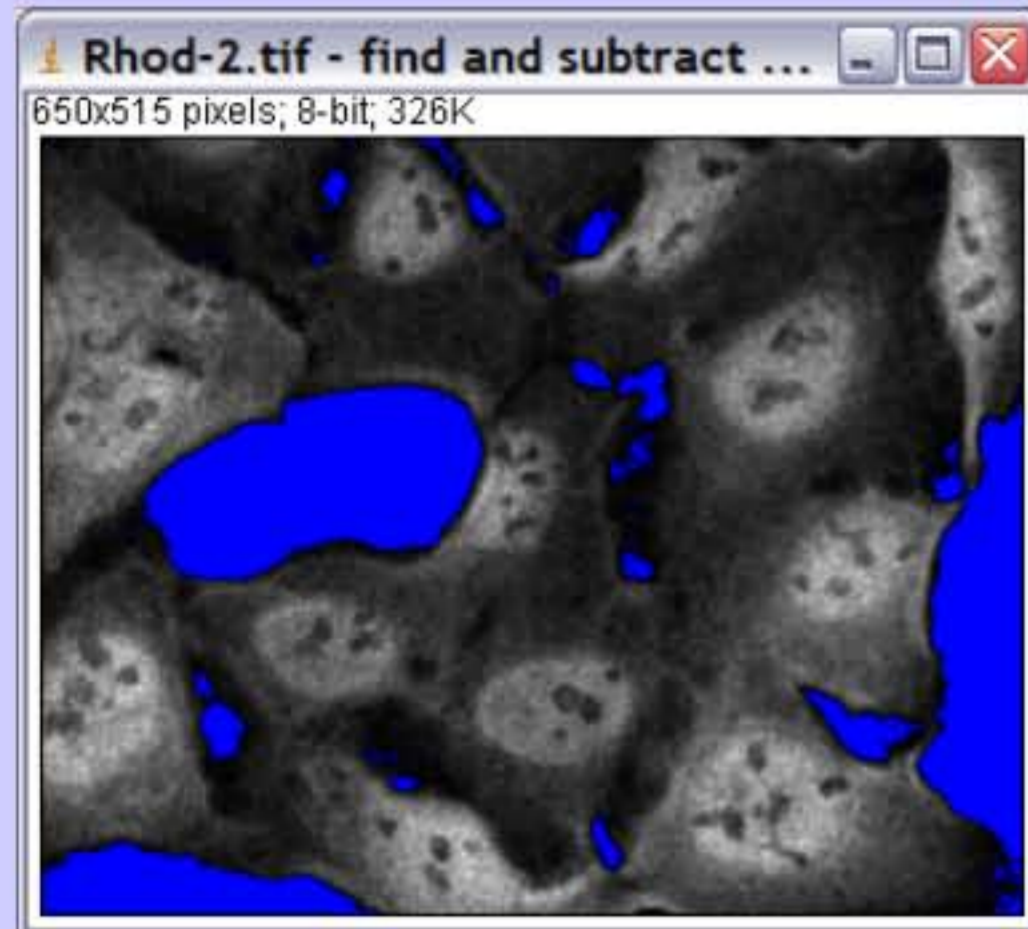
measure

- area
- total intensity of nuclei

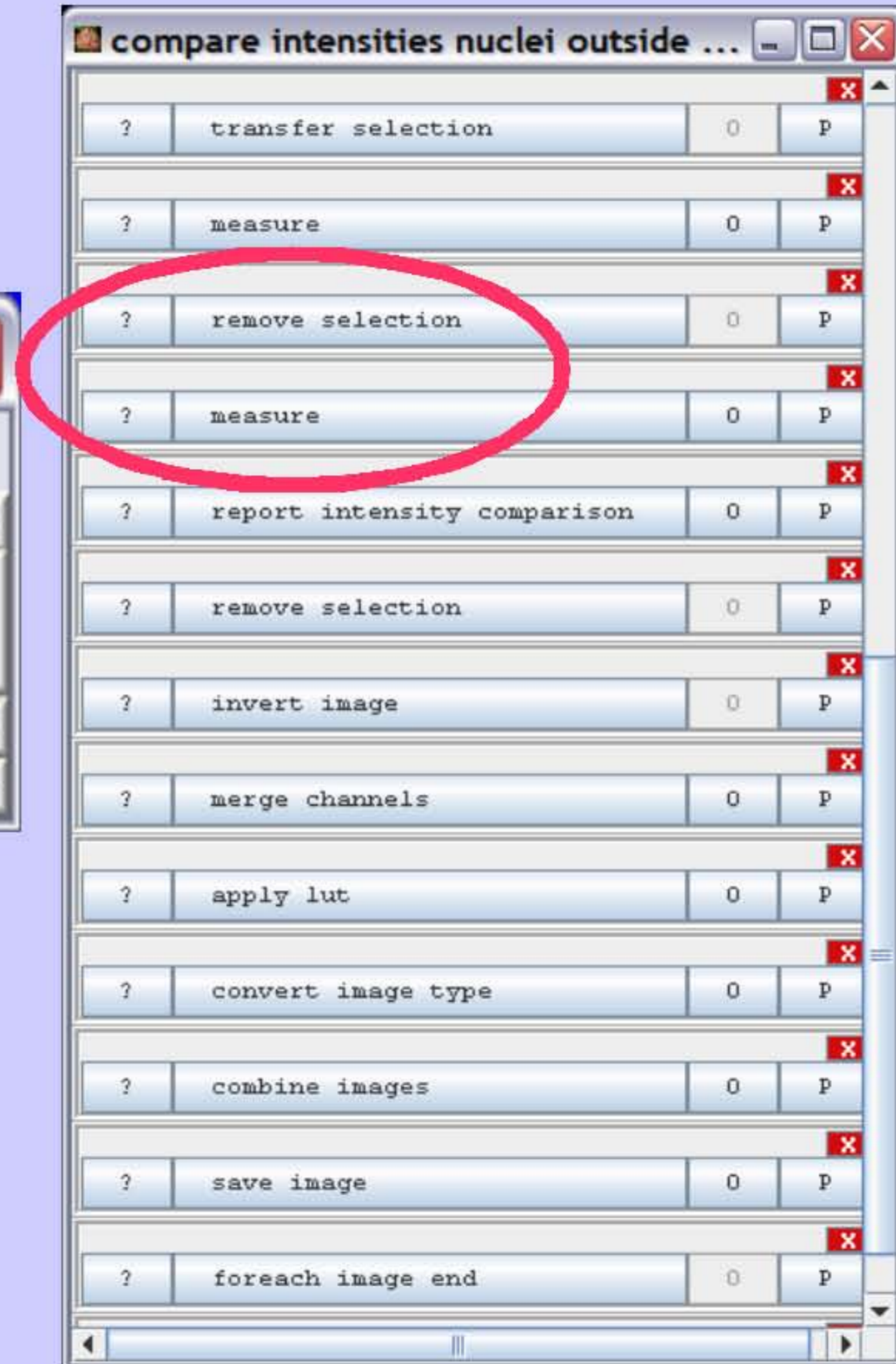




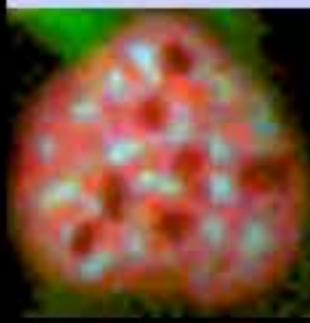
Measuring intensity ratios



	Area	IntDen
1	84203	9347903
2	334750	18258131



remove selection
measure area and intensity of the
whole image





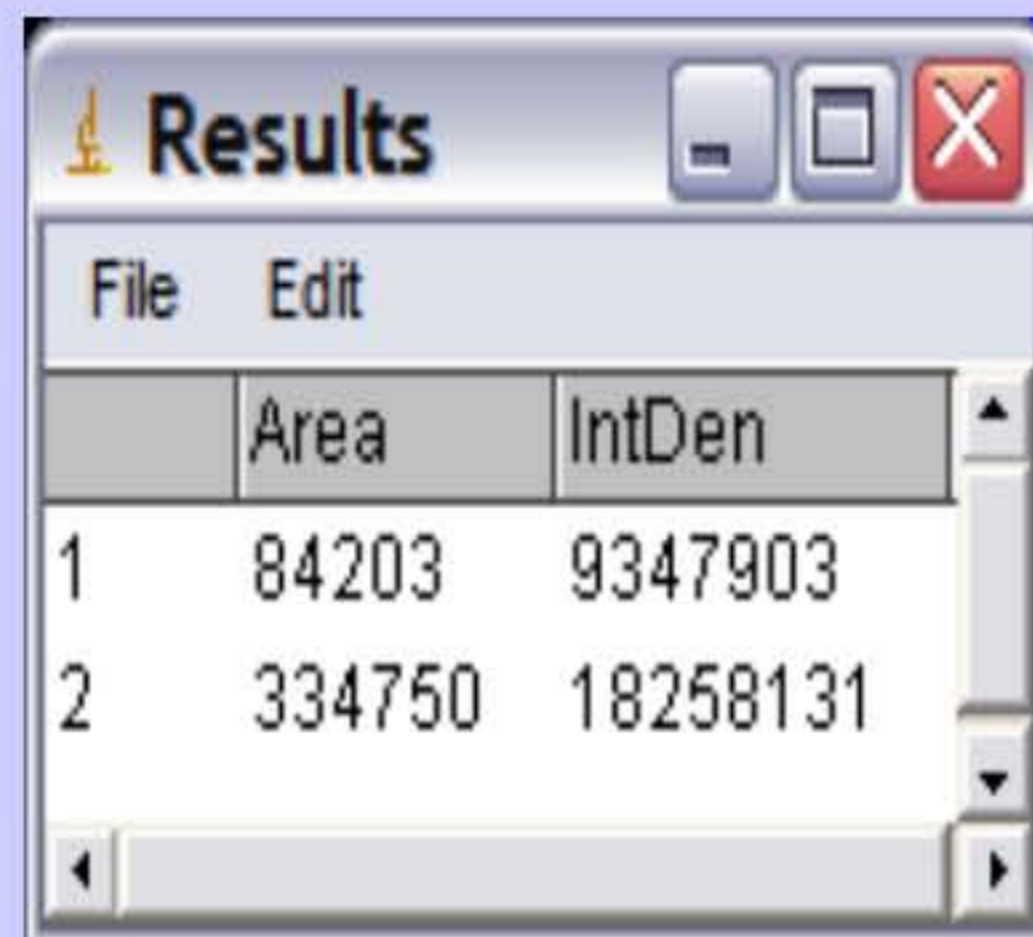
Measuring intensity ratios

calculate percentage of intensity

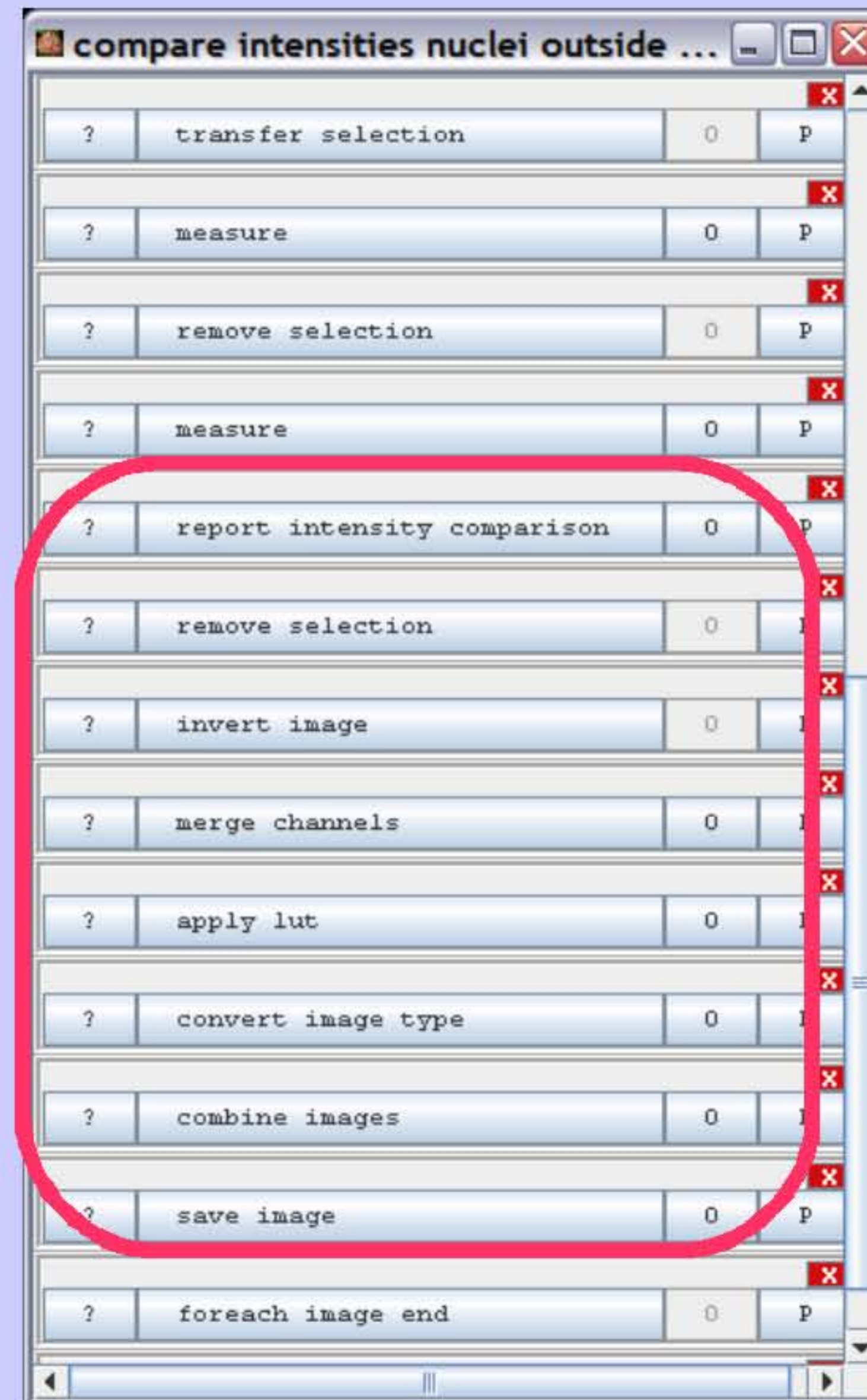
- in nuclei
- outside nuclei

$$p1 = d1 / d2$$

$$p2 = (d2 - d1) / d2$$



	Area	IntDen
1	84203	9347903
2	334750	18258131

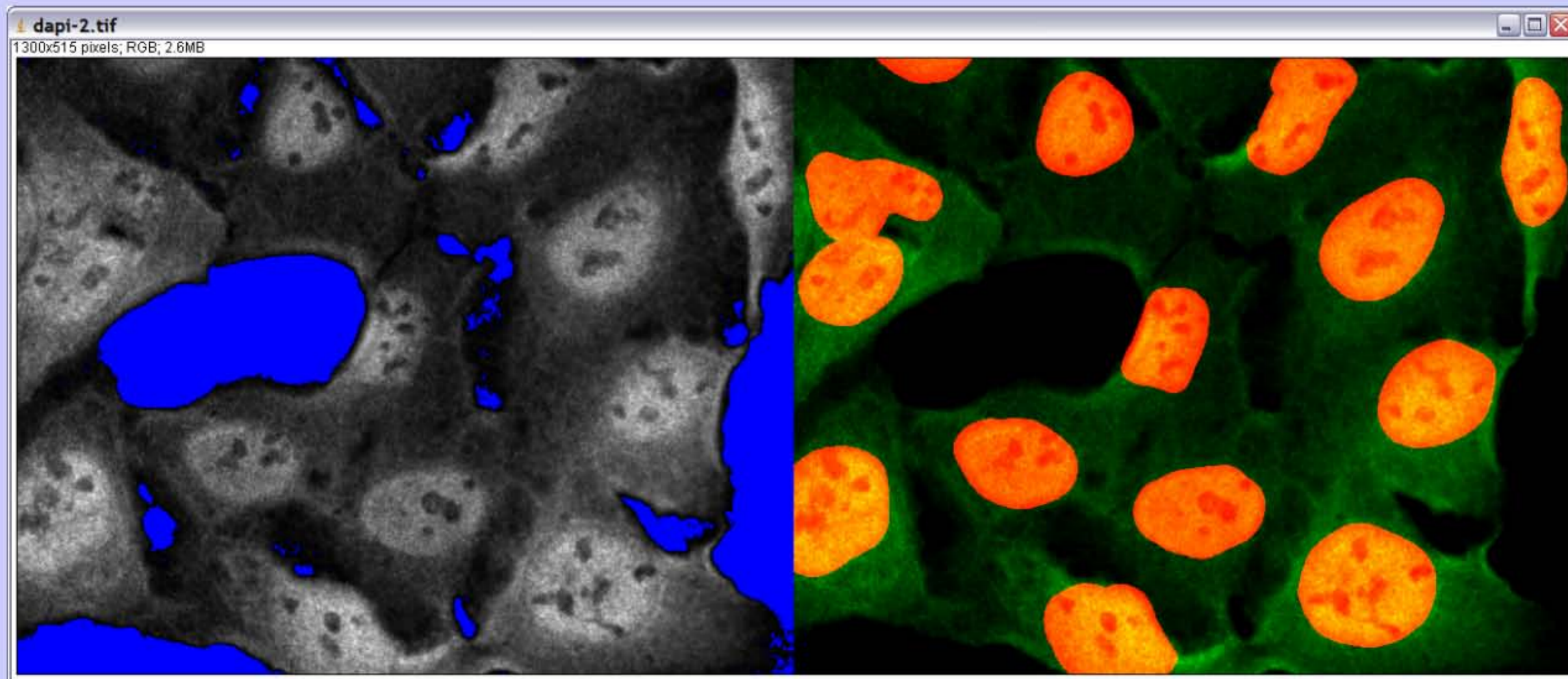




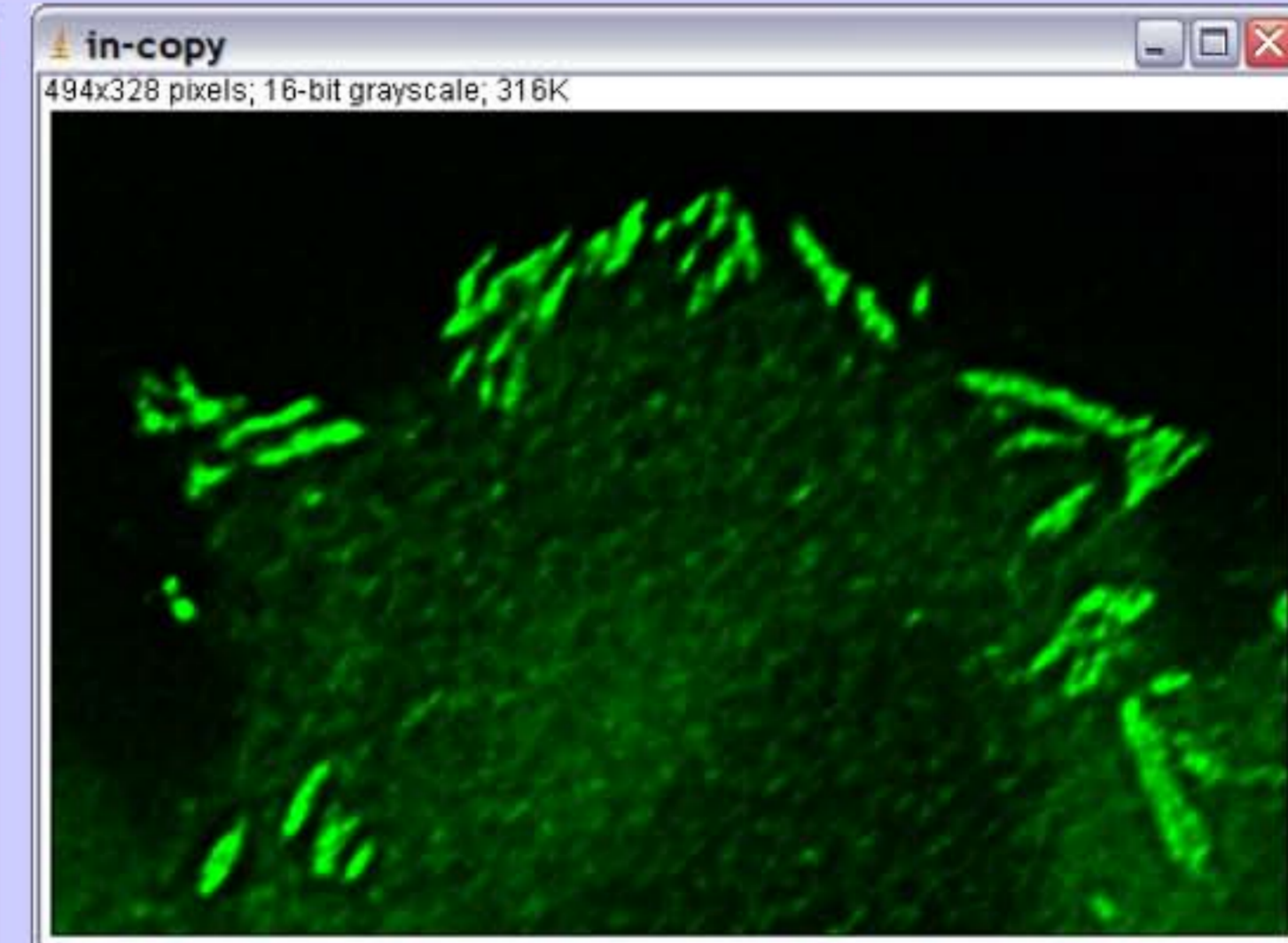
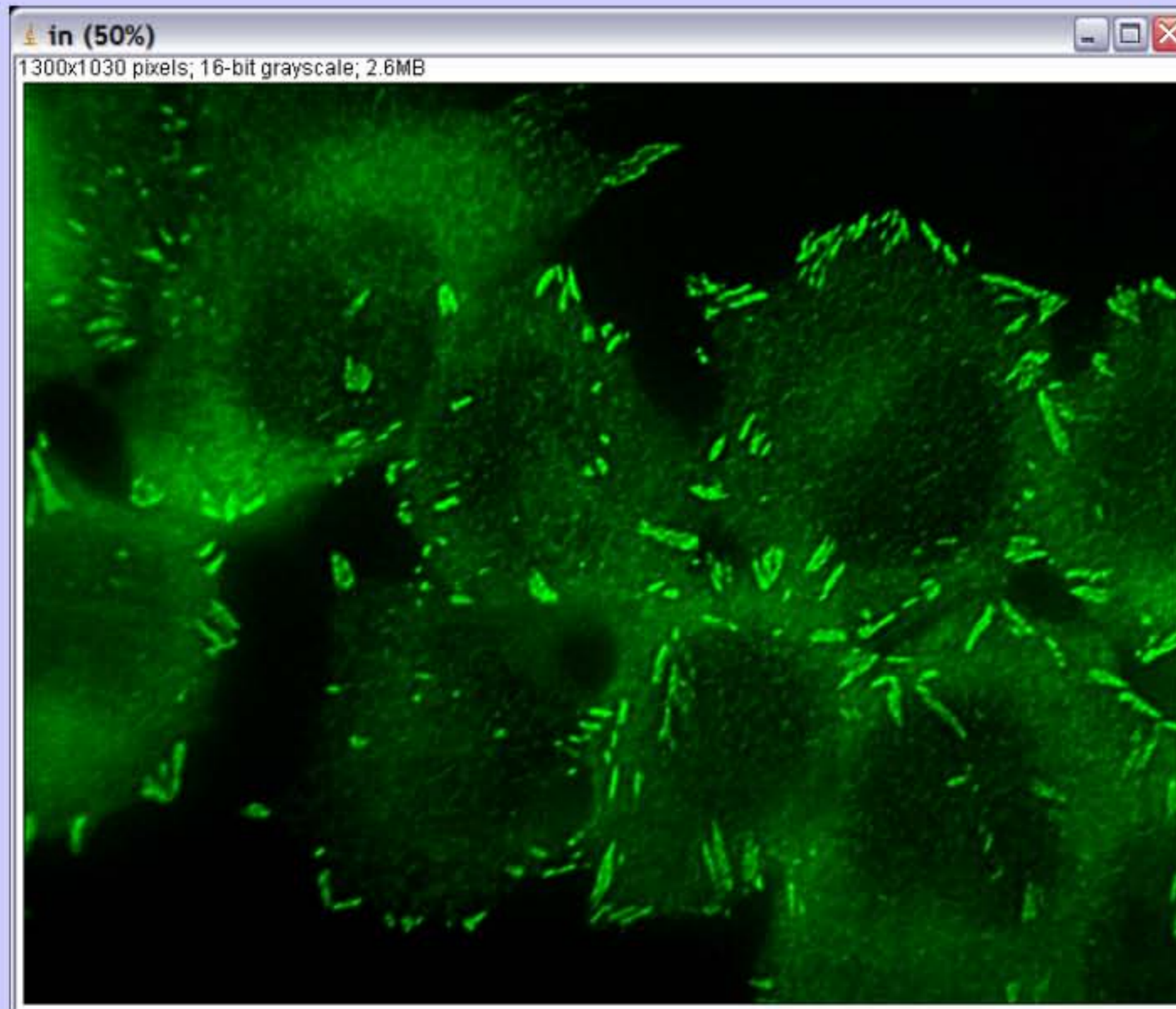
Measuring intensity ratios

image	icn factor	percent nuclei	percent cytoplasm	av.
dapi-2.tif	1,05	0,51	0,49	

report to spreadsheet-file
save control images

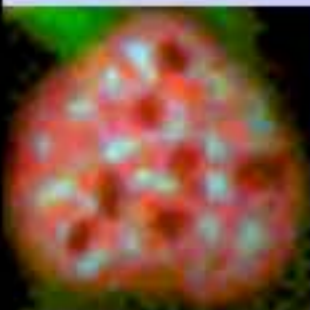


Measuring stained proteins

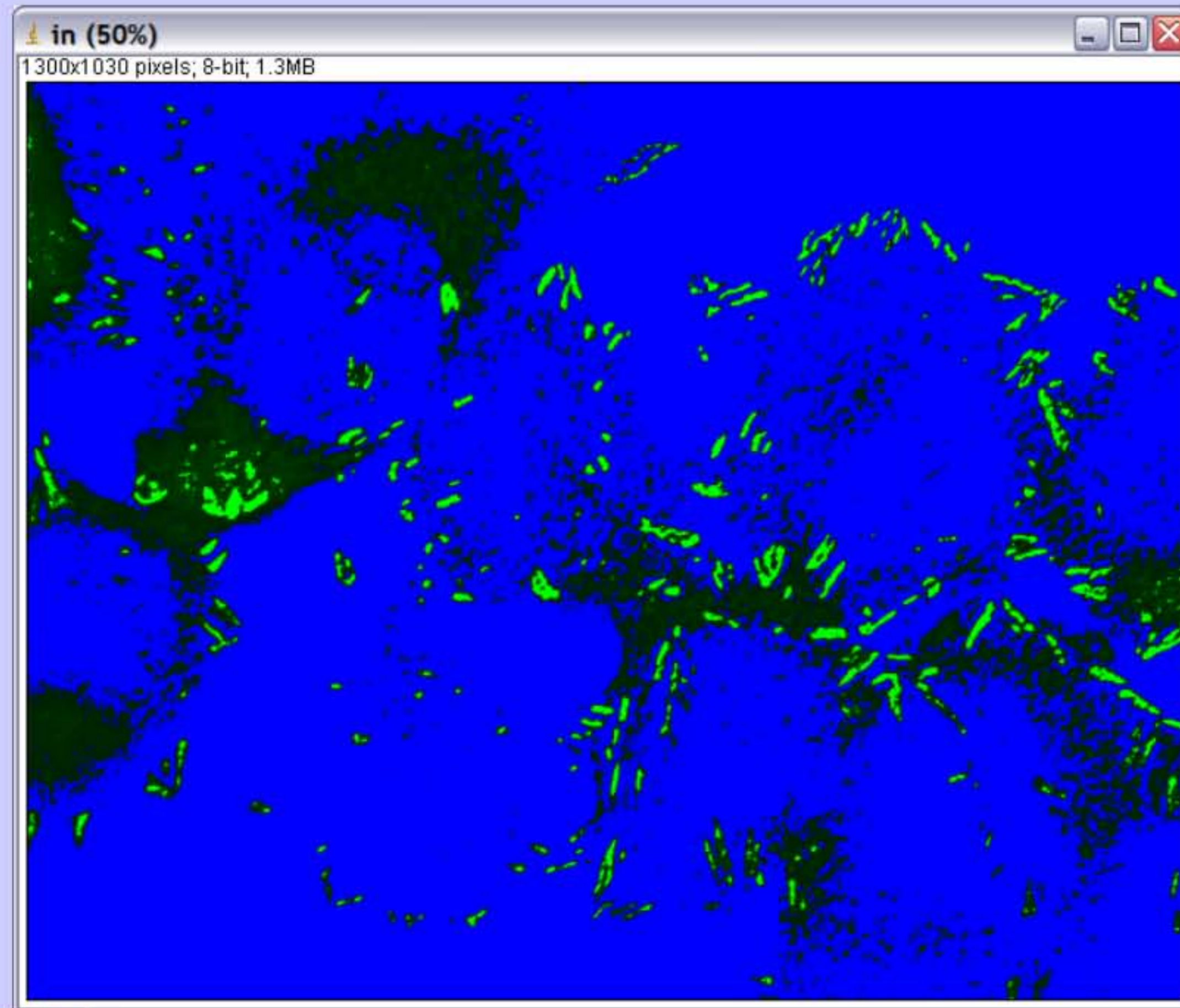


task:

- count plaques
- measure size of plaques



Measuring stained proteins



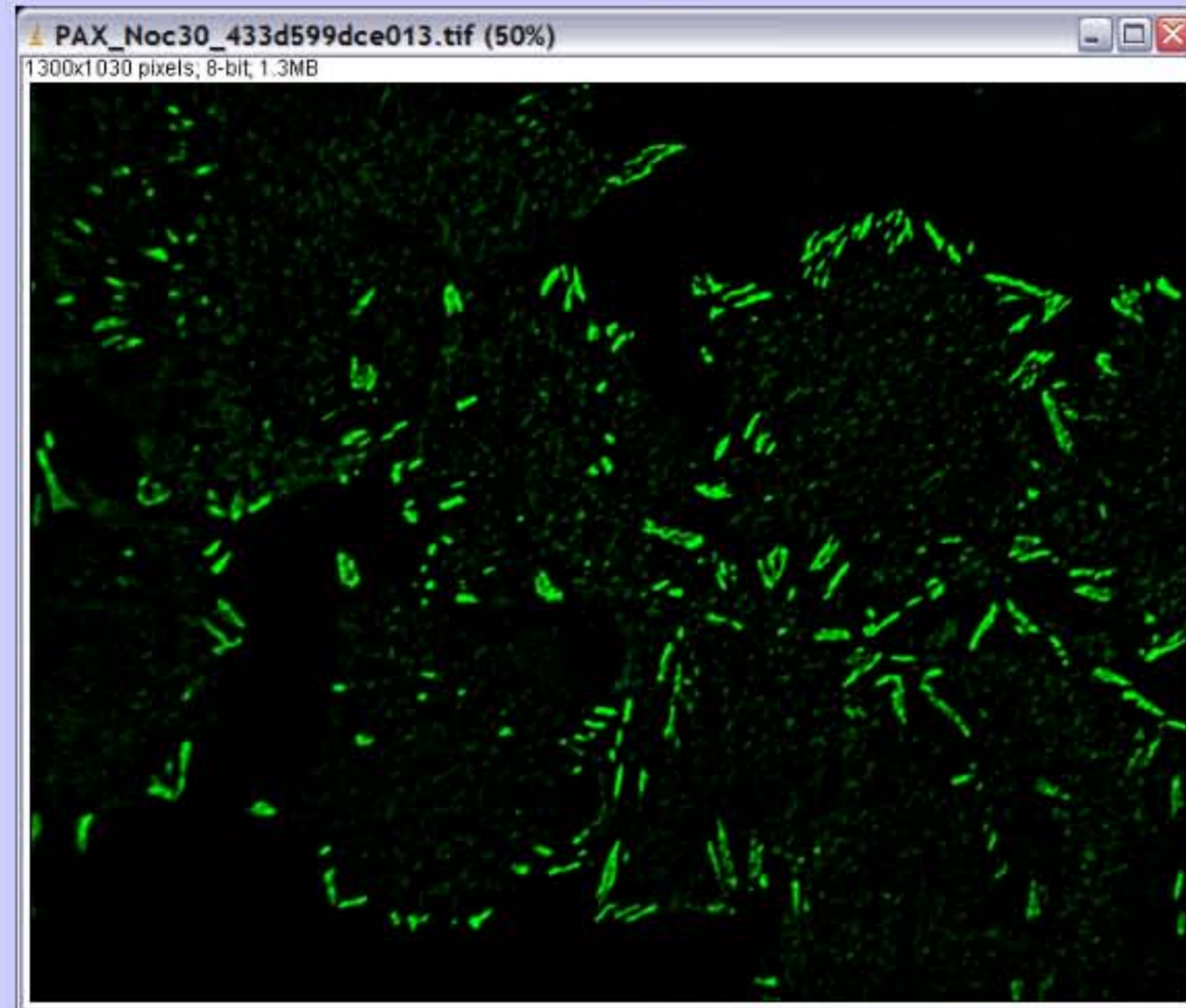
problem:

- same intensities in plaques and cytoplasm
- simple threshold doesn't separate plaques from background

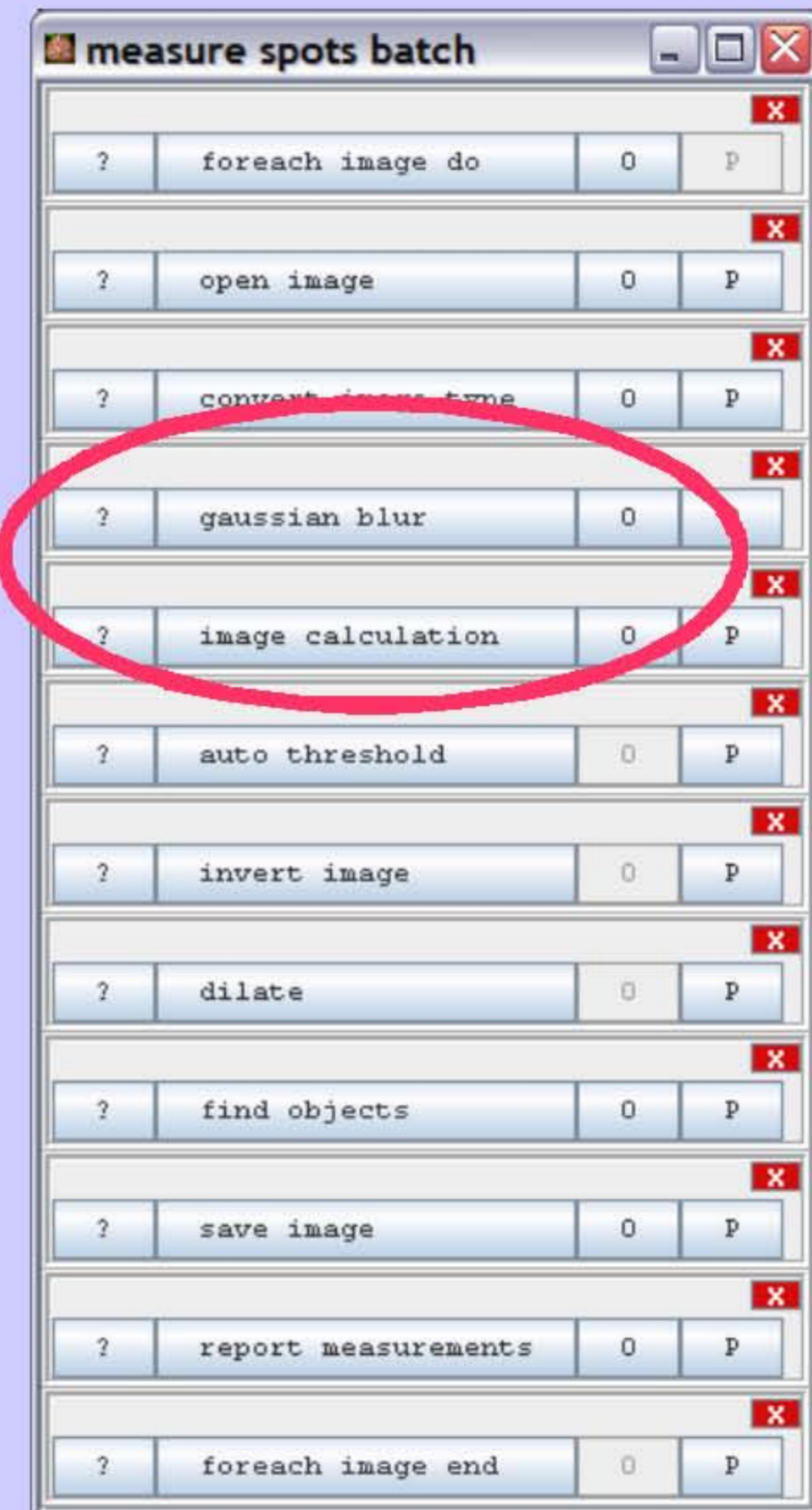




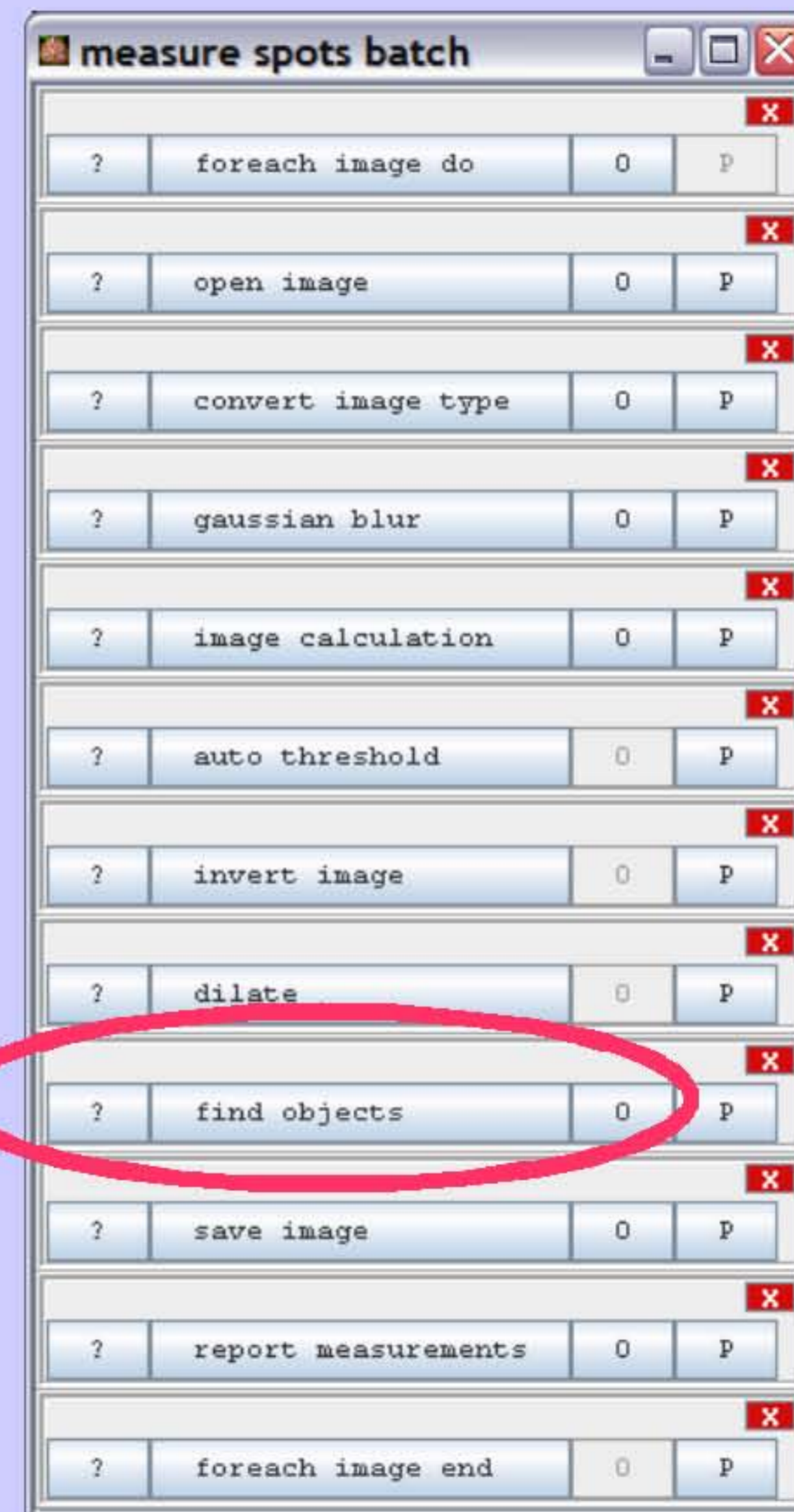
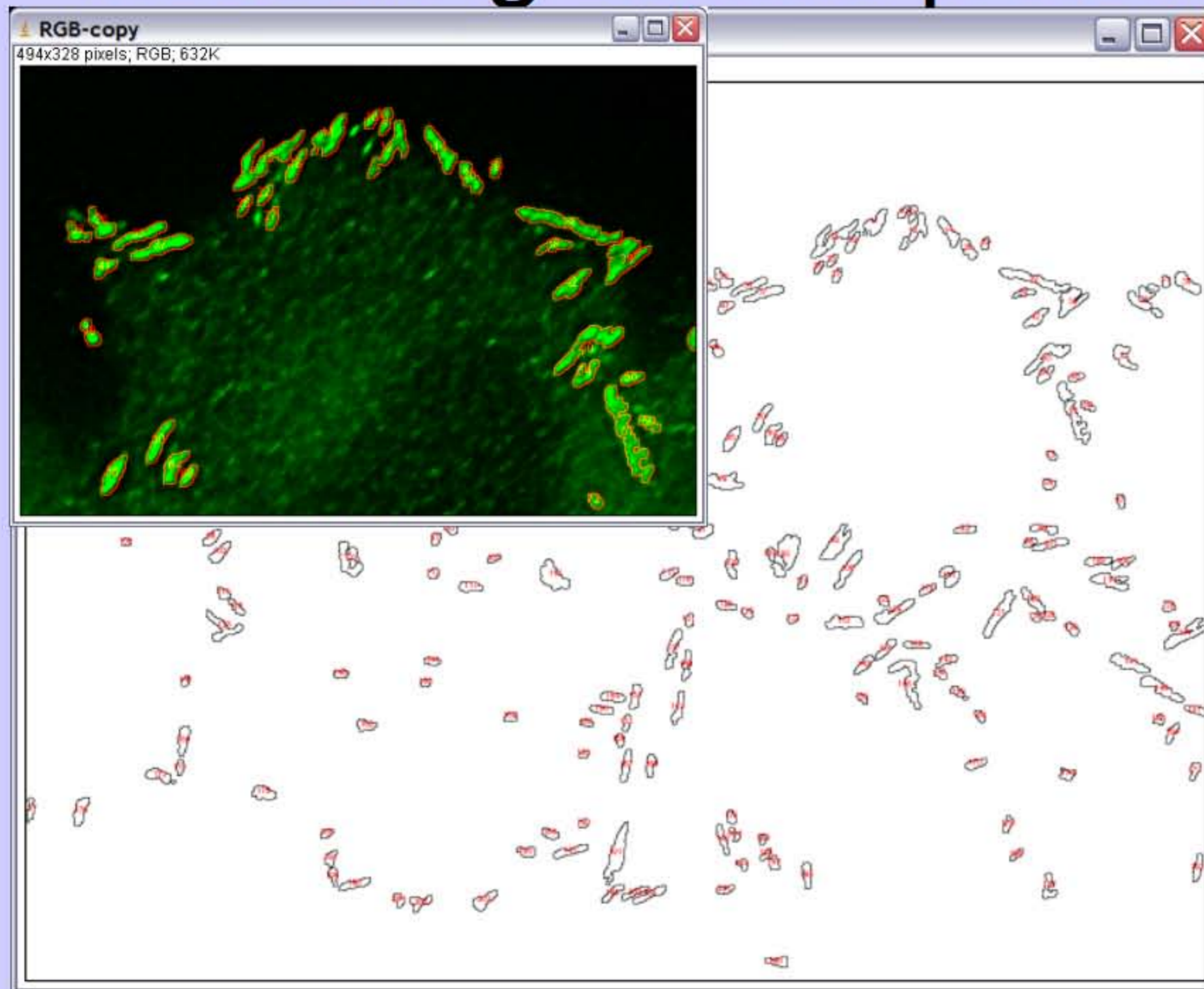
Measuring stained proteins



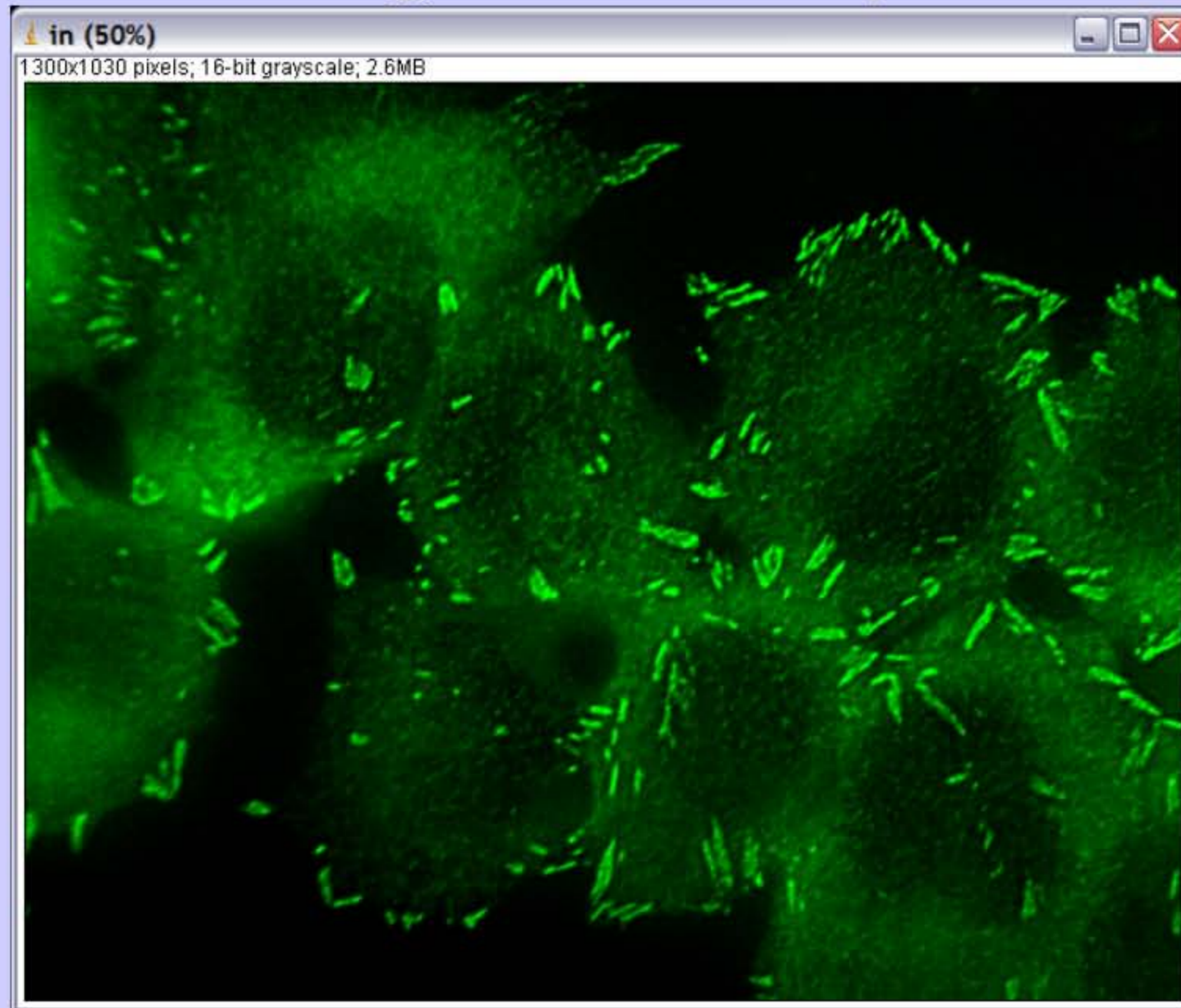
- subtract a blurred version of the image from the original
- only small, isolated dots of the background remain



Measuring stained proteins

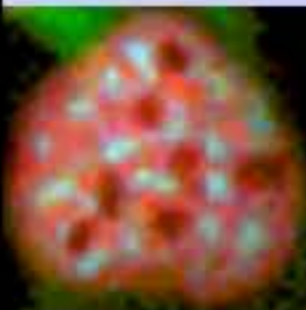


Measuring stained proteins



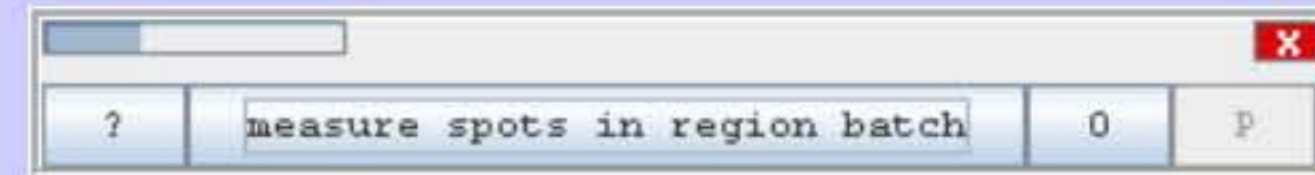
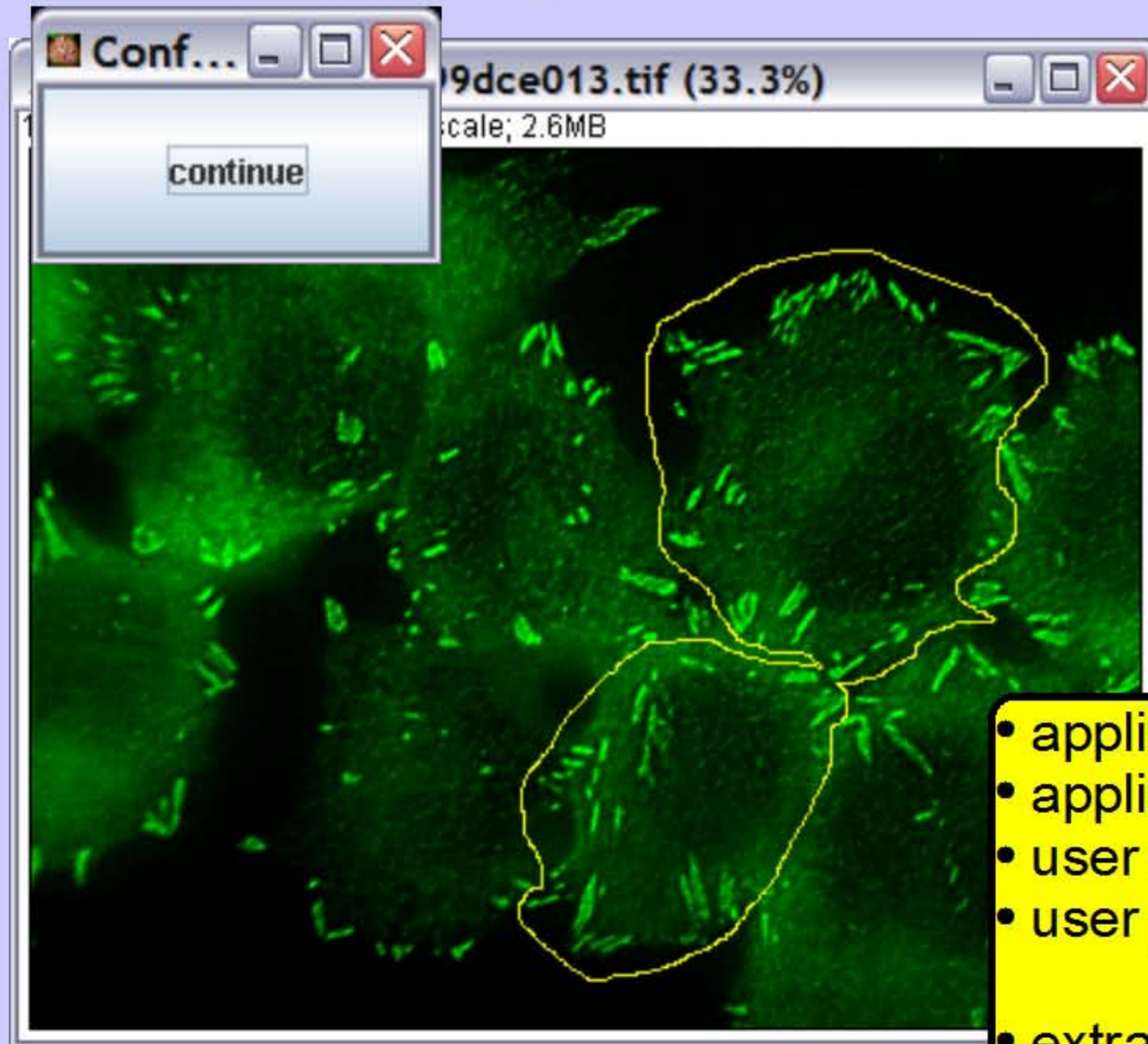
new requirement:

- count and measure plaques only for some selected cells





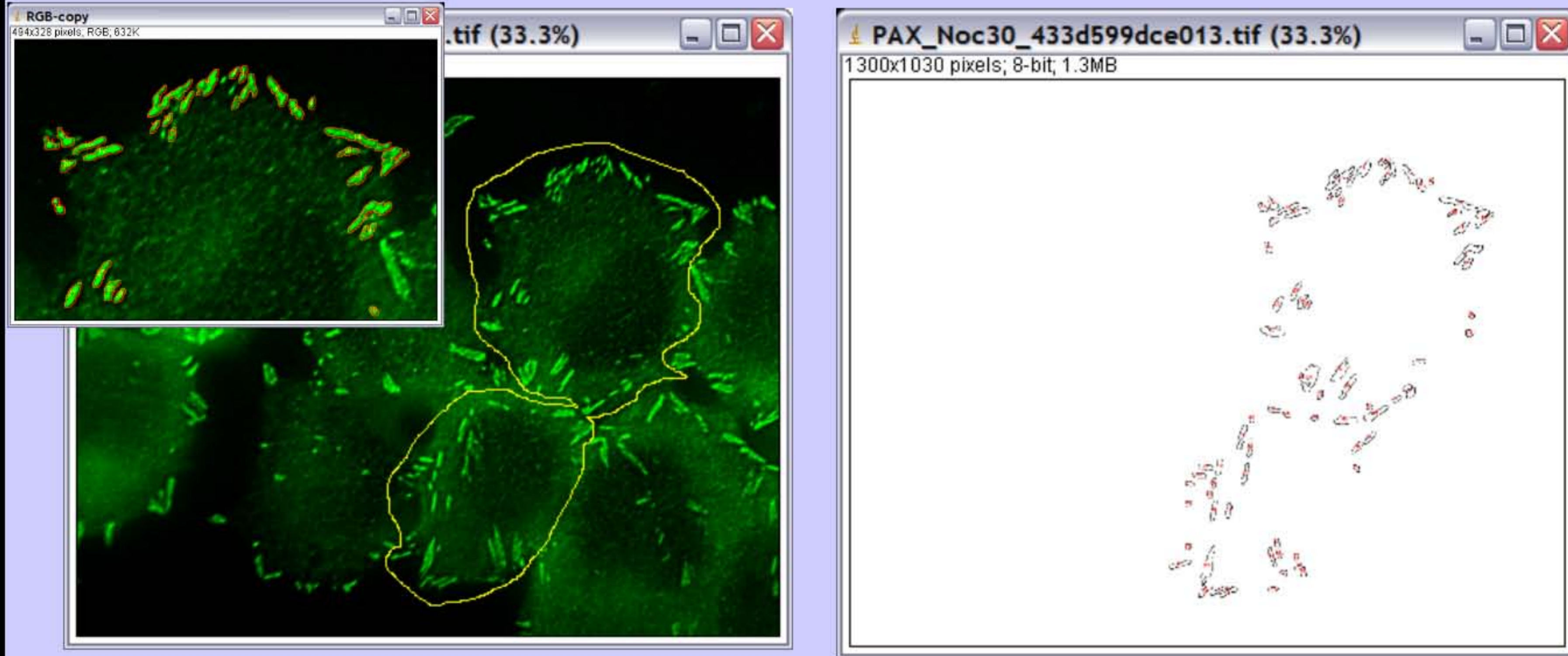
Measuring stained proteins



- application shows image
- application opens « continue » button
- user draws selection
- user presses « continue »
- extra processing to avoid problems at the selection borders



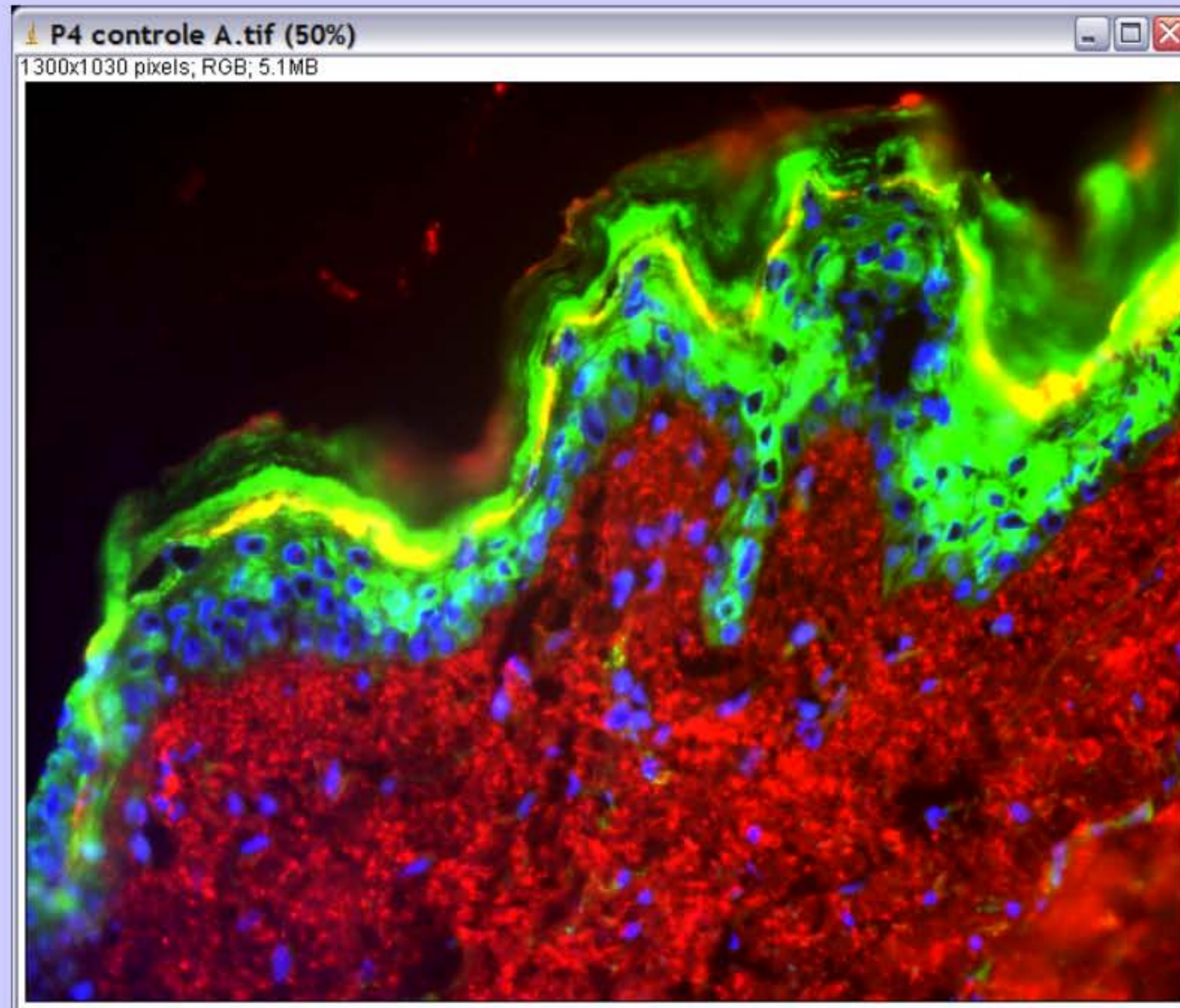
Measuring stained proteins



only the plaques within the selection are taken into account



Counting nuclei

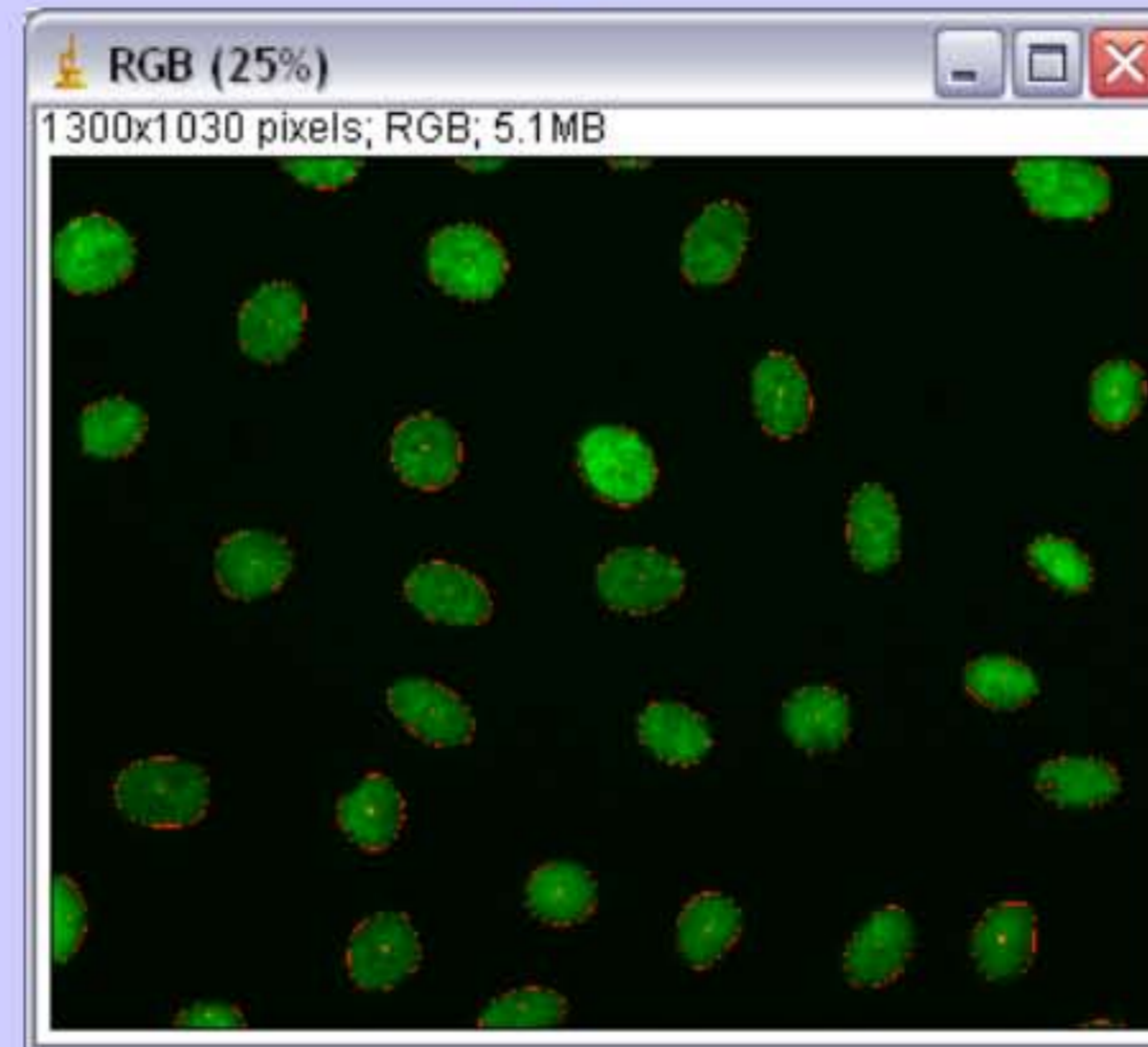
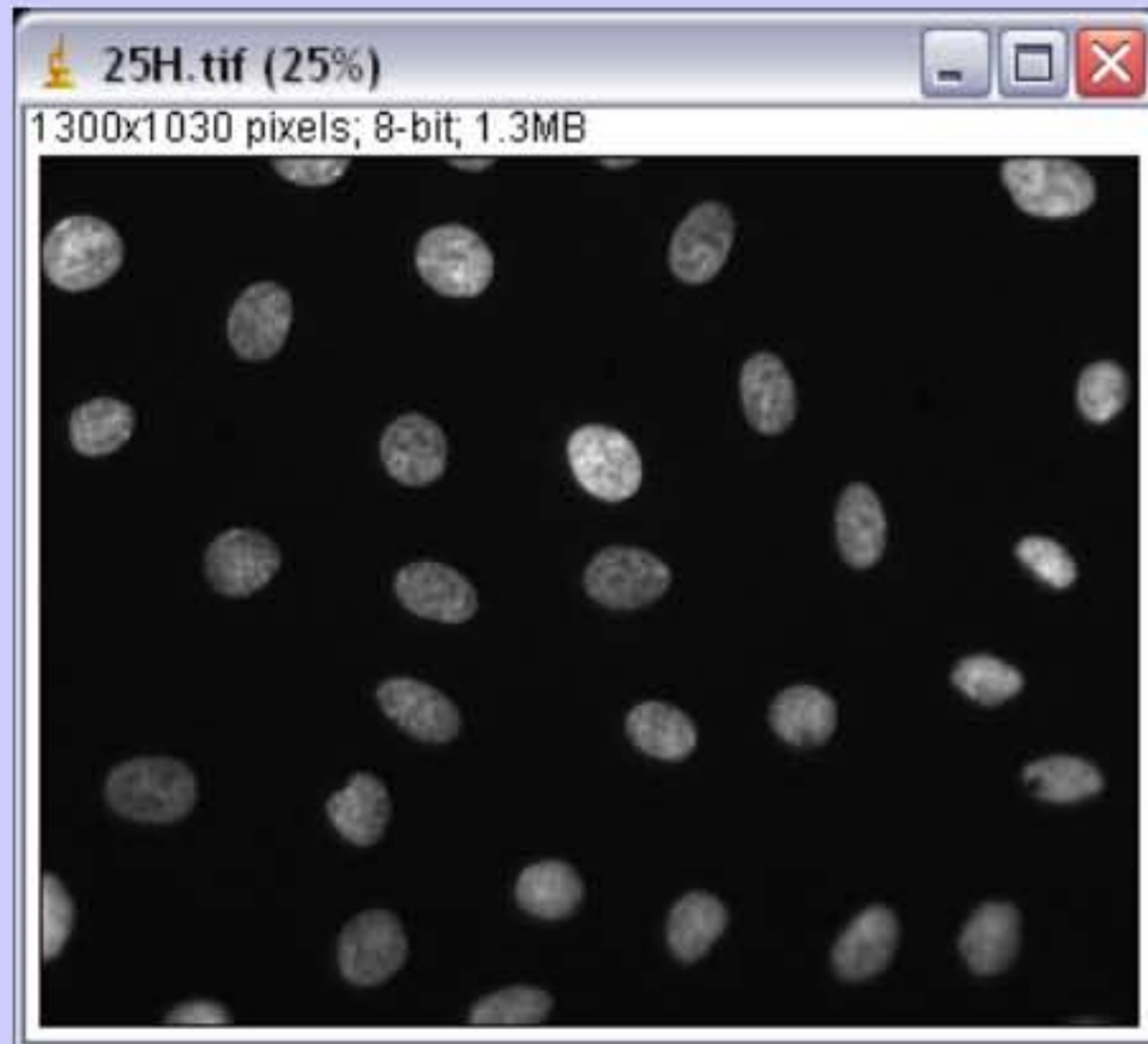


applications

- count cells or nuclei
- count stain combinations
 - blue only,
 - red and blue,
 - ...



Counting nuclei

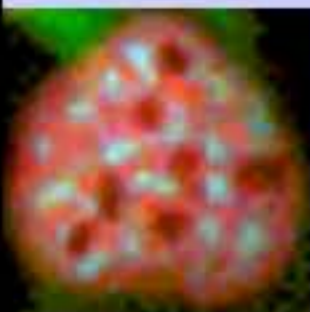


How many cells
are there?

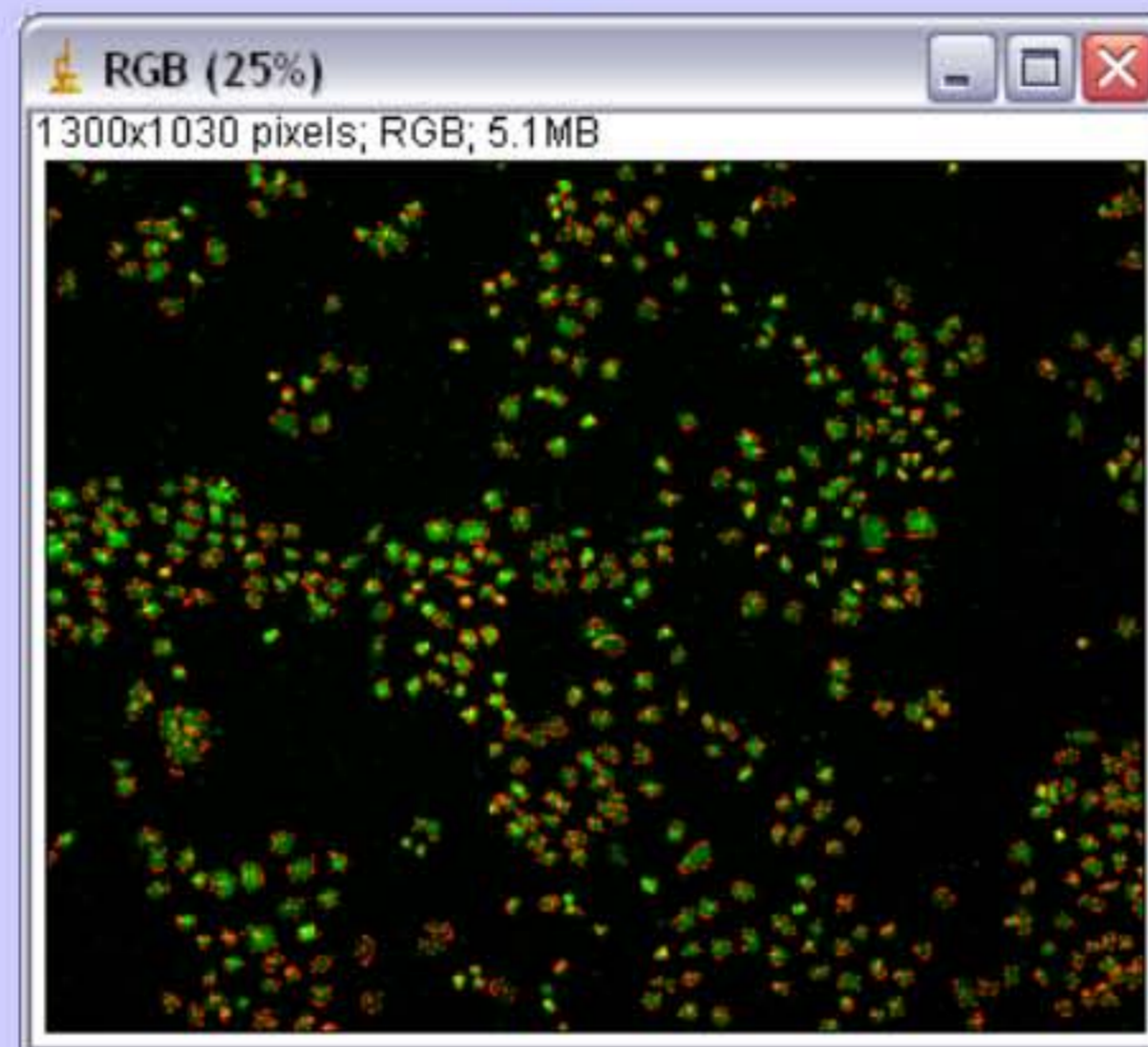
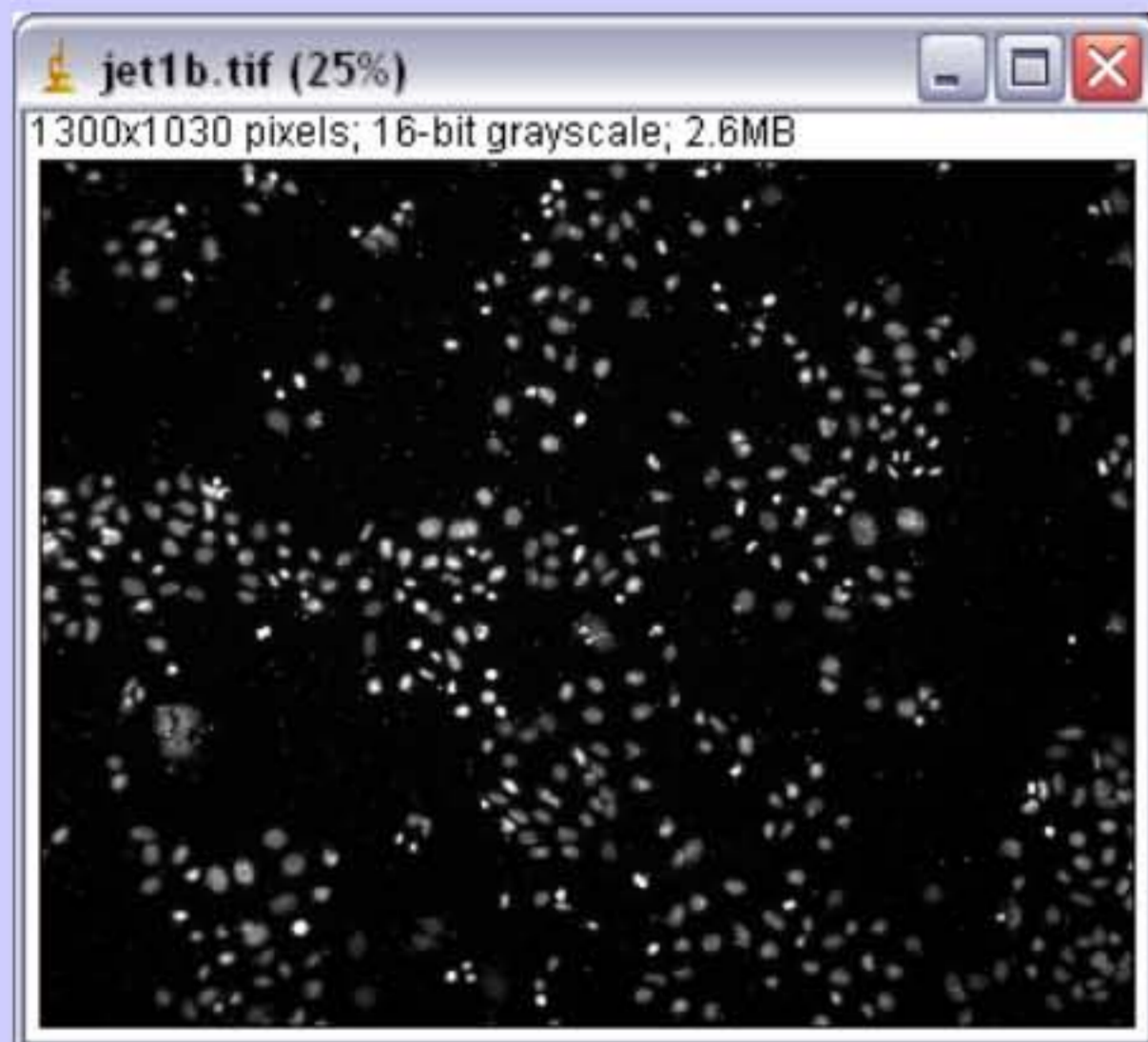
image	number of objects	folder
25H.tif	34	E:\besnard\
jet1b.tif	495	E:\etienne delepine\

- watershed
- derivative
- local threshold

- full automatic
- interactive



Counting nuclei



How many cells
are there?

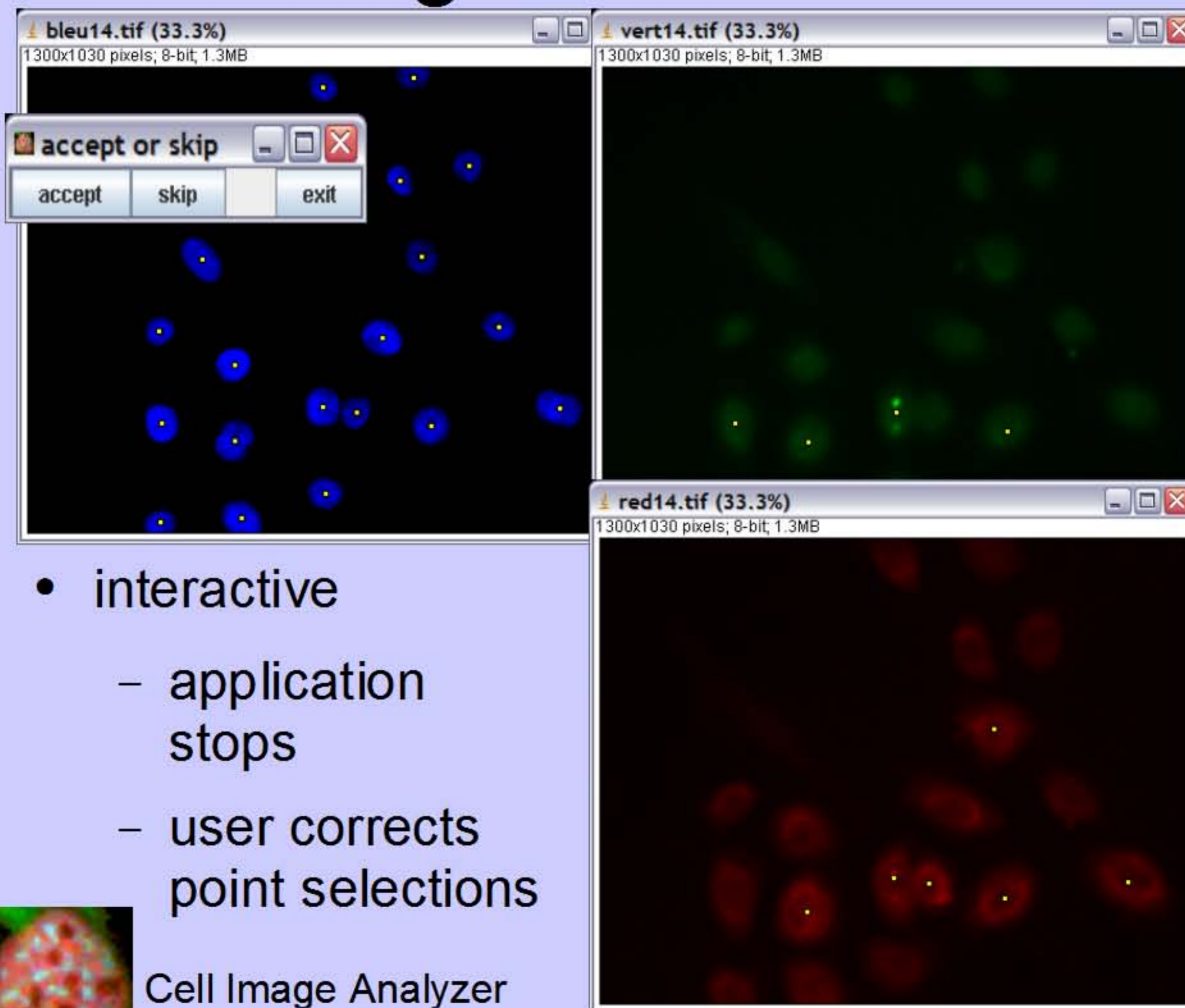
image	number of objects	folder
25H.tif	34	E:\besnard\
jet1b.tif	495	E:\etienne delepine\

- watershed
- derivative
- local threshold

- full automatic
- interactive



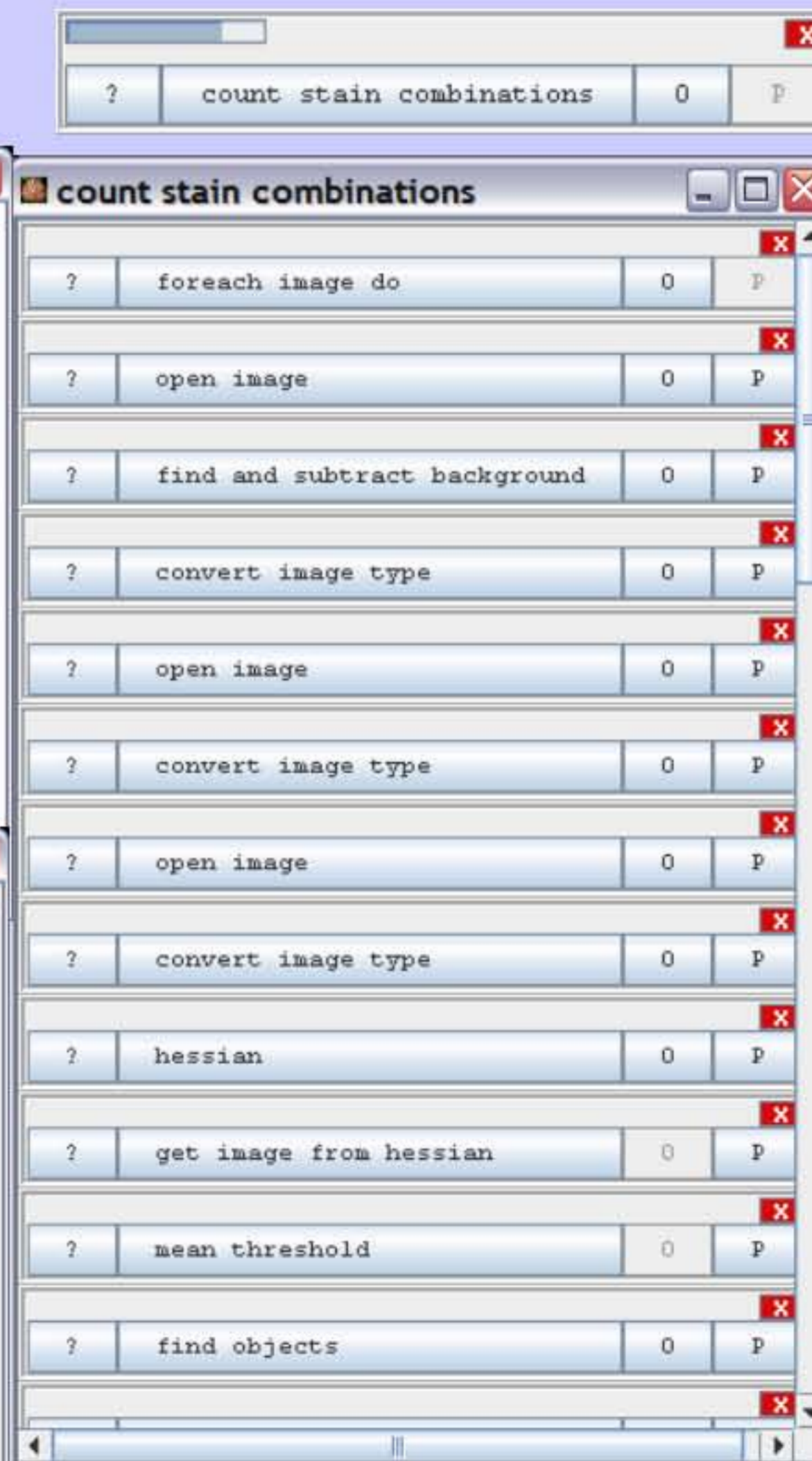
Counting nuclei



- interactive
 - application stops
 - user corrects point selections

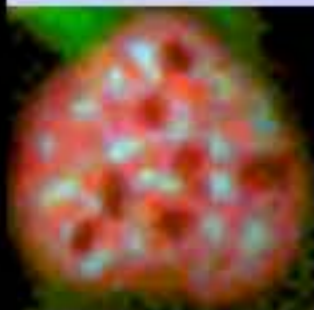
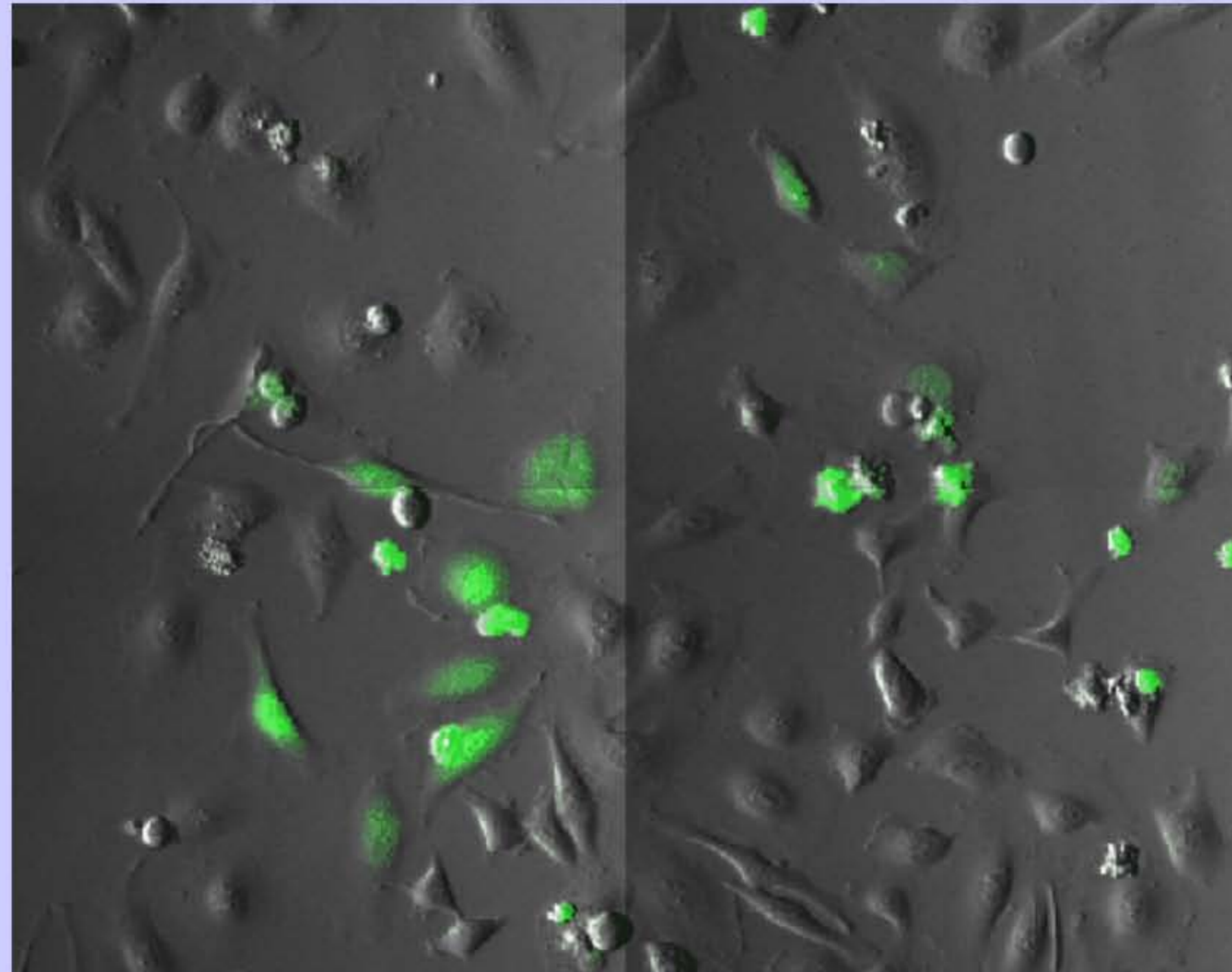


Cell Image Analyzer



Automation of simple tasks

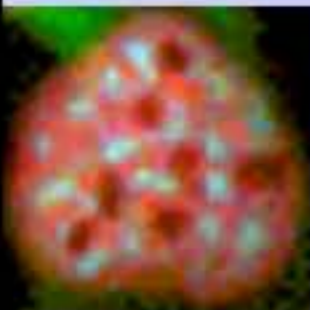
- batch type conversions
- creating compressed movies from large data sets
 - use virtual stack opener + QuickTime Stack Writer plugin
- creating overlay of fluorescence and phase contrast image series
- ...



Further projects and projects in work



- measuring growth of plant surface (first version deployed)
- measuring growth of plant roots
- 3d modelling from histological sections
 - mouse
 - palm tree meristem

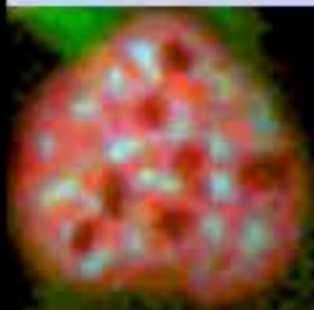


Conclusions

MRI Cell Image Analyzer

ImageJ + visual scripting

- rapidly
 - create image analysis applications
 - automate image analysis tasks
 - batch applications
 - semi-automatic applications
- interface application developer - scientist
 - easy to use by non computer experts
- the framework helps create new operations at the programming level
- all ImageJ methods can be accessed with the help of proxies
- used at Montpellier RIO Imaging
 - to create image analysis applications on demand
 - in close collaboration with the biologists
- made it possible to provide solutions that could not be obtained with standard image analysis packages
- outlook:
 - allow hierarchical applications (the use of applications in applications)
 - allow to work seamlessly with very big data sets (elaborate the virtual stack concept)



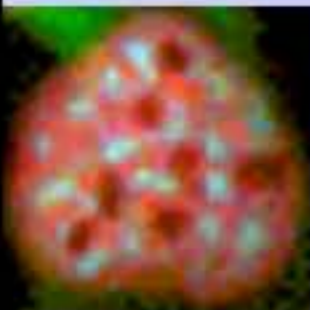
The last slide



- Thanks to
 - Wayne Rasband
 - all ImageJ plugin and macro authors
 - the ImageJ community
 - The ImageJ conference - organisation
 - Montpellier RIO Imaging
 - the research groups that participated in developing image analysis applications

Thank you, for your attention!

? Questions ?



References

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